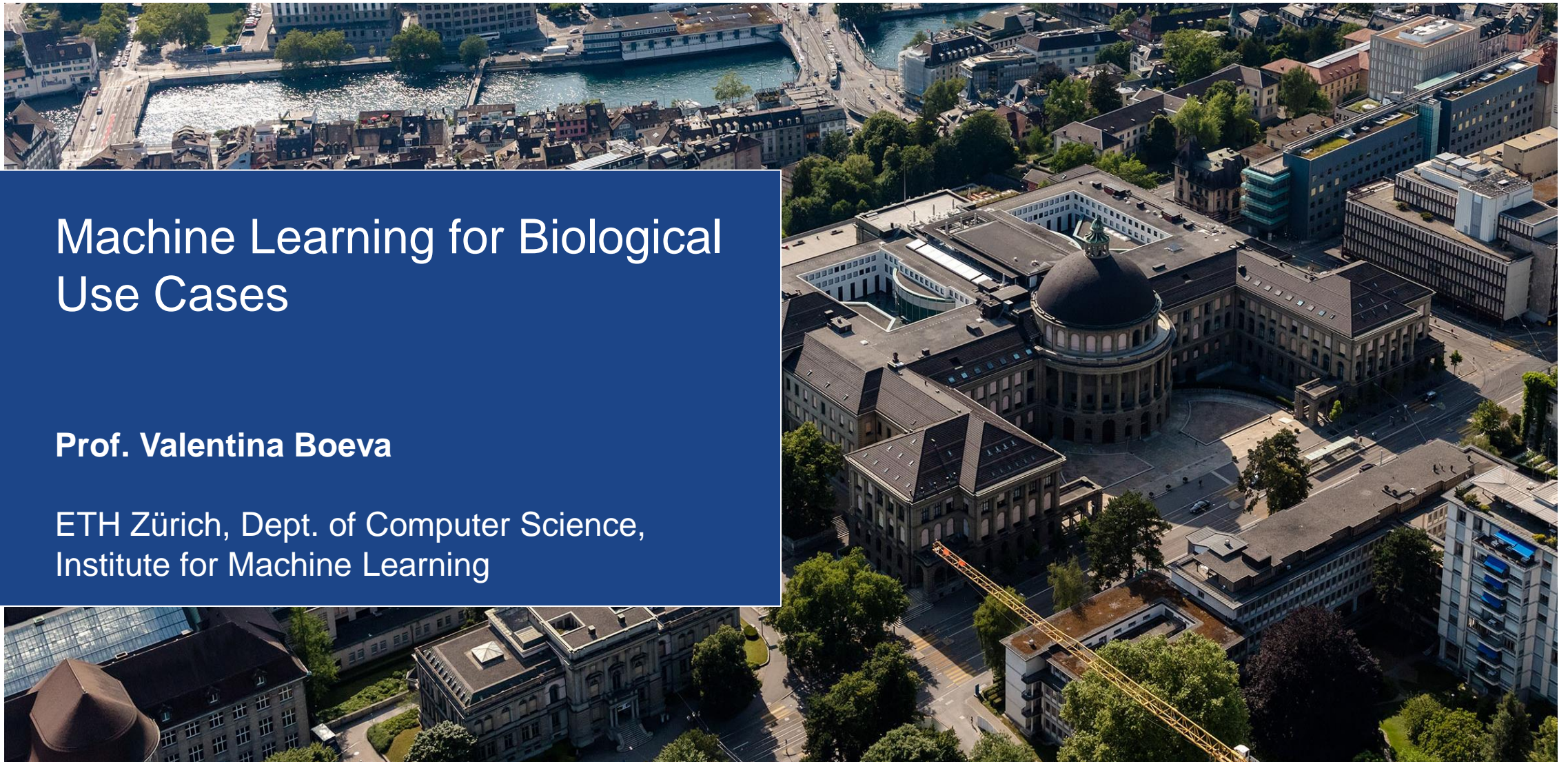


# Machine Learning for Biological Use Cases

**Prof. Valentina Boeva**

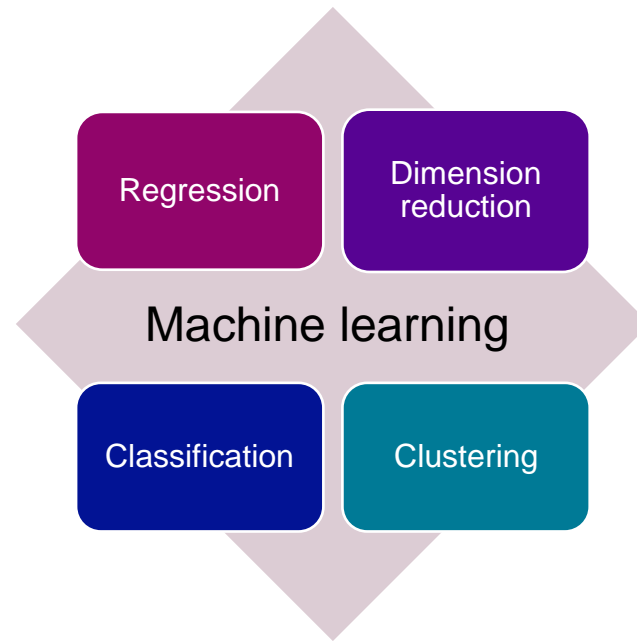
ETH Zürich, Dept. of Computer Science,  
Institute for Machine Learning



# Machine learning



# Map of classical machine learning methods



# Map of classical machine learning methods

**Supervised**

learns on data **with labels**

**Unsupervised**

learns on data **without labels**

**Continuous**

**Discrete/Categorical**

Regression

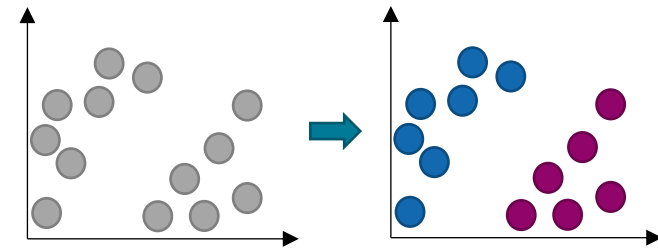
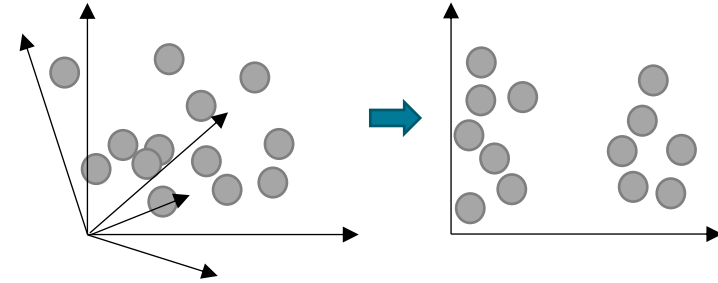
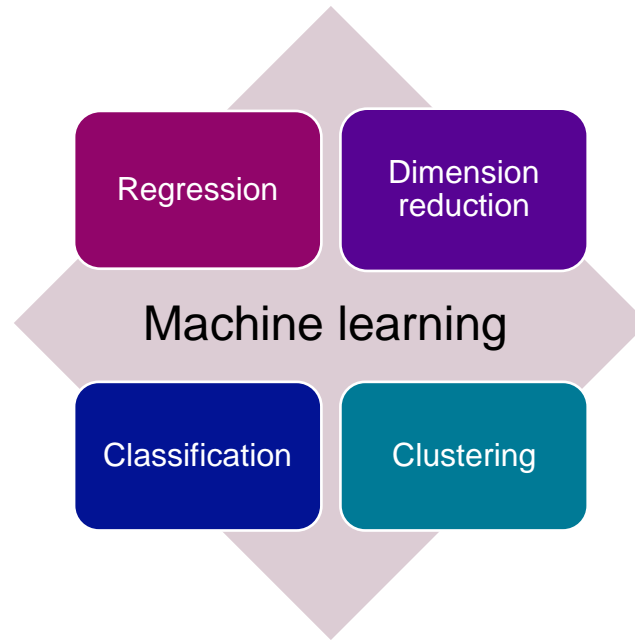
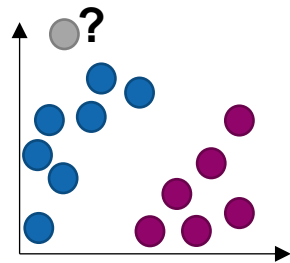
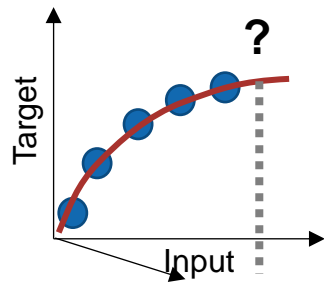
Dimension  
reduction

Machine learning

Classification

Clustering

# Map of classical machine learning methods



# Map of classical machine learning methods

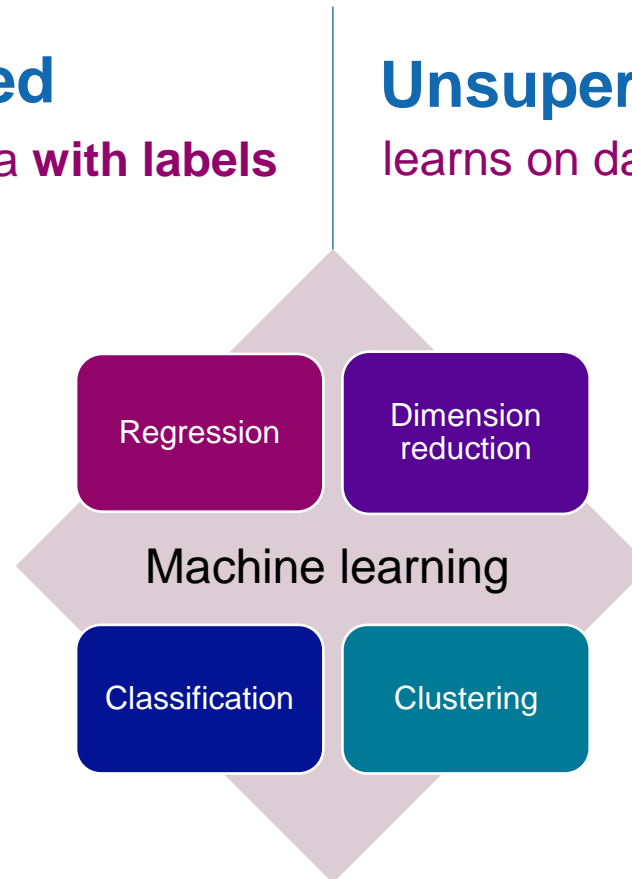
## Supervised

learns on data **with labels**

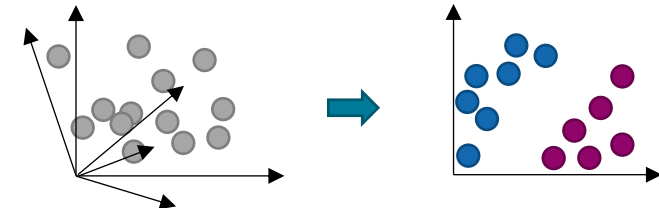
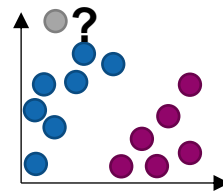
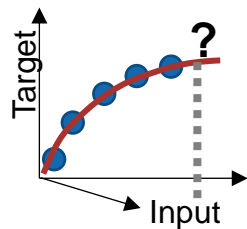
## Unsupervised

learns on data **without labels**

- Automate decisions
  - Primary diagnosis
  - Choice of treatment
- Predict future
  - Treatment response
  - Side effects
- Detect the most “important” features



- Understand data structure
- Visualize data in 2D/3D
- Detect hidden features



# Types of input data in molecular biology and genetics

## Type of data:

- **Genomic:**
  - Variants common in general population (SNPs)
  - Rare variants
    - Single nucleotide variants, SNV (a.k.a. mutations)
    - Structural aberrations (e.g., translocations, amplifications)
    - Copy number profiles
- **Transcriptomic:**
  - RNA-seq (or expression microarrays) – bulk and single cell
  - mi-RNA
- **Epigenetic:**
  - DNA methylation (sequencing or methylation arrays)
  - Histone modifications (ChIP-seq) and open chromatin (ATAC-seq, DHS-seq)
- **Proteomic:**
  - RPPA (bulk), CyTOF (single cell)

## Context:

- Common diseases (Alzheimer, asthma, hypertension,...)
- Genetic syndromes (Down syndrome, CHARGE syndrome, ...)
- Cancer

## Type of samples:

- Blood samples
- Saliva samples
- Tissue samples
- Maternal blood samples

# Omics data are high dimensional

SNPs	Mutations (SNVs)	Structural variants	Copy number alterations	mRNA expression	miRNA expression	DNA methylation
3M-100M	~10-10K	~1-1000	~10-25K	~25K	~1000	27K-28M

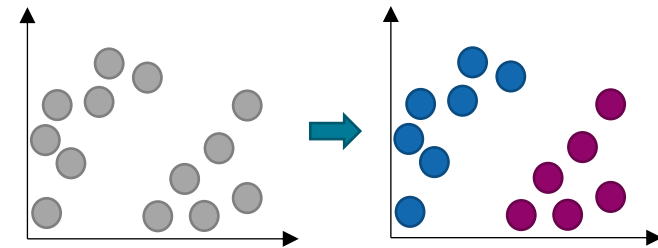
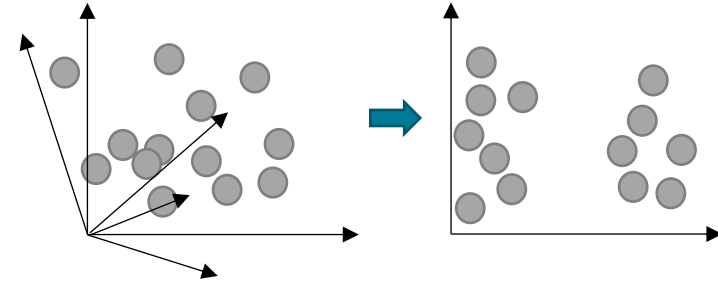
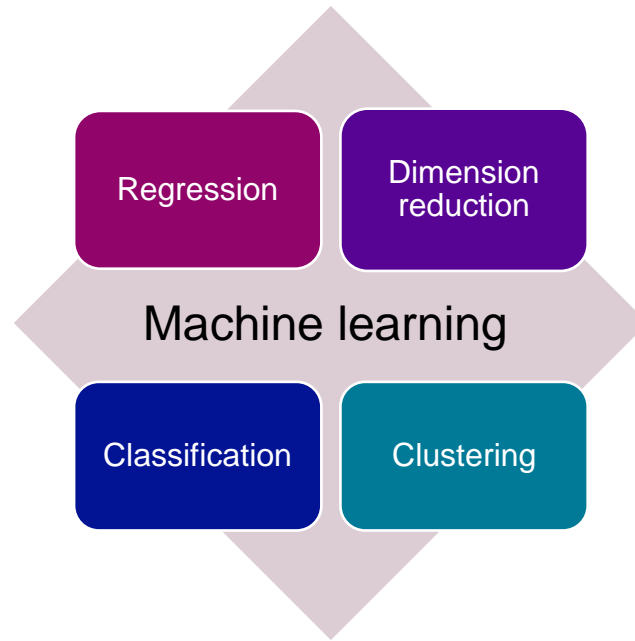
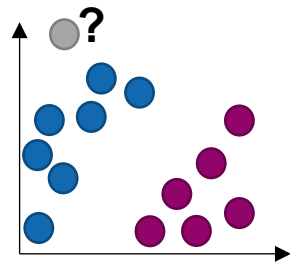
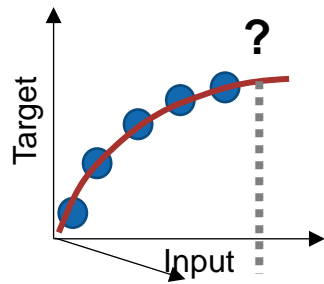
+ sometimes these data is complemented with proteomics data (expression of hundreds of proteins)

Full –omics dataset millions of observations per patient:

**Great challenge to avoid over-fitting and perform feature selection!**



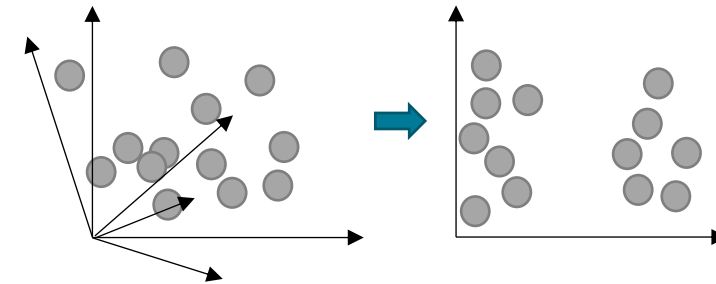
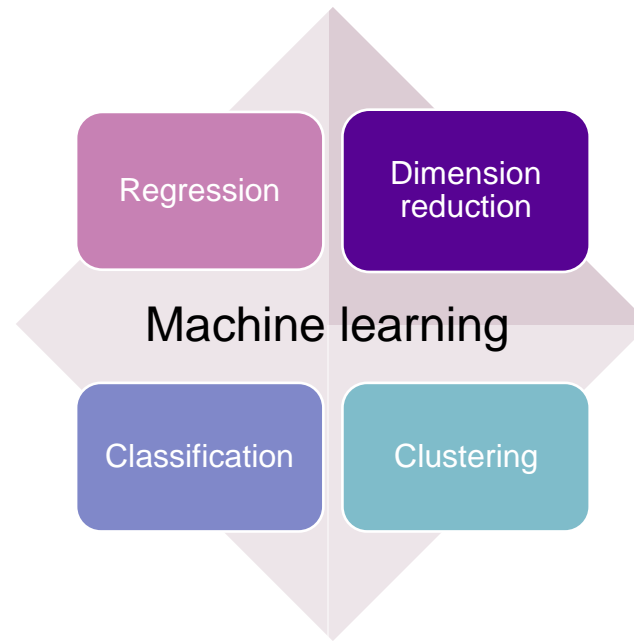
# Map of classical machine learning methods



# Map of machine learning methods

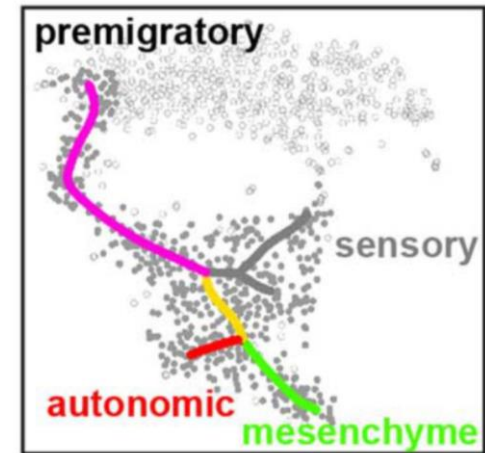
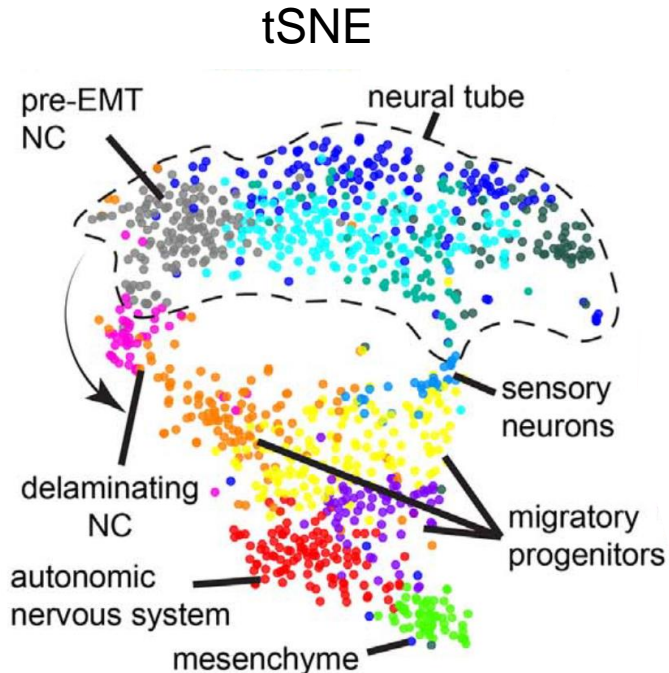
## Unsupervised

learns on data **without labels**



# Examples: Dimension reduction and clustering

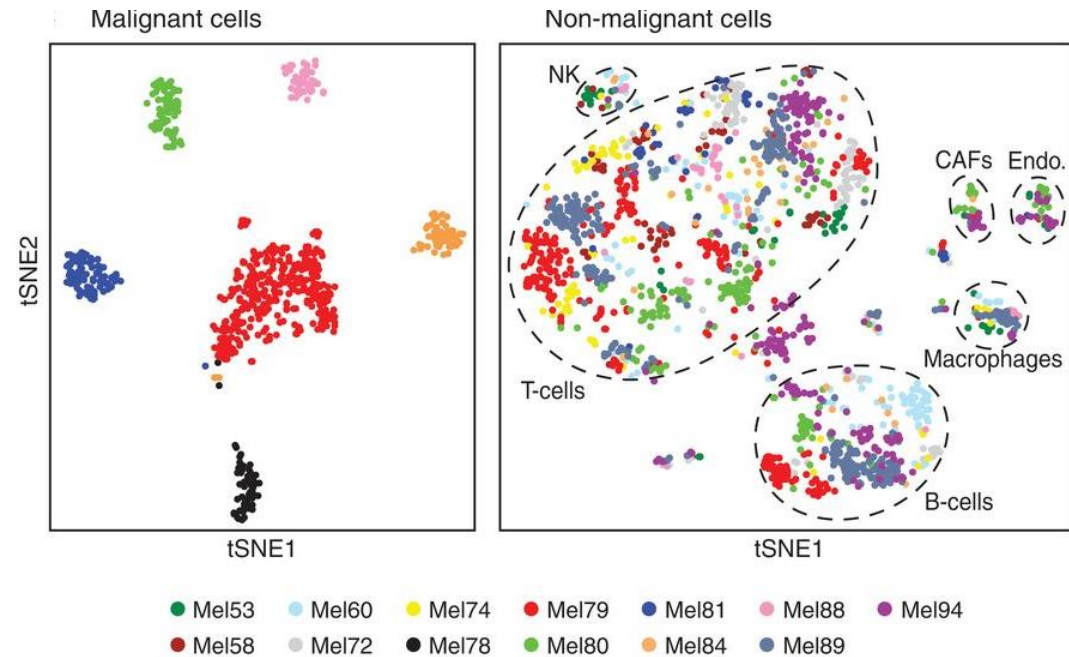
- Single cell transcriptomics: development, cell type heterogeneity and cancer
- Bulk transcriptomics and epigenetics (cancer)
- Bulk transcriptomics: Effect of mutations



**Spatiotemporal structure of cell fate decisions in murine neural crest.** Soldatov et al., *Science* 364, 971 (2019)

# Examples: Dimension reduction and clustering

- Single cell transcriptomics: development, cell type heterogeneity and cancer
- Bulk transcriptomics and epigenetics (cancer)
- Bulk transcriptomics: Effect of mutations



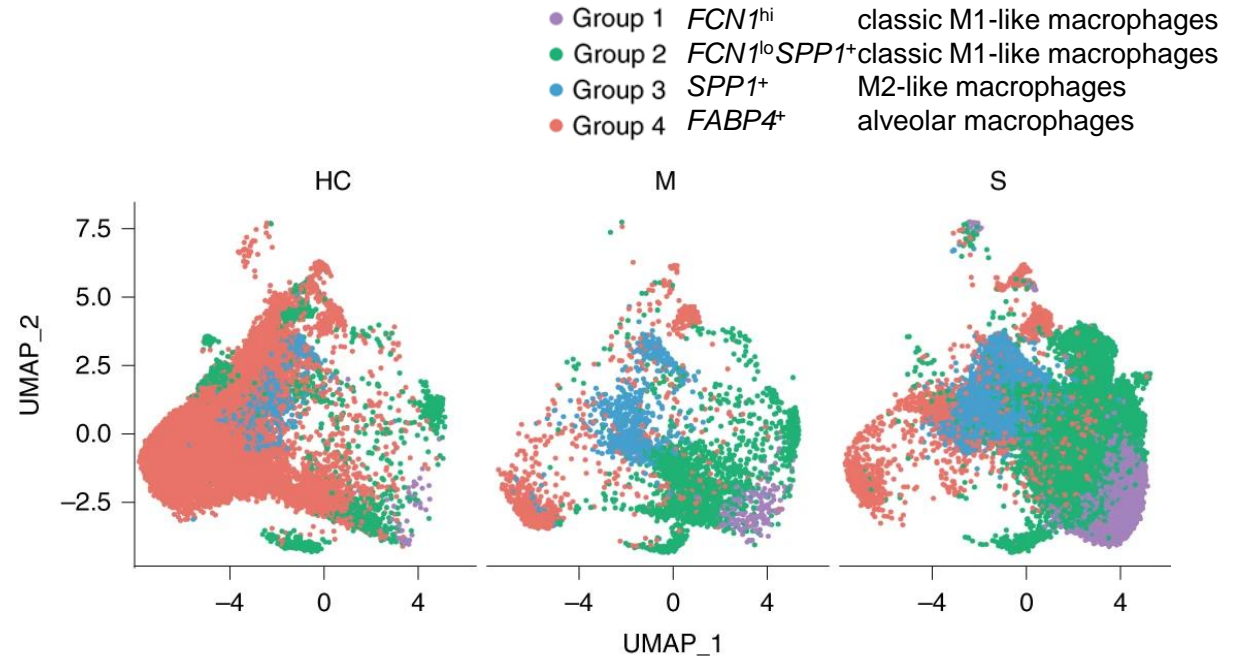
Clusters called by DBScan

**Dissecting the multicellular ecosystem of metastatic melanoma by single-cell RNA-seq.** Tirosh et al. *Science*. 2016 Apr 8;352(6282):189-96.

# Examples: Dimension reduction and clustering

- Single cell transcriptomics: development, cell type heterogeneity and cancer
- Bulk transcriptomics and epigenetics (cancer)
- Bulk transcriptomics: Effect of mutations

Bronchoalveolar lavage fluid macrophages from patients with varying severity of COVID-19 and from healthy people:

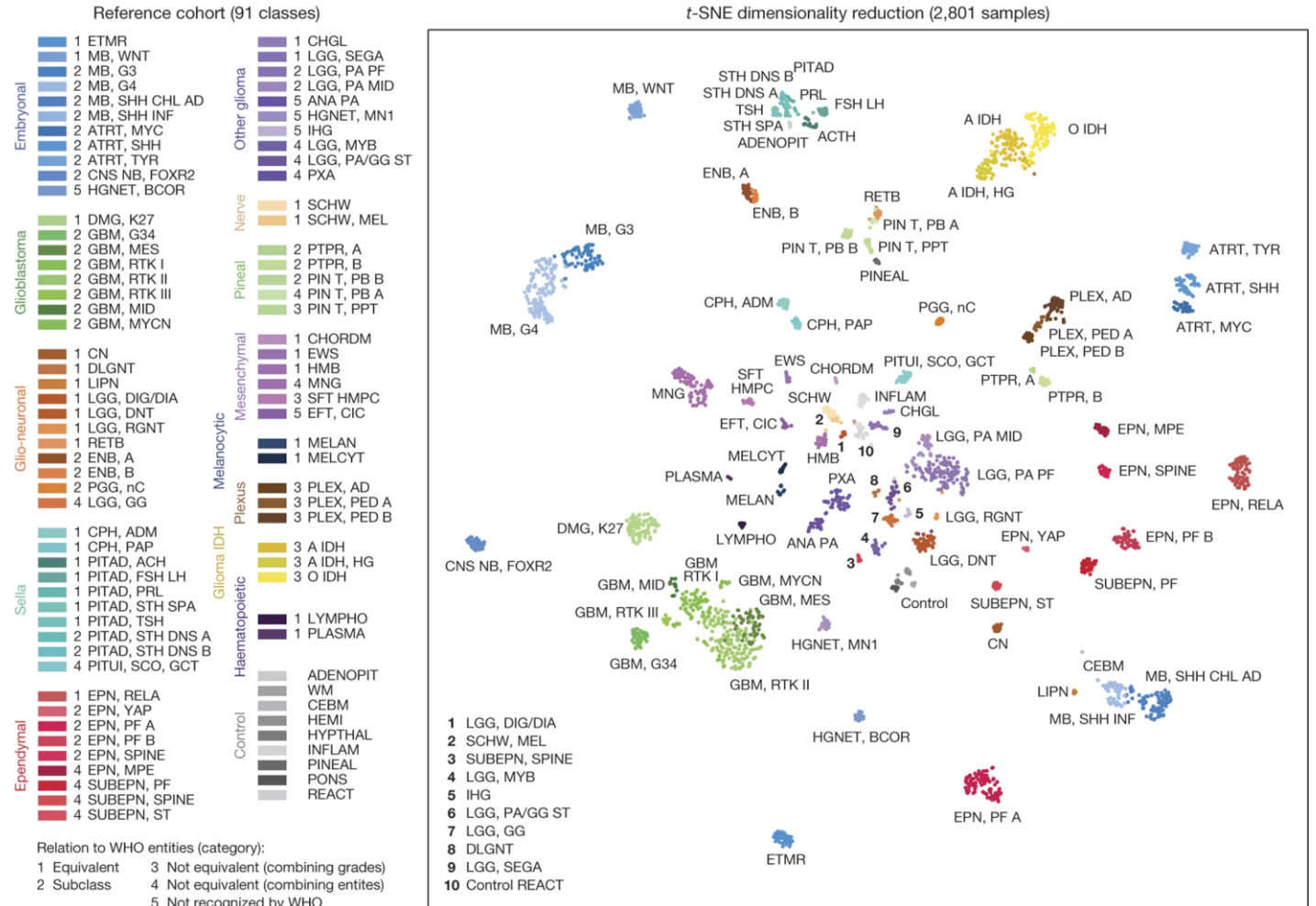


Severe cases: Presence of proinflammatory monocyte-derived macrophages

**Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19.** Liao et al., *Nature Medicine*. 2020

# Examples: Dimension reduction and clustering

- Single cell transcriptomics: development, cell type heterogeneity and cancer
- Bulk transcriptomics and epigenetics (cancer)
- Bulk transcriptomics: Effect of mutations

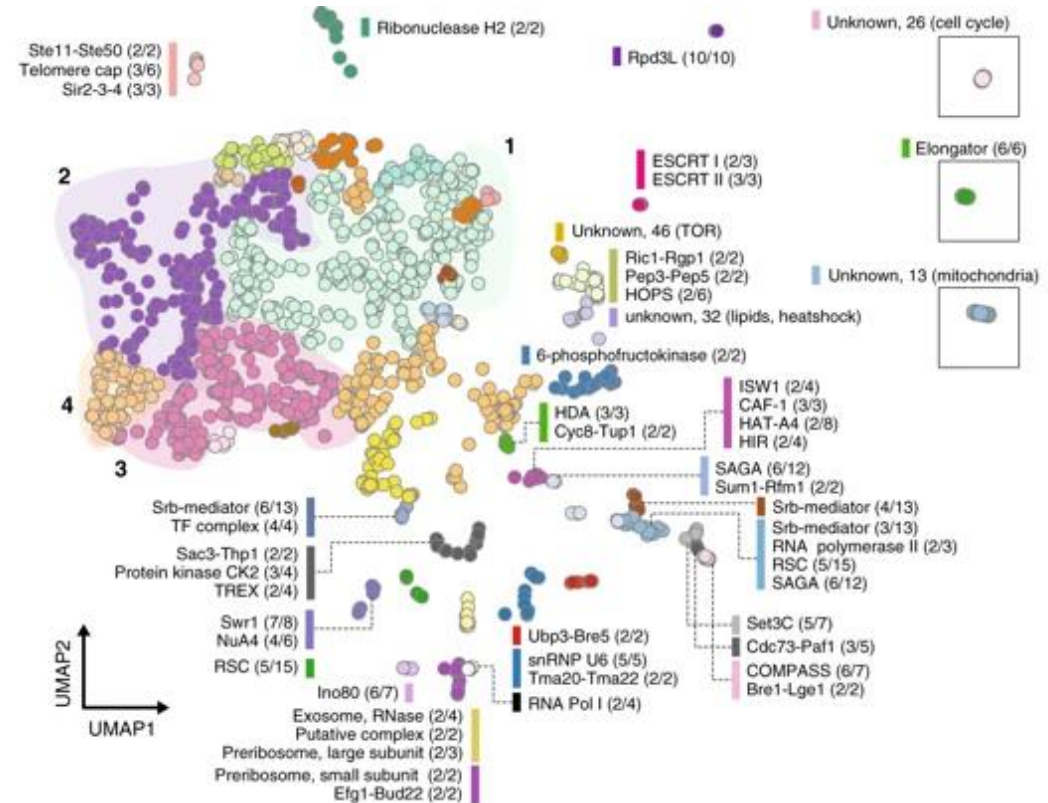


**DNA methylation-based classification of central nervous system tumours.** Capper et al. *Nature* 2018

# Examples: Dimension reduction and clustering

- Single cell transcriptomics: development, cell type heterogeneity and cancer
- Bulk transcriptomics and epigenetics (cancer)
- Bulk transcriptomics: Effect of mutations

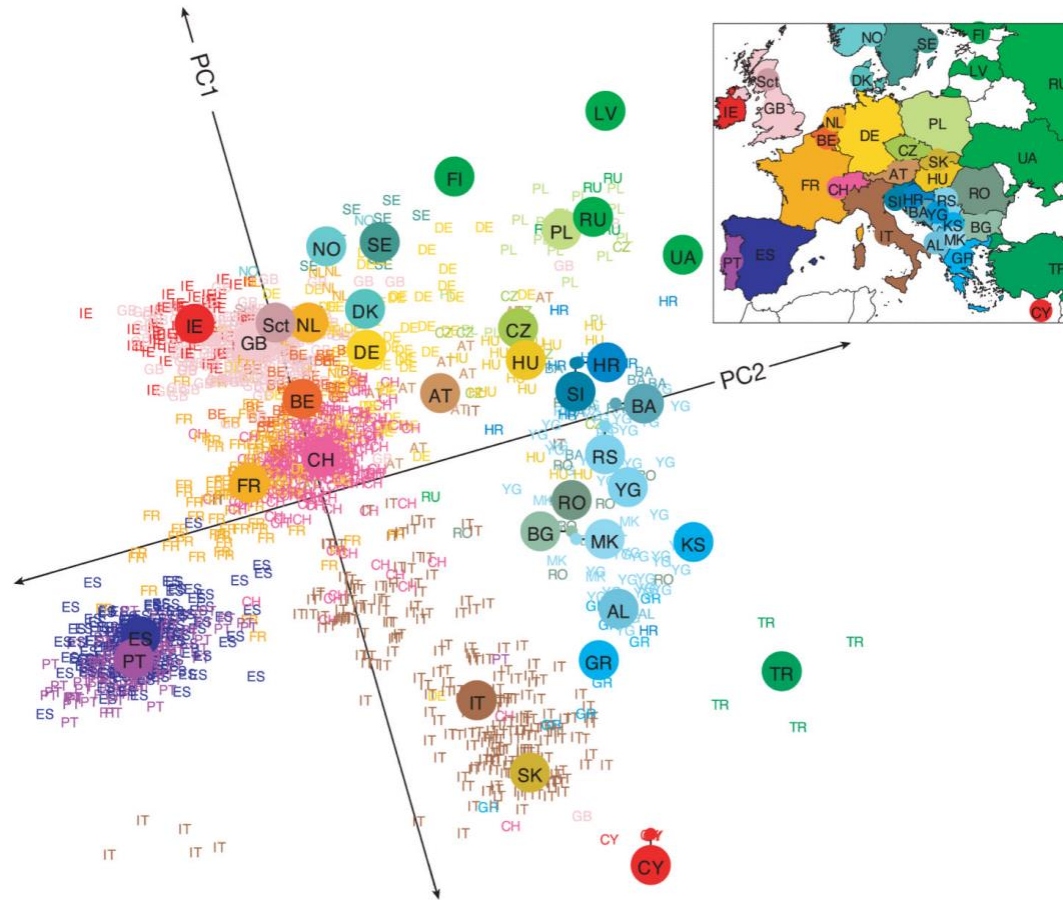
Transcript profiles of 1484 single gene deletions of *Saccharomyces cerevisiae* (baker's yeast)



**Dimensionality reduction by UMAP to visualize physical and genetic interactions.** Michael W. Dorrity et al., *Nature Comm.* 2020

# Examples: Dimension reduction and clustering

- Population genetics



PCA

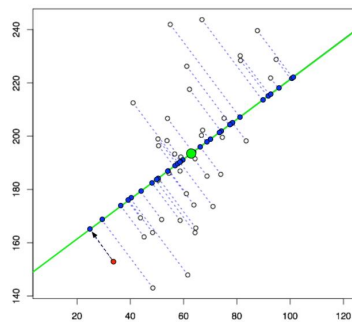
«Genes mirror geography within Europe» Novembre et al, *Nature*, 2008



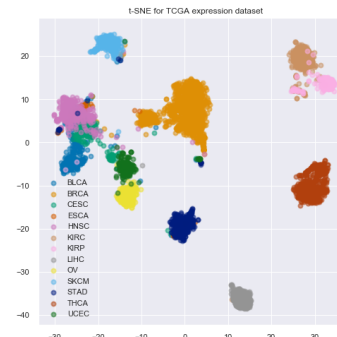
# Methods for dimension reduction

## Most widely used methods:

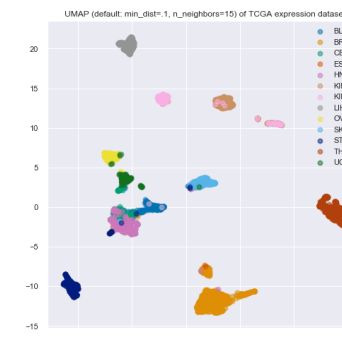
- Principal component analysis (PCA)
- t-distributed Stochastic Neighbor Embedding (tSNE) – check!
- Uniform Manifold Approximation and Projection (UMAP) – check!



PCA



t-SNE



UMAP

# Hands-on: gene expression data from several cancer types

- <https://github.com/BoevaLab/Teaching>
- Input: The Cancer Genome Atlas (TCGA) mRNA expression data

Harmonized Cancer Datasets  
Genomic Data Commons Data Portal

Get Started by Exploring:

Projects Exploration Analysis Repository

Quick Search Manage Sets Login Cart GDC Apps

Search: e.g. BRAF, Breast, TCGA-BLCA, TCGA-A5-A0G2

Data Portal Summary [Data Release 23.0 - April 07, 2020](#)

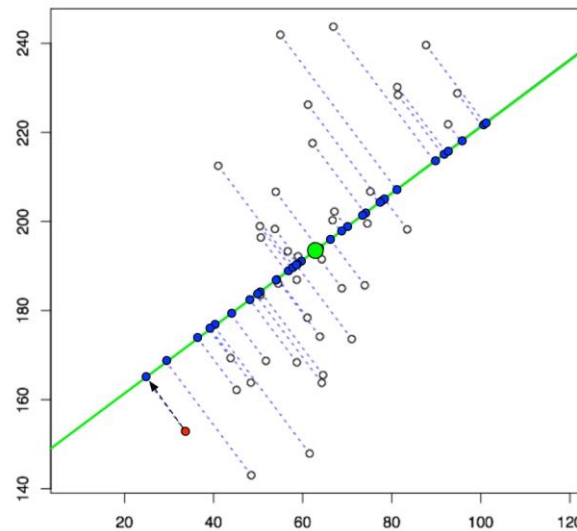
Category	Count
PROJECTS	64
FILES	559,345
PRIMARY SITES	67
GENES	22,872
CASES	83,709
MUTATIONS	3,142,246

Cases by Major Primary Site

Primary Site	Cases (in 1000s)
Adrenal Gland	0.1
Bile Duct	0.1
Bladder	0.1
Blood	0.1
Bone	0.1
Bone Marrow	0.1
Brain	0.1
Breast	0.1
Cervix	0.1
Colorectal	0.1
Esophagus	0.1
Eye	0.1
Head and Neck	0.1
Kidney	0.1
Liver	0.1
Lung	11.0
Lymph Nodes	0.1
Nervous System	0.1
Ovary	0.1
Pancreas	0.1
Pleura	0.1
Prostate	0.1
Skin	0.1
Soft Tissue	0.1
Stomach	0.1
Testis	0.1
Thymus	0.1
Thyroid	0.1
Uterus	0.1

# Principal component analysis (PCA)

- **PCA**: an **orthogonal linear transformation** that transforms the data to a new coordinate system such that the greatest variance by some scalar projection of the data comes to lie on the first coordinate (called the first principal component), the second greatest variance on the second coordinate, and so on.



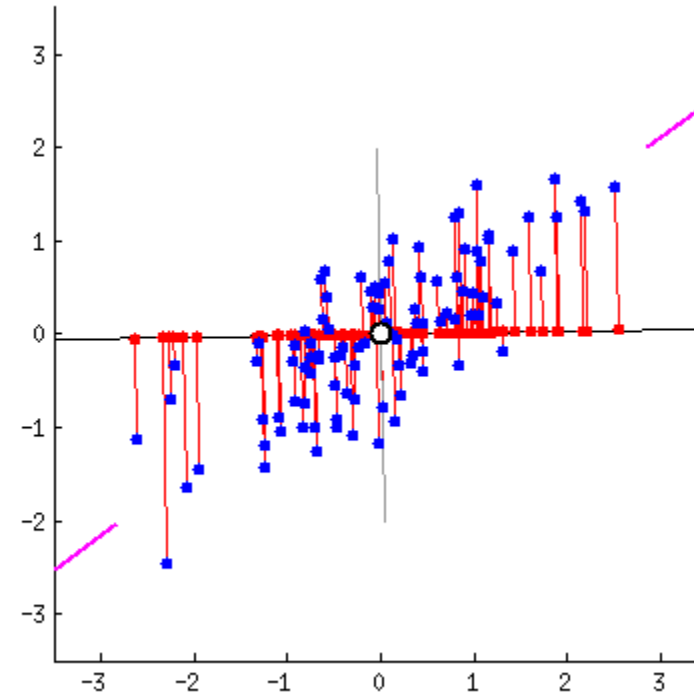
From <https://liorpachter.wordpress.com/2014/05/26/what-is-principal-component-analysis/>

# Principal component analysis (PCA)

The **first** principal component:  
the line that maximizes the variance (the average of the squared distances from the projected points (red dots) to the origin “o”).

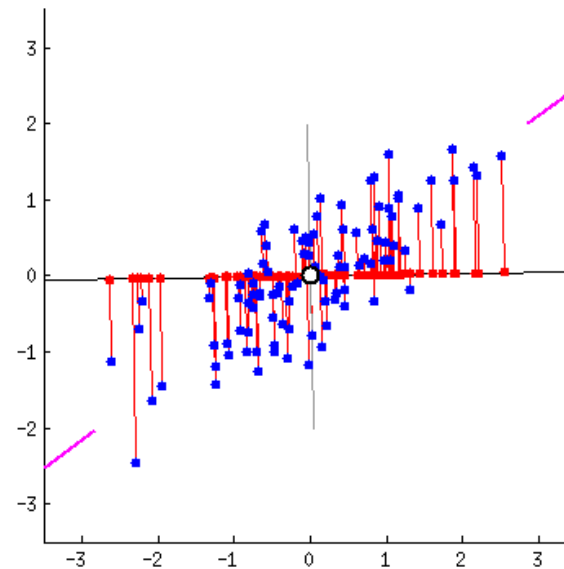
The **second** principal component is calculated in the same way, with the condition that it is perpendicular to the first principal component and that it accounts for the next highest variance.

Etc.



# Principal component analysis (PCA)

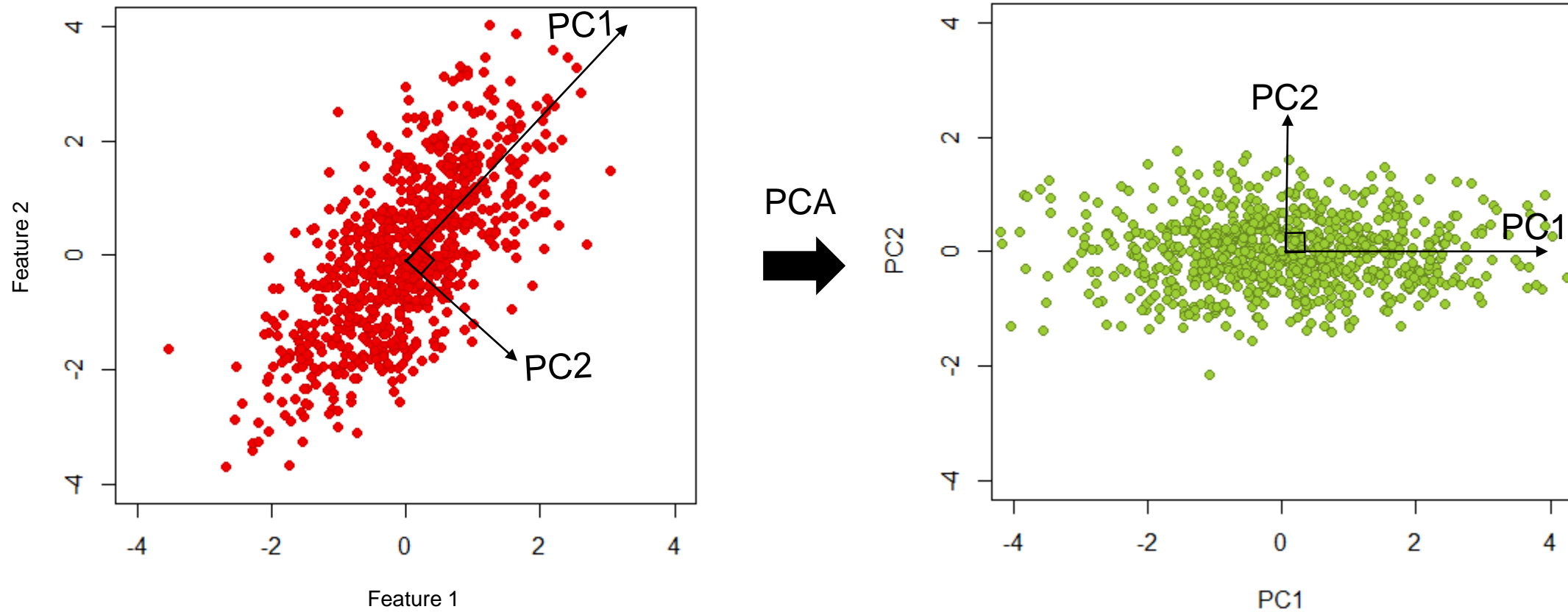
- Given  $n$  points in  $\mathbb{R}^p$ , principal components analysis consists of choosing a dimension  $k < p$  and then finding the affine space of dimension  $k$  with the property that the squared distance of the points to their orthogonal projection onto the space is minimized.



# Principal component analysis (PCA)

Generally, one standardizes the data along each dimension before applying PCA.

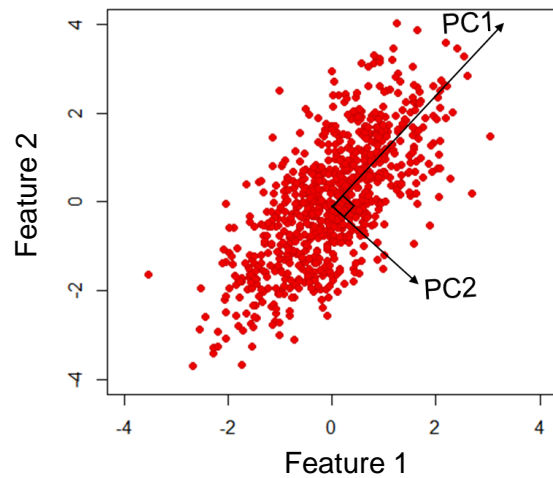
PCA: an orthogonal linear transformation



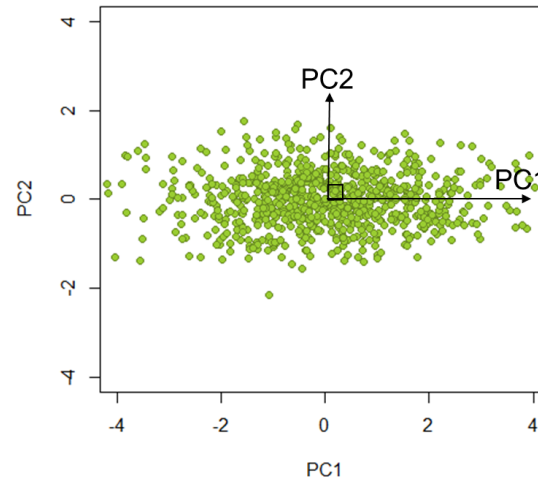
# Principal component analysis (PCA)

Generally, one standardizes the data along each dimension before applying PCA.

**Q:** How will look the PCA transformation of a PCA?

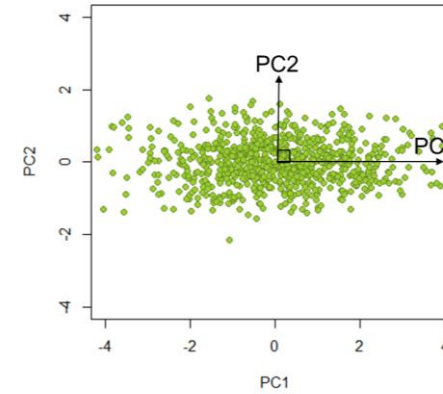


PCA  
➔

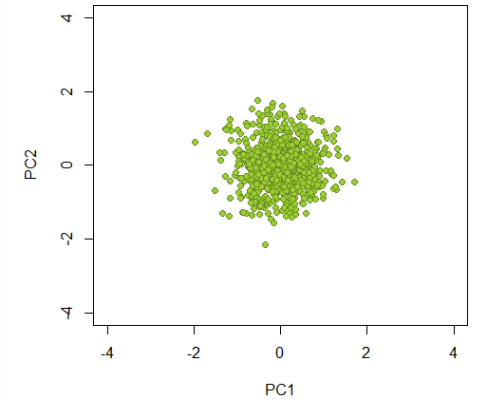


?  
PCA  
➔

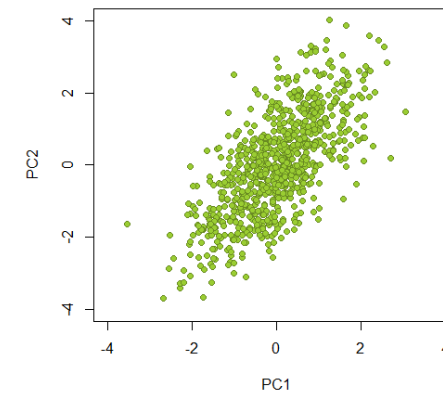
(1) the same



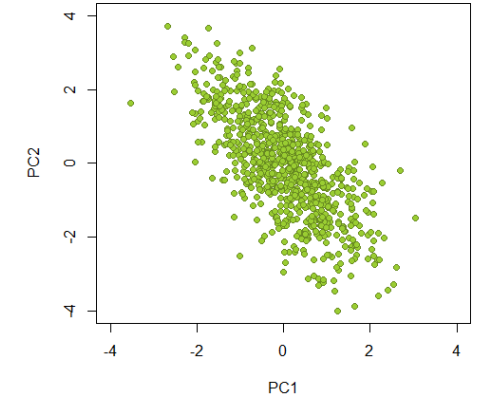
(2) the "ball"



(3) as original set

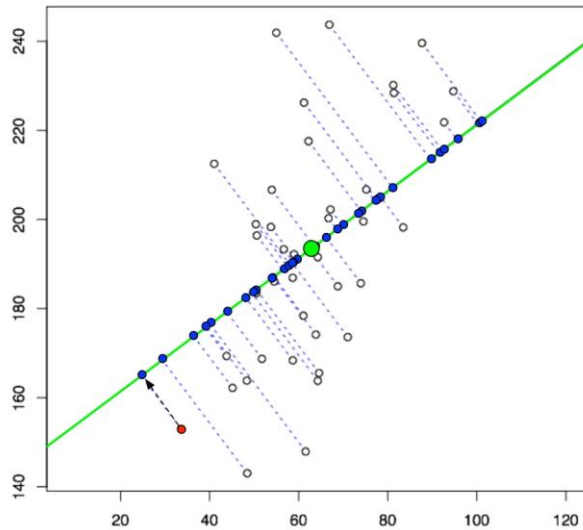


(4) Inverted original



# Principal component analysis (PCA)

- PCA: an orthogonal linear transformation that transforms the data to a new coordinate system such that the greatest variance by some scalar projection of the data comes to lie on the first coordinate (called the first principal component), the second greatest variance on the second coordinate, and so on.

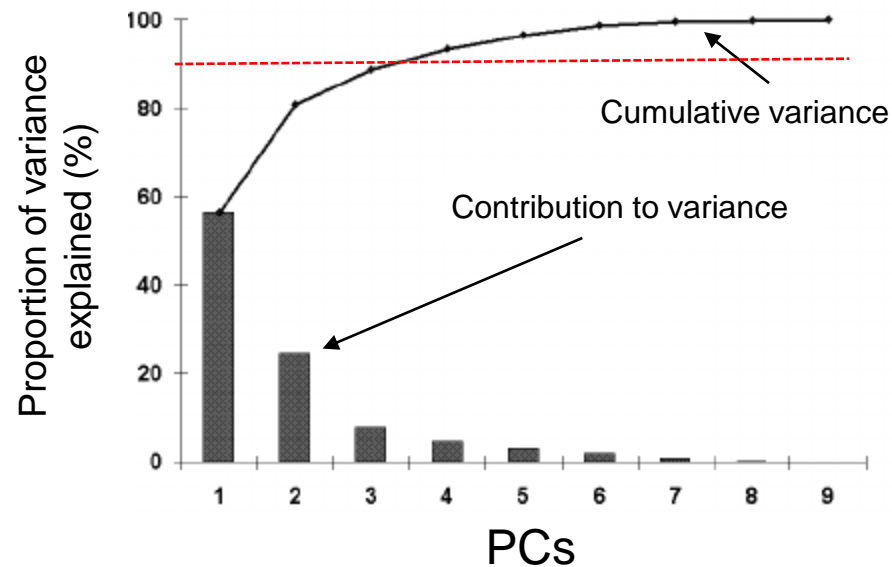


Given  $n$  points in  $R^p$ , principal components analysis consists of choosing a dimension  $k < p$  and then finding the affine space of dimension  $k$  with the property that the squared distance of the points to their orthogonal projection onto the space is minimized.



# Principal component analysis (PCA)

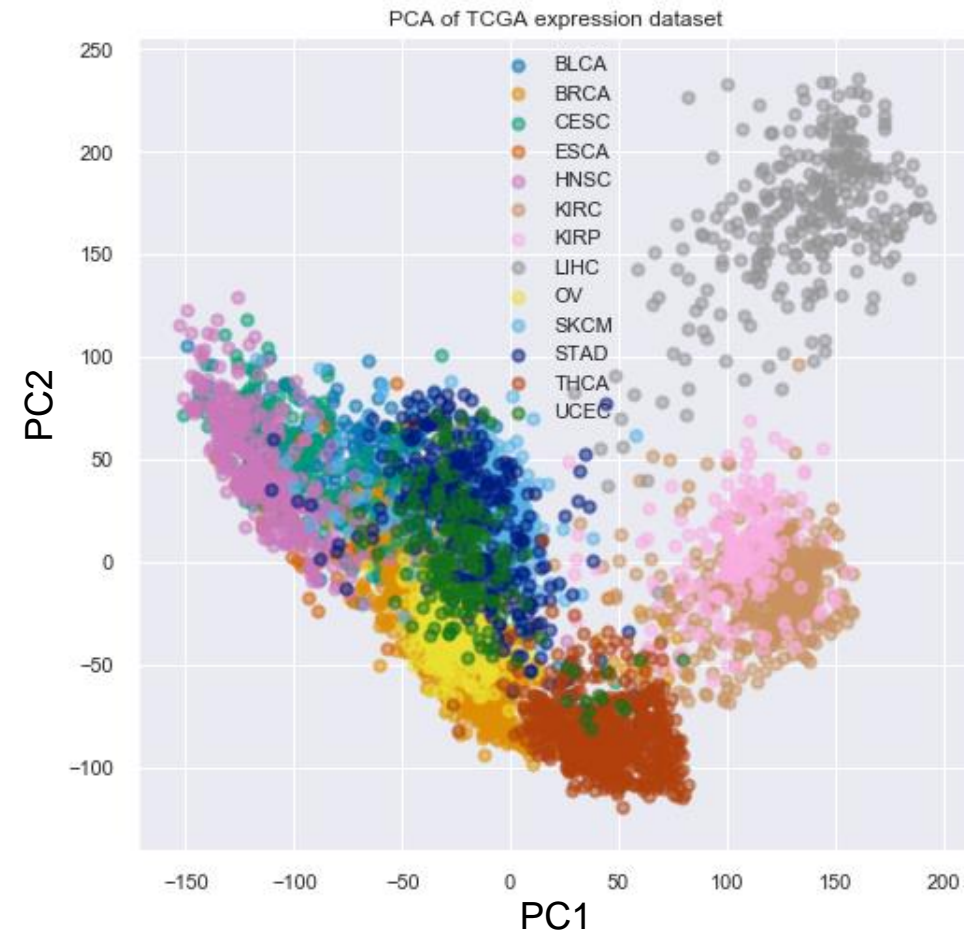
- PCA is a deterministic method, the only parameter one can choose is  $k$  for how many principal components to keep.
- Choosing  $k$  based on the proportion of the variance explained.



$$Var\_explained_k = \frac{\lambda_k}{\sum_i \lambda_i}, \text{ where } \lambda_k \text{ is } k^{\text{th}} \text{ eigenvalue.}$$

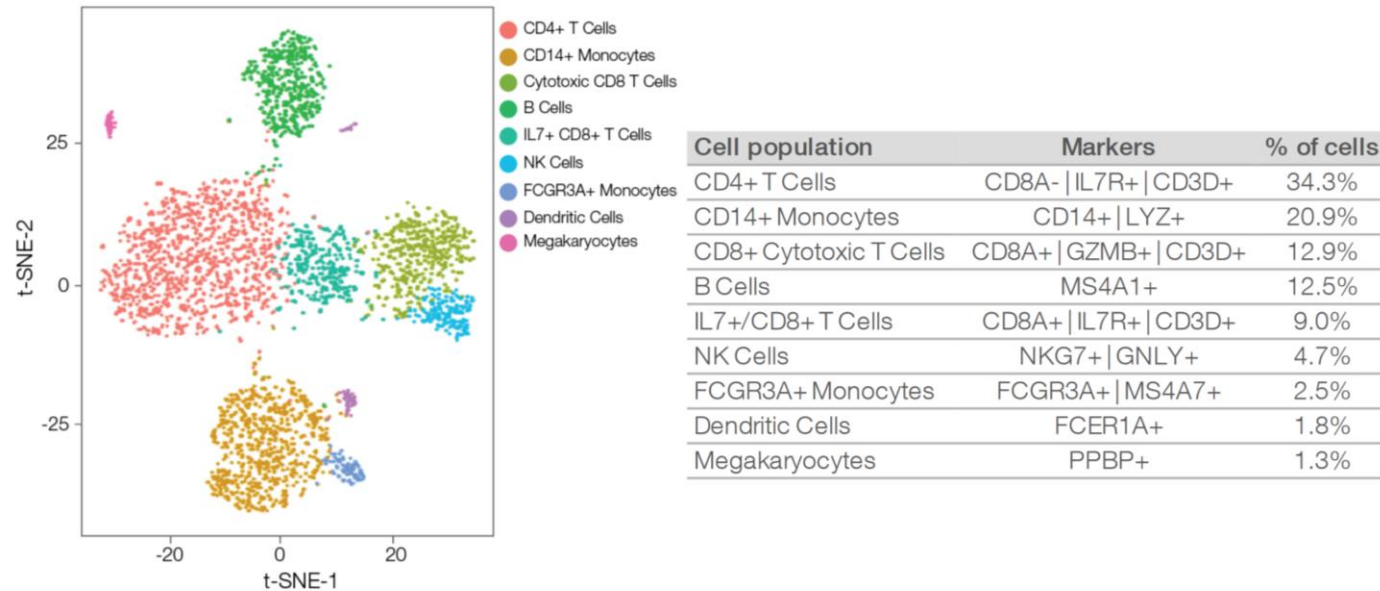
# Let's go to the Jupiter Notebook to see the result of PCA on our toy data set

- <https://github.com/BoevaLab/Teaching>
- Input: The Cancer Genome Atlas (TCGA) mRNA expression data



# t-distributed Stochastic Neighbor Embedding (tSNE)

- tSNE: nonlinear dimensionality reduction technique, converts similarities between data points to joint probabilities and tries to minimize the Kullback-Leibler divergence between the joint probabilities of the low-dimensional embedding and the high-dimensional data.



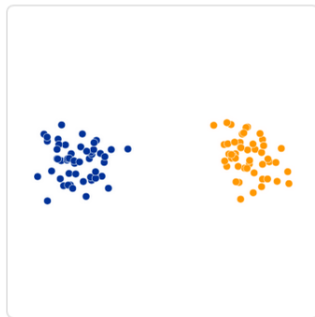
- t-SNE has a cost function that is not convex, i.e., with different initializations we can get different results.

[First introduced by van der Maaten & Hinton paper from 2008.](#)

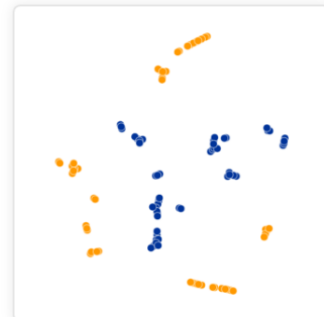
# t-distributed Stochastic Neighbor Embedding (tSNE)

tSNE method's the most important parameter:

- **Perplexity**, scikit-learn recommended range: [5, 50], default: 30



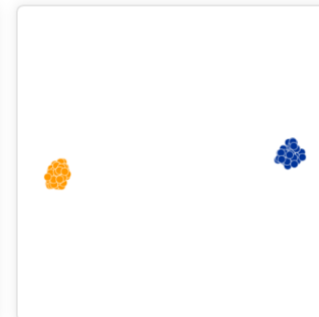
*Original*



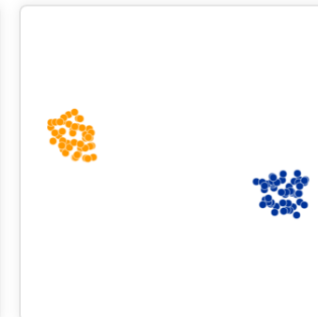
Perplexity: 2  
Step: 5,000



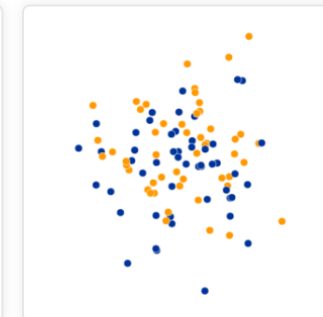
Perplexity: 5  
Step: 5,000



Perplexity: 30  
Step: 5,000



Perplexity: 50  
Step: 5,000

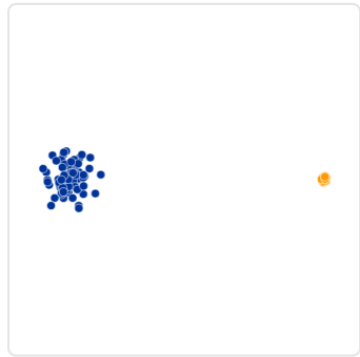


Perplexity: 100  
Step: 5,000

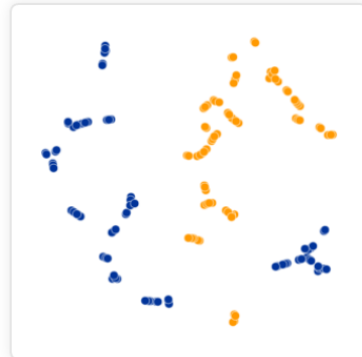
# t-distributed Stochastic Neighbor Embedding (tSNE)

tSNE method's particularities:

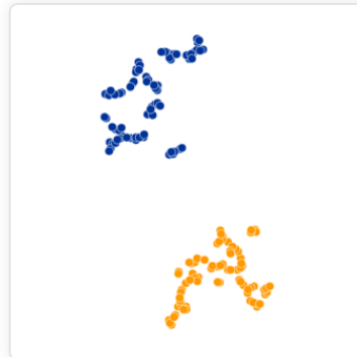
- Cluster sizes in a t-SNE plot mean nothing



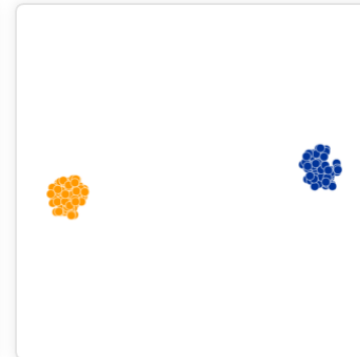
*Original*



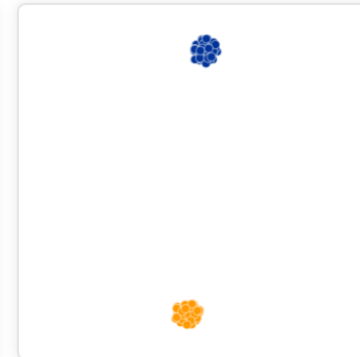
Perplexity: 2  
Step: 5,000



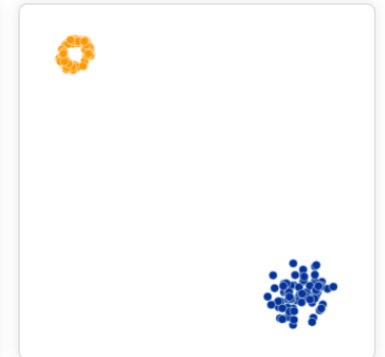
Perplexity: 5  
Step: 5,000



Perplexity: 30  
Step: 5,000



Perplexity: 50  
Step: 5,000



Perplexity: 100  
Step: 5,000

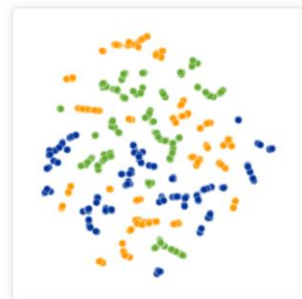
# t-distributed Stochastic Neighbor Embedding (tSNE)

tSNE method's particularities:

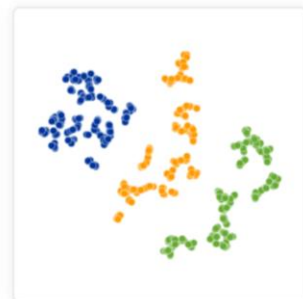
- Cluster sizes in a t-SNE plot mean nothing
- Distances between clusters might not mean anything



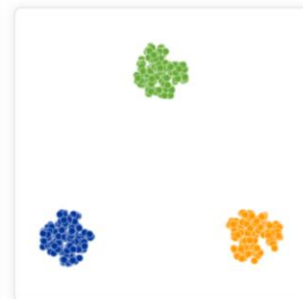
*Original*



Perplexity: 2  
Step: 5,000



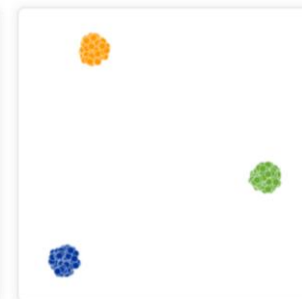
Perplexity: 5  
Step: 5,000



Perplexity: 30  
Step: 5,000



Perplexity: 50  
Step: 5,000



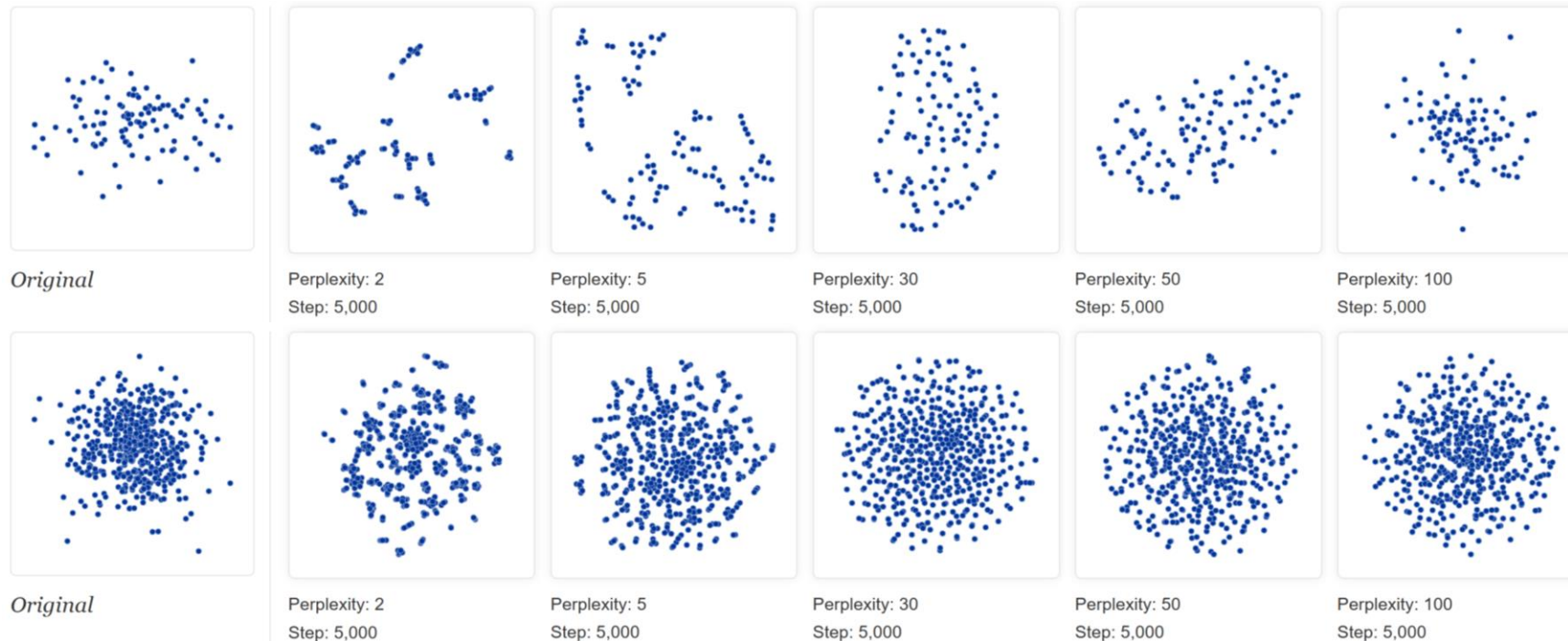
Perplexity: 100  
Step: 5,000

200 points/cluster

# t-distributed Stochastic Neighbor Embedding (tSNE)

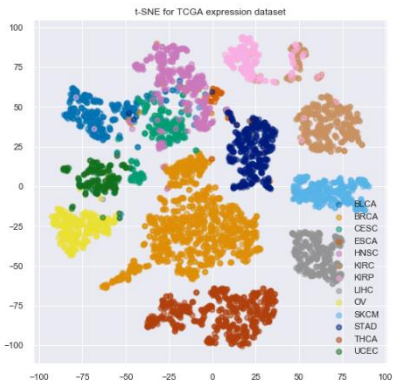
tSNE method's particularities:

- Cluster sizes in a t-SNE plot mean nothing
- Distances between clusters might not mean anything
- Random noise does not always look random, and sometimes one can see some shapes

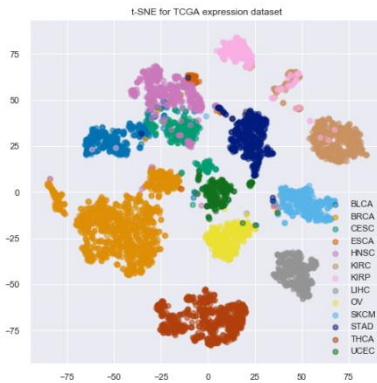


# What would you choose as the best value of perplexity in this example?

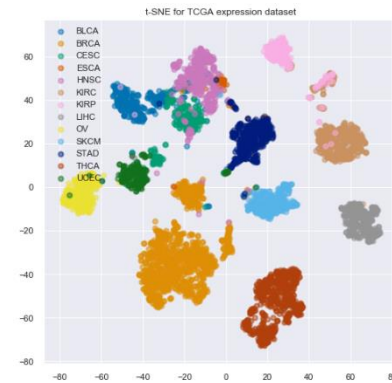
- BLCA Bladder
- BRCA Breast
- CESC Cervical squamous cell
- ESCA Esophageal
- HNSC Head and Neck squamous cell
- KIRC Kidney clear cell
- KIRP Kidney papillary
- LIHC Liver
- OV Ovarian
- SKCM Skin Cutaneous Melanoma
- STAD Stomach
- THCA Thyroid
- UCEC Uterine Corpus Endometrial



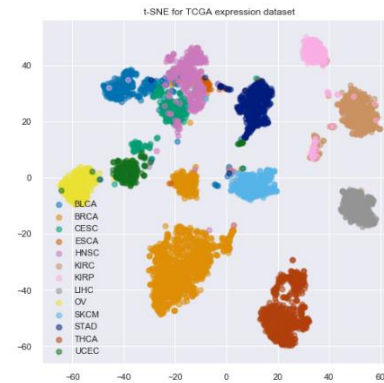
Perplexity: 10



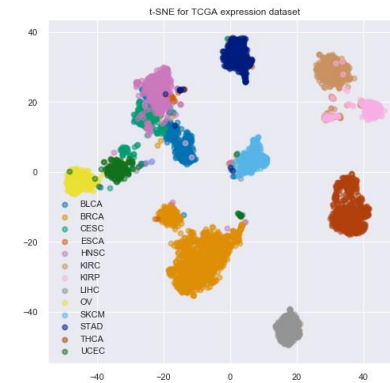
20



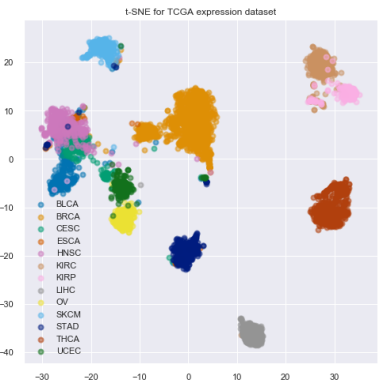
30



50



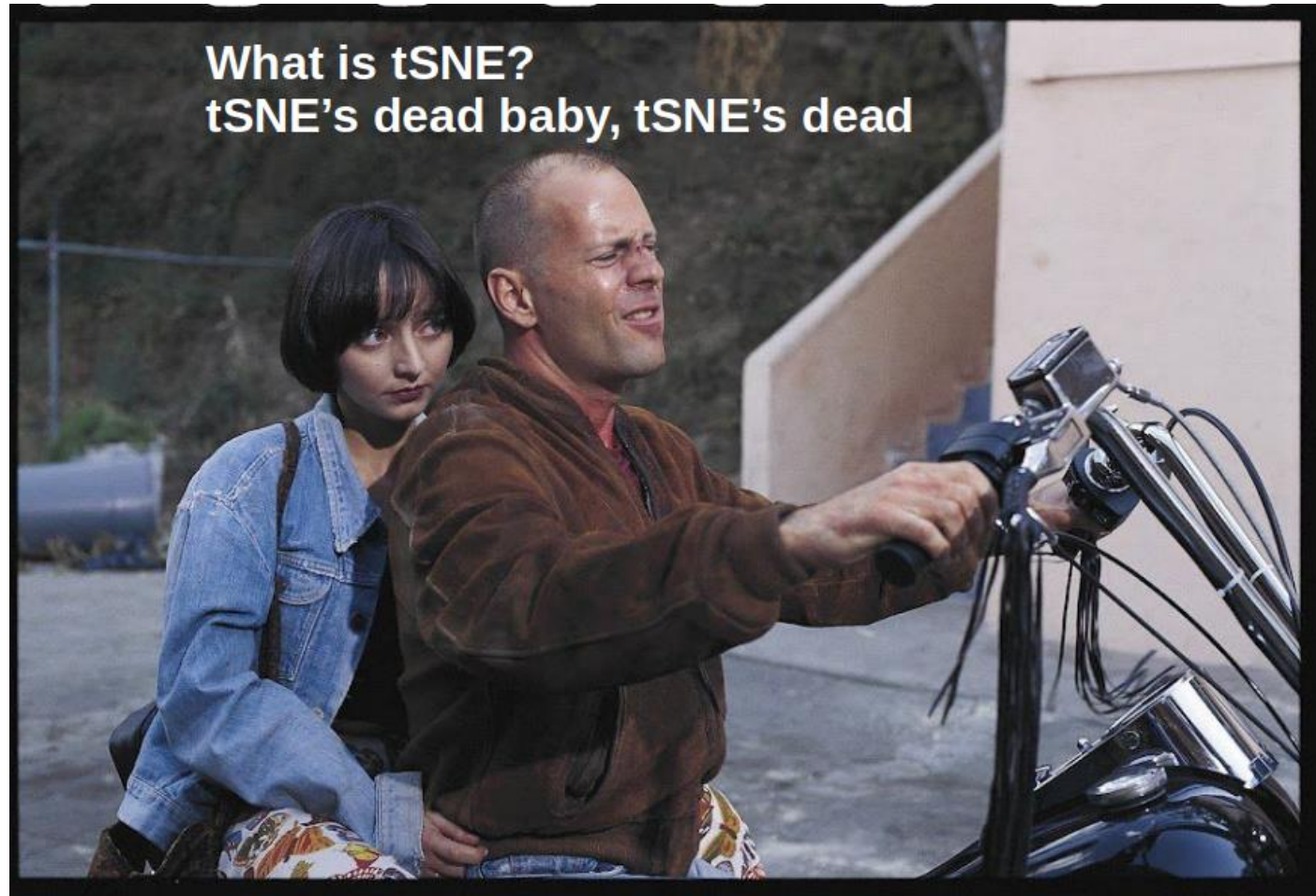
100



200



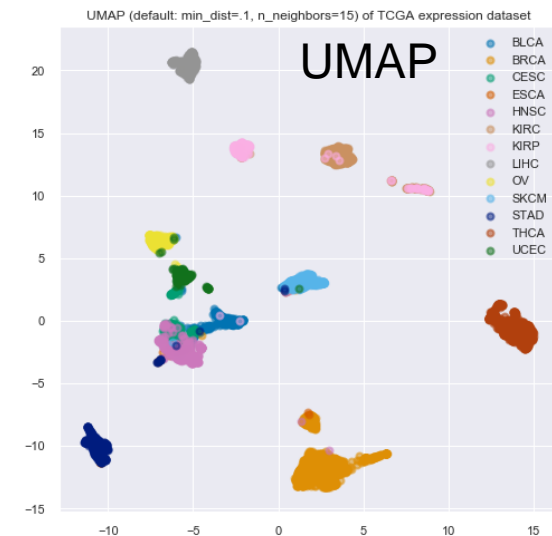
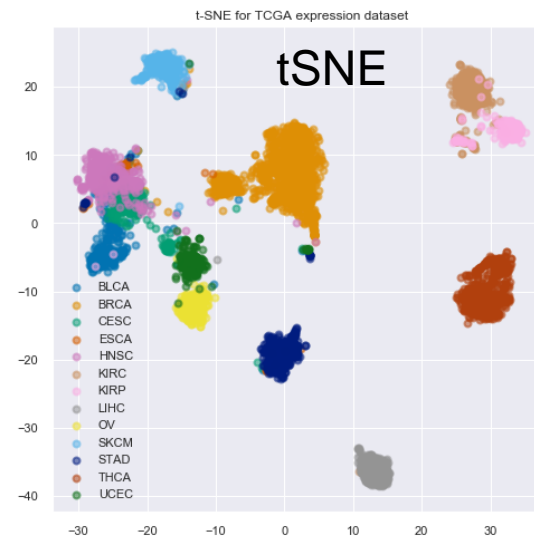
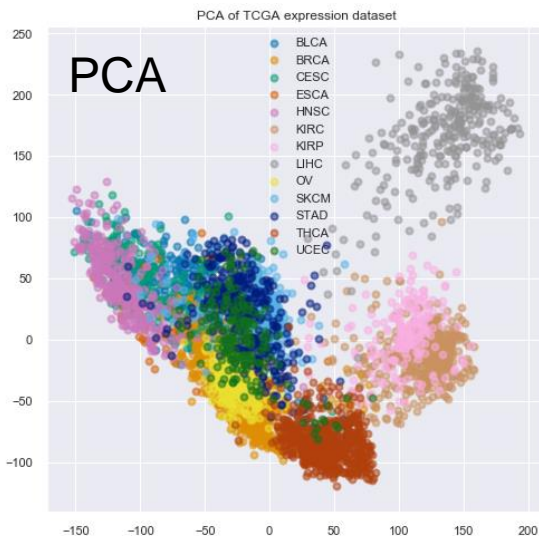
# Uniform Manifold Approximation and Projection (UMAP)



# Uniform Manifold Approximation and Projection (UMAP)

UMAP: nonlinear dimensionality reduction technique. Idea is similar to tSNE, but

- Much faster
- Not limited to the first 2-3 dimensions
- Uses binary cross-entropy as a cost function instead of the KL-divergence
- Better preserves global structure
- Uses the number of nearest neighbors and min-dist correction instead of perplexity



First introduced by [McInnes, L, Healy, J, ArXiv e-prints 1802.03426, 2018](#)

# Uniform Manifold Approximation and Projection (UMAP)

UMAP's the most important parameters:

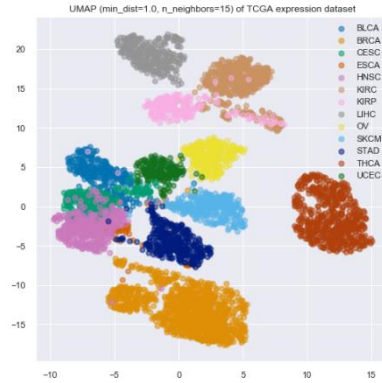
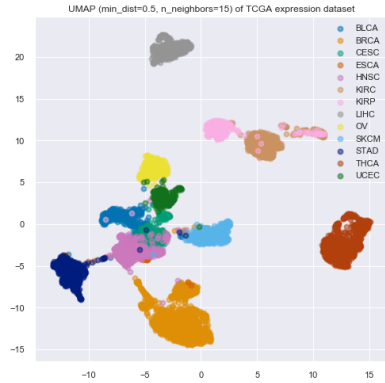
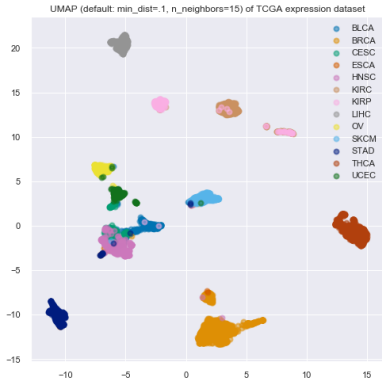
**min\_dist**, recommended range: [0.1, 1], default: 0.1

- Controls how tightly the embedding is allowed compress points together. Larger values ensure embedded points are more evenly distributed, while smaller values allow the algorithm to optimise more accurately with regard to local structure. Sensible values are in the range 0.001 to 0.5, with 0.1 being a reasonable default.

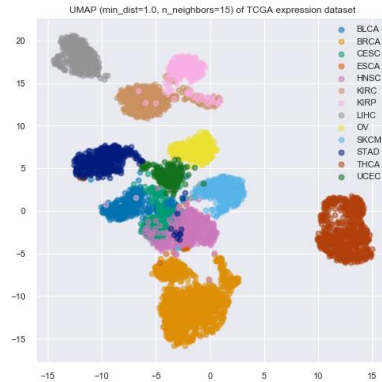
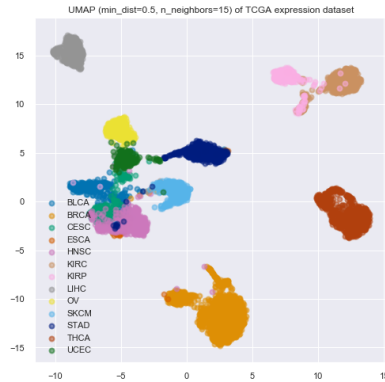
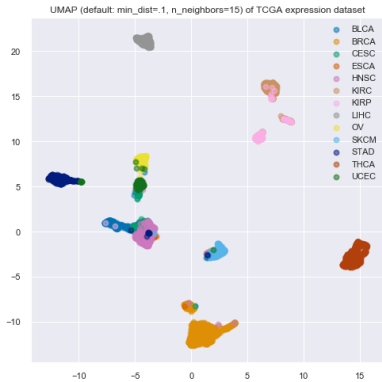
**n\_neighbors**, recommended range: [2, 100], default: 15

- Determines the number of neighboring points used in local approximations of manifold structure. Larger values will result in more global structure being preserved at the loss of detailed local structure. In general, this parameter should often be in the range 5 to 50, with a choice of 10 to 15 being a sensible default.

# What would you choose as the best value of **min\_dist** and **n\_neighbors** for this example?



n\_neighbors: 15



n\_neighbors: 50

Min\_dist: 0.1

0.5

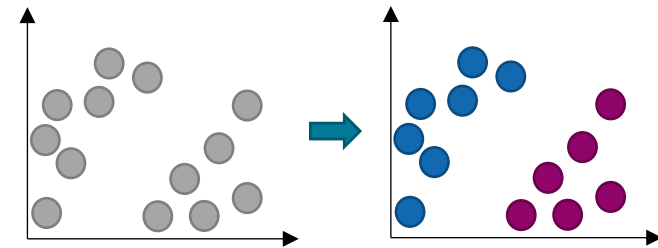
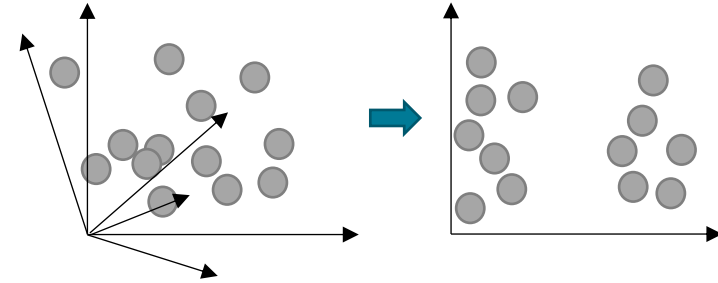
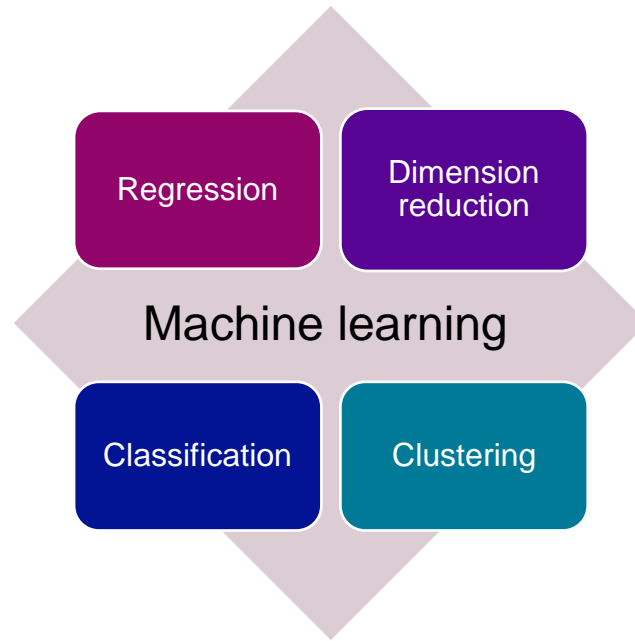
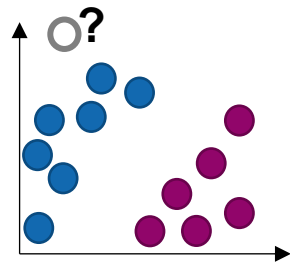
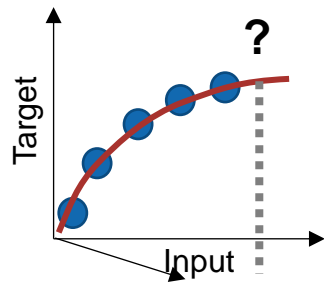
1

● BLCA	Bladder
● BRCA	Breast
● CESC	Cervical squamous cell
● ESCA	Esophageal
● HNSC	Head and Neck squamous cell
● KIRC	Kidney clear cell
● KIRP	Kidney papillary
● LIHC	Liver
● OV	Ovarian
● SKCM	Skin Cutaneous Melanoma
● STAD	Stomach
● THCA	Thyroid
● UCEC	Uterine Corpus Endometrial

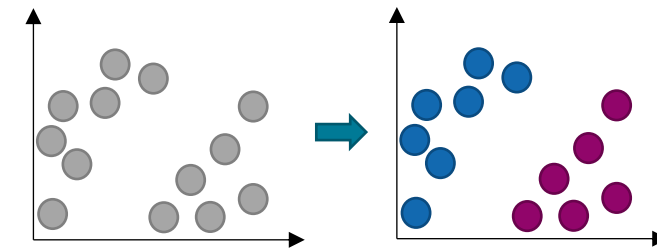
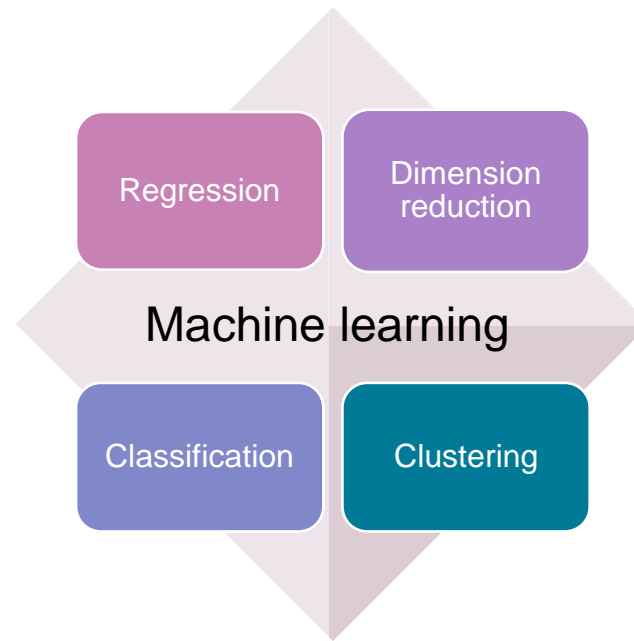
# Take home message: dimensionality reduction

- One should often try different methods with different parameters to choose the one that fits the best to our expectations from the data
- Random noise does not always look random, and sometimes one can see shapes
- Projections on the first  $n$  principal components can be used as input (instead of the original  $X$ ) to other dimension reduction methods such as tSNE to reduce execution time.

# Map of classical machine learning methods

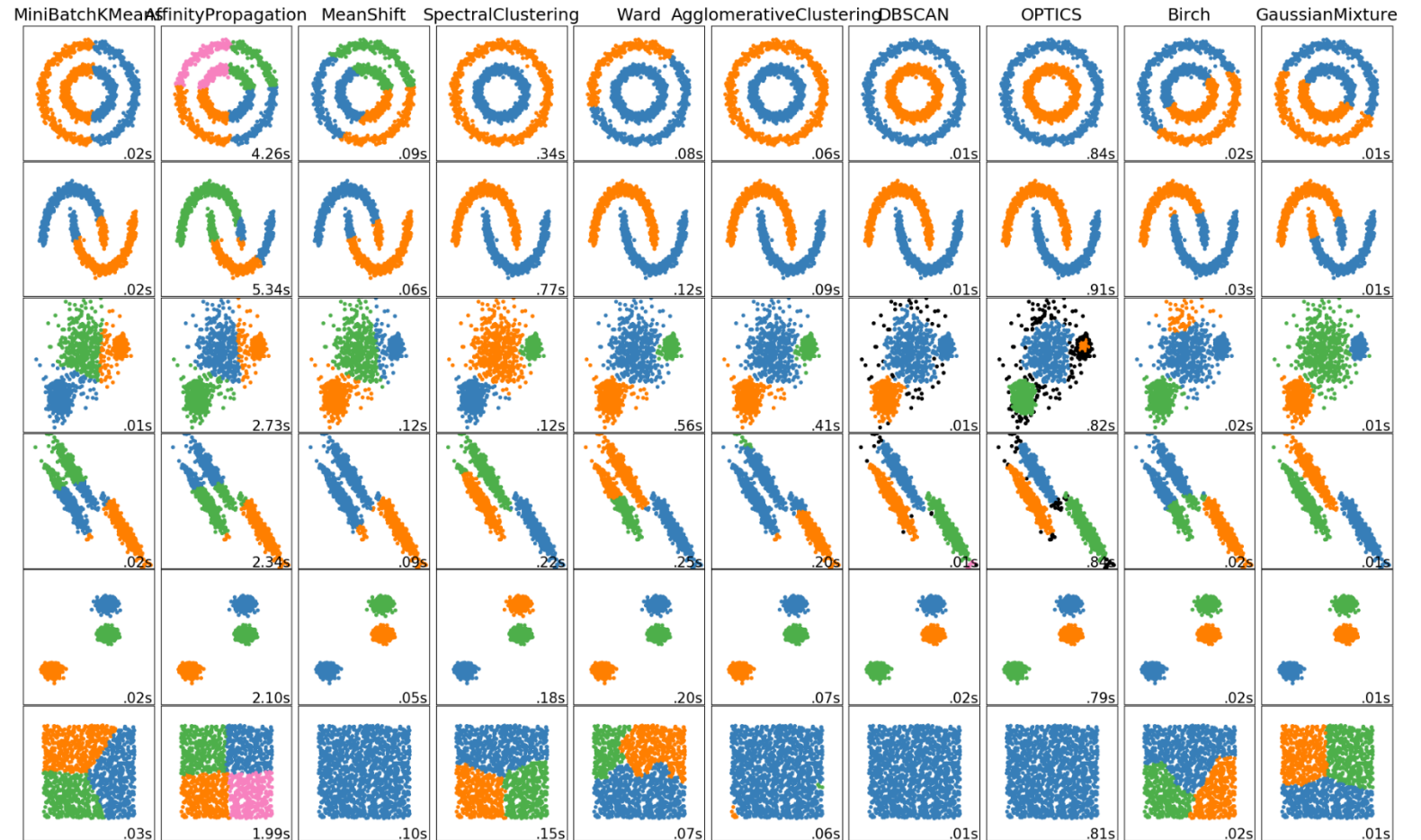


# Map of classical machine learning methods



# Clustering methods

- K-means
- Gaussian mixture models
- Spectral clustering
- Hierarchical clustering

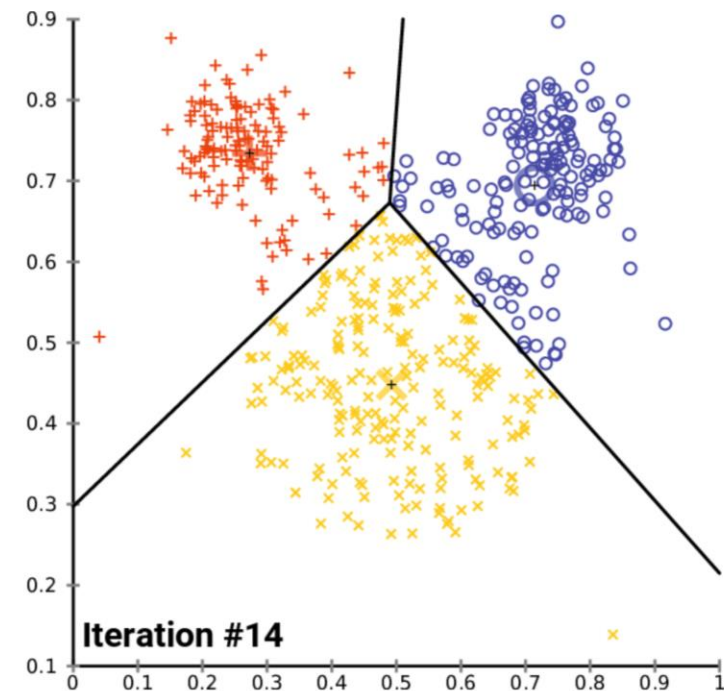
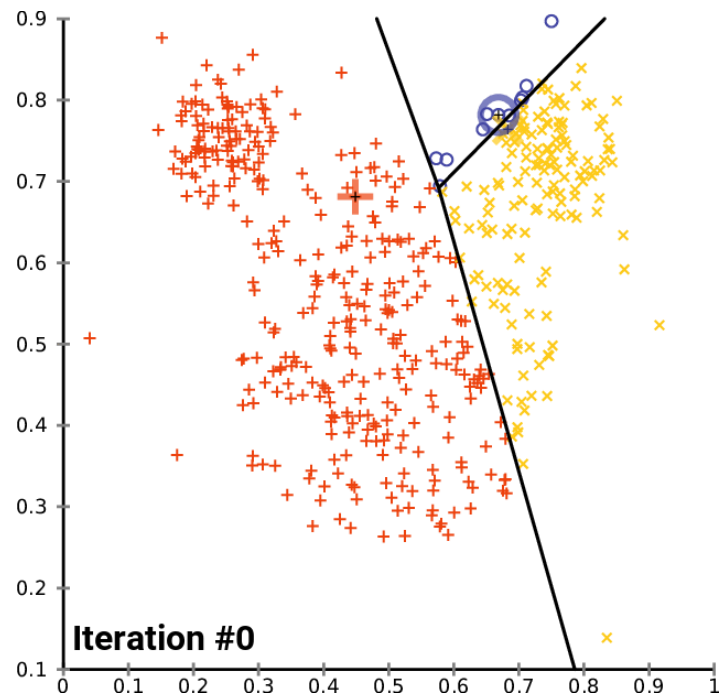


From <https://scikit-learn.org/stable/modules/clustering.html>



# K-means

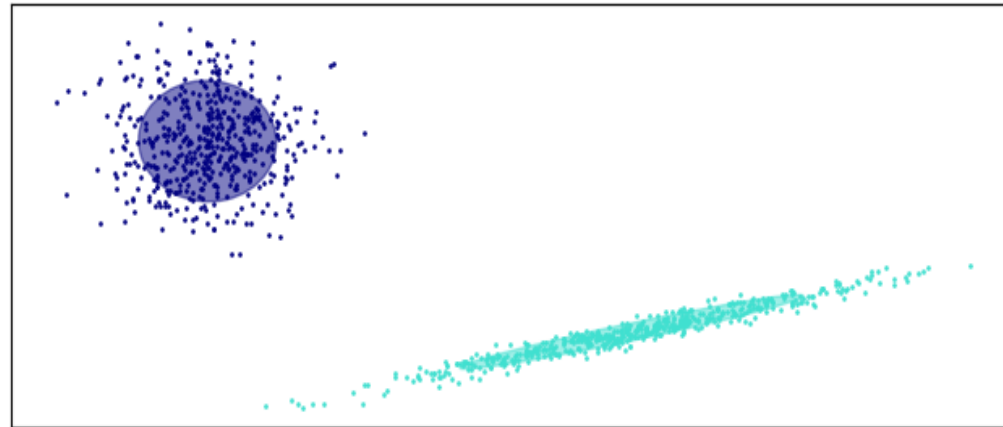
- k-means: aims to partition  $n$  observations into  $k$  clusters in which each observation belongs to the cluster with the nearest mean (cluster centers)



- k-means clustering tends to find clusters of comparable spatial extent

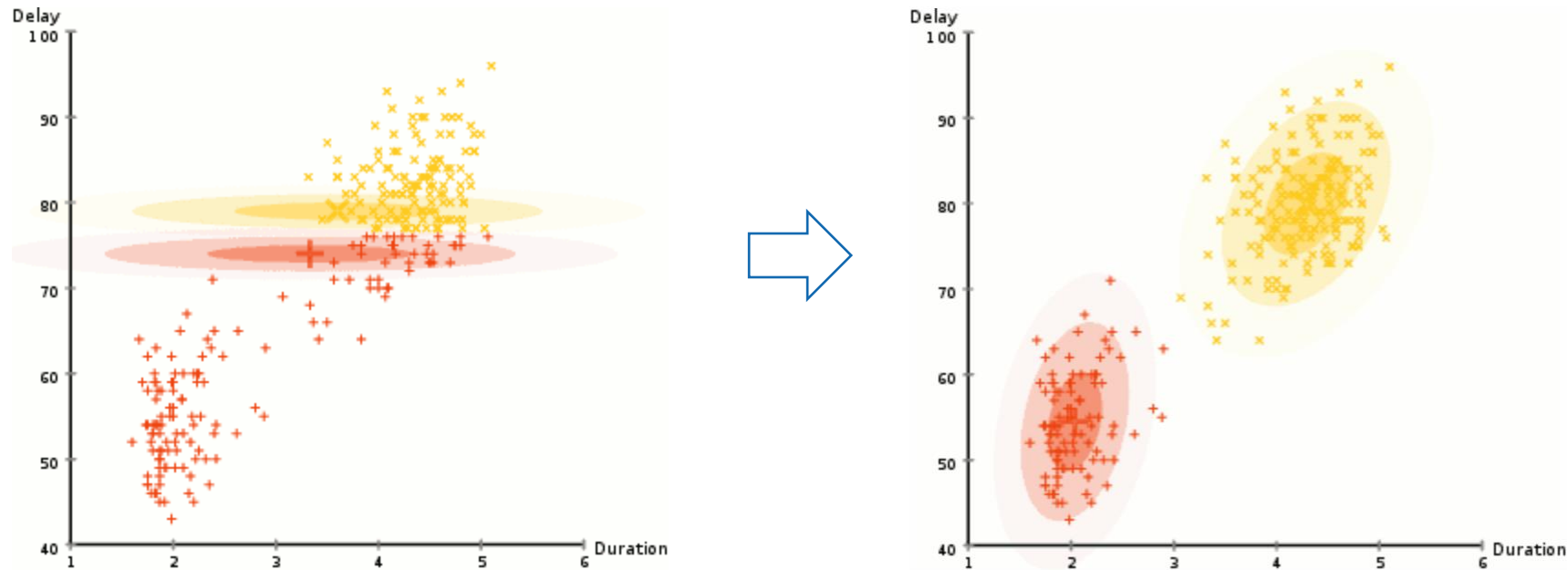
# Gaussian mixture models (GMM)

**GMM:** probabilistic model that assumes all the data points are generated from a mixture of a finite number of Gaussian distributions with unknown parameters



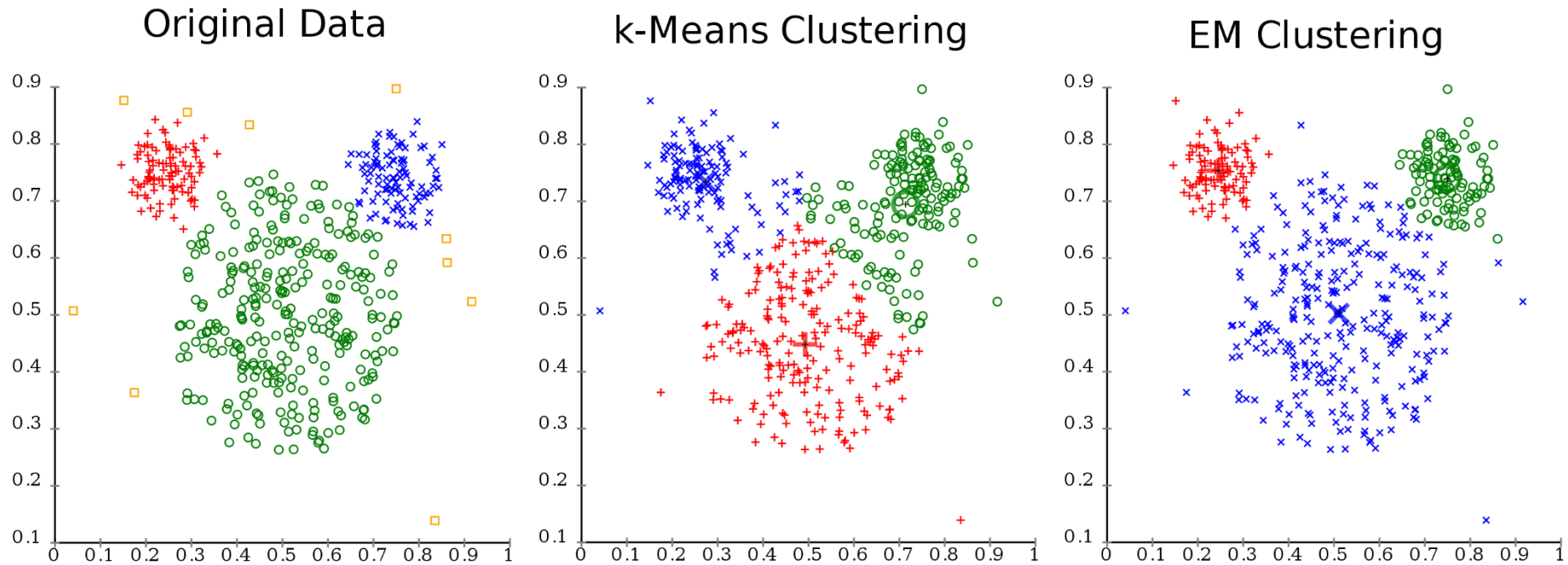
# Gaussian mixture models (GMM)

**GMM:** probabilistic model that assumes all the data points are generated from a mixture of a finite number of Gaussian distributions with unknown parameters



k-means clustering tends to find clusters of comparable spatial extent, while the GMM expectation-maximization mechanism allows clusters to have different shapes.

Different cluster analysis results on "mouse" data set:

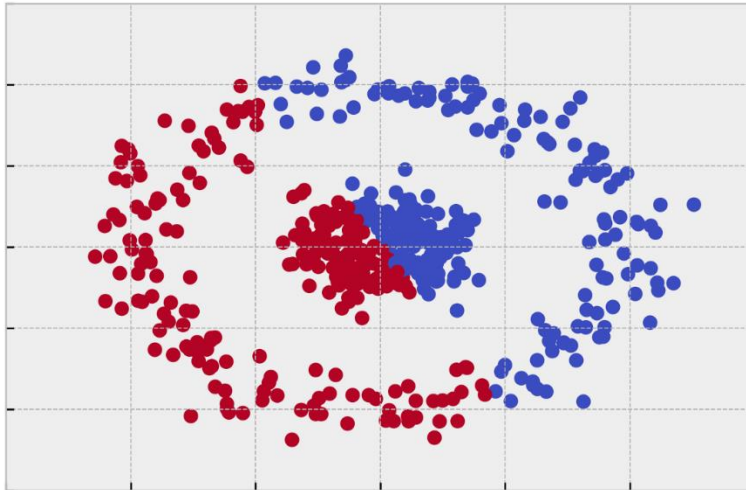


Also, GMMs support mixed membership

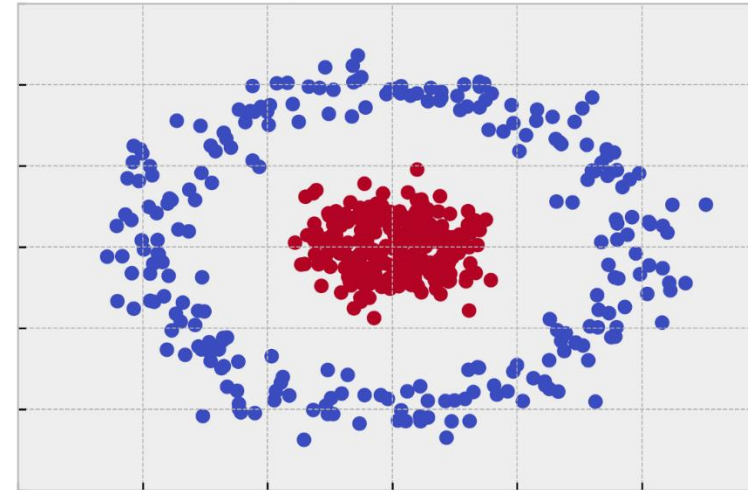
# Spectral clustering: relies on the assumption that “close” points should belong in the same cluster

- Spectral clustering: uses a standard clustering method (e.g., k-means) on relevant eigenvectors of a Laplacian matrix  $L$  of symmetric data similarity matrix  $A$ . – can also use k-nearest neighbors graphs for construction of  $A$ .
- $L := D - A$ , where  $D$  is the diagonal matrix, such as  $D_{ii} = \sum_j A_{ij}$ .

K-Means Circles

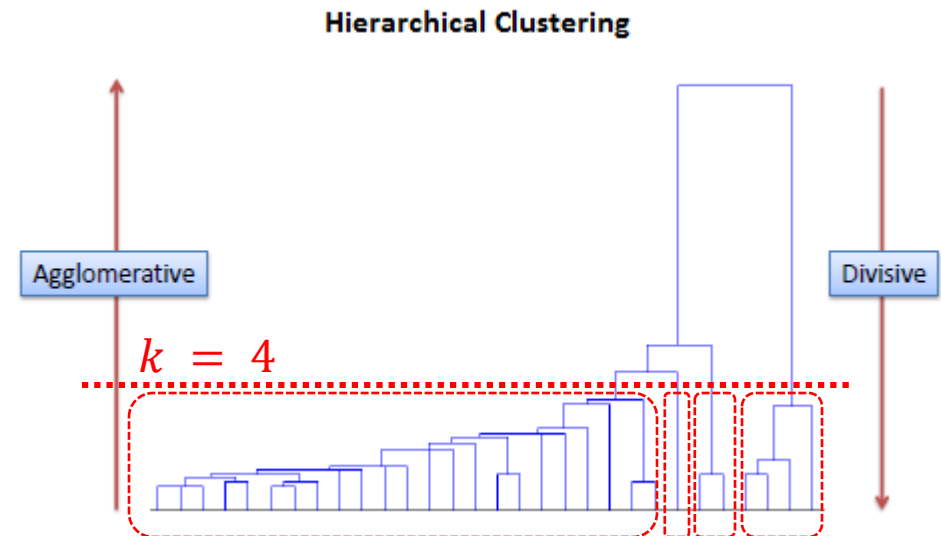


Spectral Clustering



# Hierarchical clustering

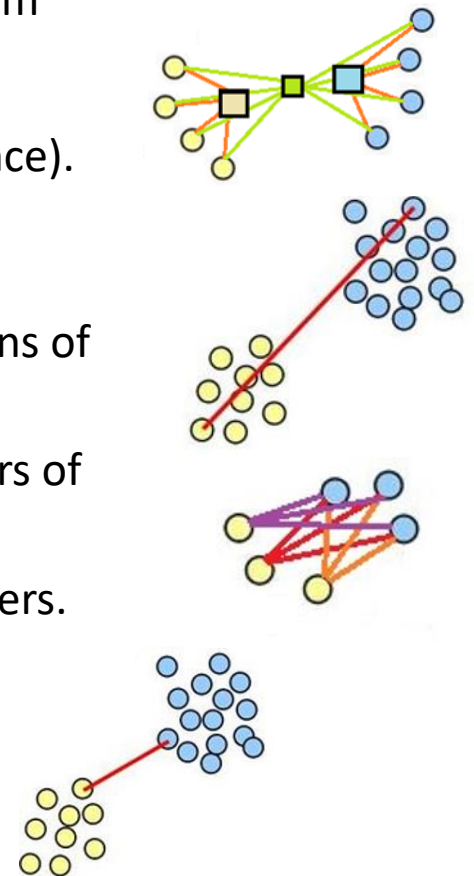
- Hierarchical clustering: family of clustering algorithms that build nested clusters by merging or splitting them successively. This hierarchy of clusters is represented as a tree (or dendrogram).
- **Agglomerative approach:** This is a "bottom-up" approach: each observation starts in its own cluster, and pairs of clusters are merged as one moves up the hierarchy.
- **Divisive approach:** This is a "top-down" approach: all observations start in one cluster, and splits are performed recursively as one moves down the hierarchy.



# Hierarchical clustering: important parameters

You choose:

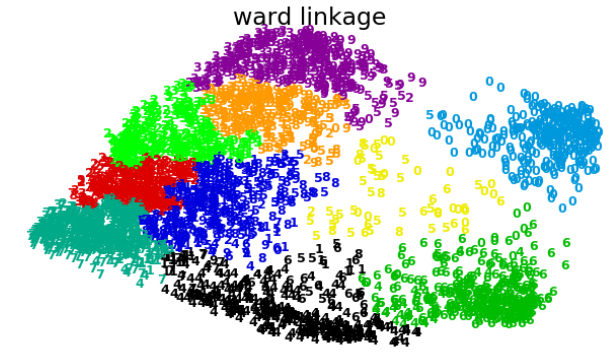
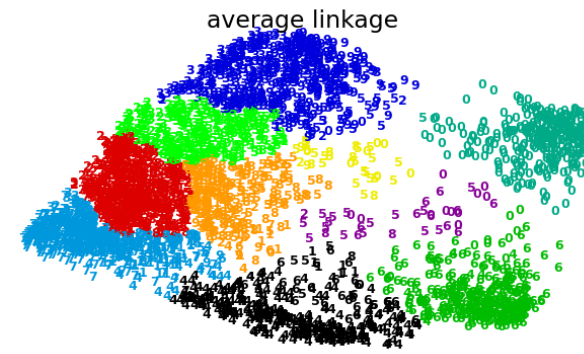
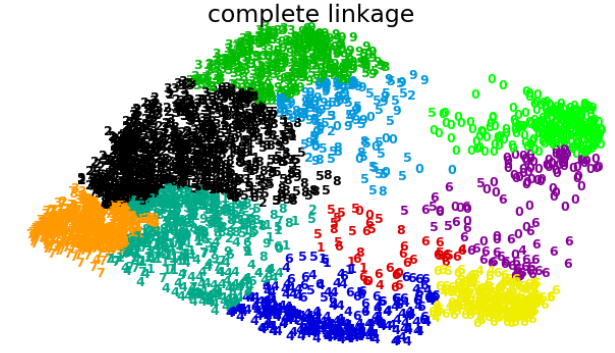
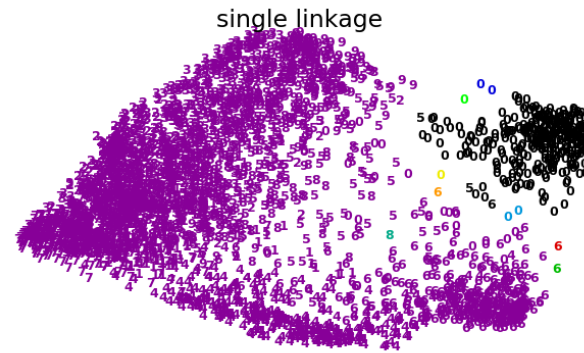
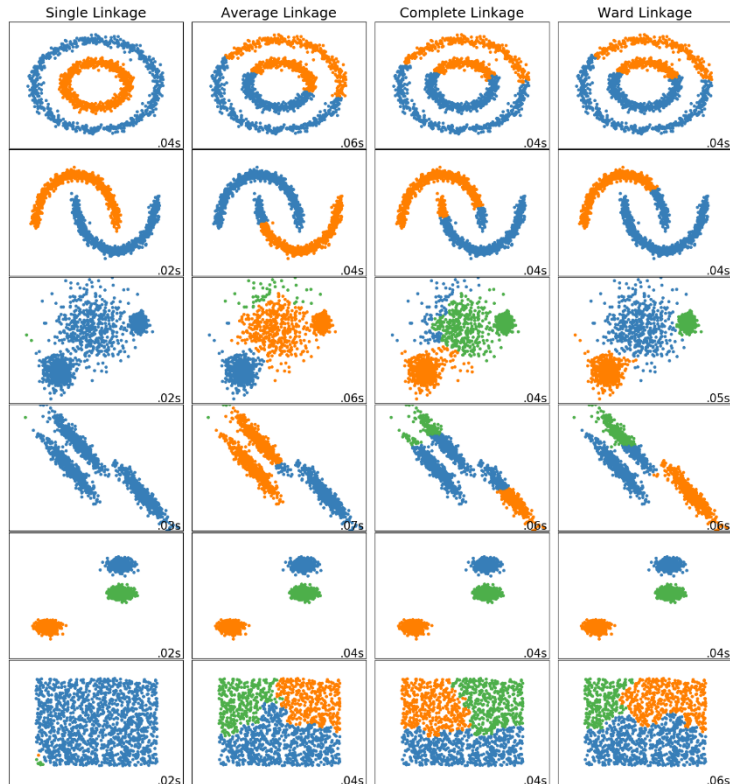
- **Distance metric** (between observations): Euclidean, Squared Euclidean, Manhattan, Maximum
- **Linkage criterium** (distance between sets of observations):
  - “**Ward**” minimizes the sum of squared differences within all clusters (within-cluster variance). It is a variance-minimizing approach and in this sense is similar to the k-means objective function but tackled with an agglomerative hierarchical approach.
  - “**Maximum**” or “complete linkage” minimizes the maximum distance between observations of pairs of clusters.
  - “**Average linkage**” minimizes the average of the distances between all observations of pairs of clusters.
  - “**Single linkage**” minimizes the distance between the closest observations of pairs of clusters.



<https://blog.tdwi.eu/hierarchical-clustering-in-python/>

# Hierarchical clustering: important parameters

- **Linkage criterium** (distance between sets of observations):



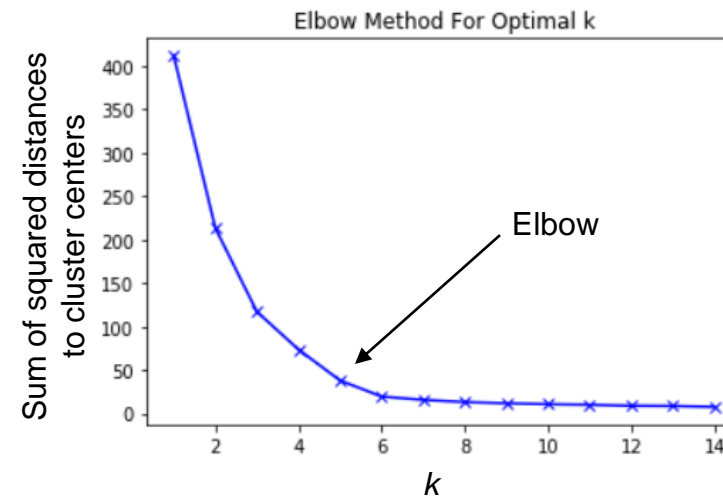
“Ward” gives the most regular cluster sizes

From <https://scikit-learn.org/stable/modules/clustering.html#hierarchical-clustering>  
[https://scikit-learn.org/stable/auto\\_examples/cluster/plot\\_digits\\_linkage.html](https://scikit-learn.org/stable/auto_examples/cluster/plot_digits_linkage.html)



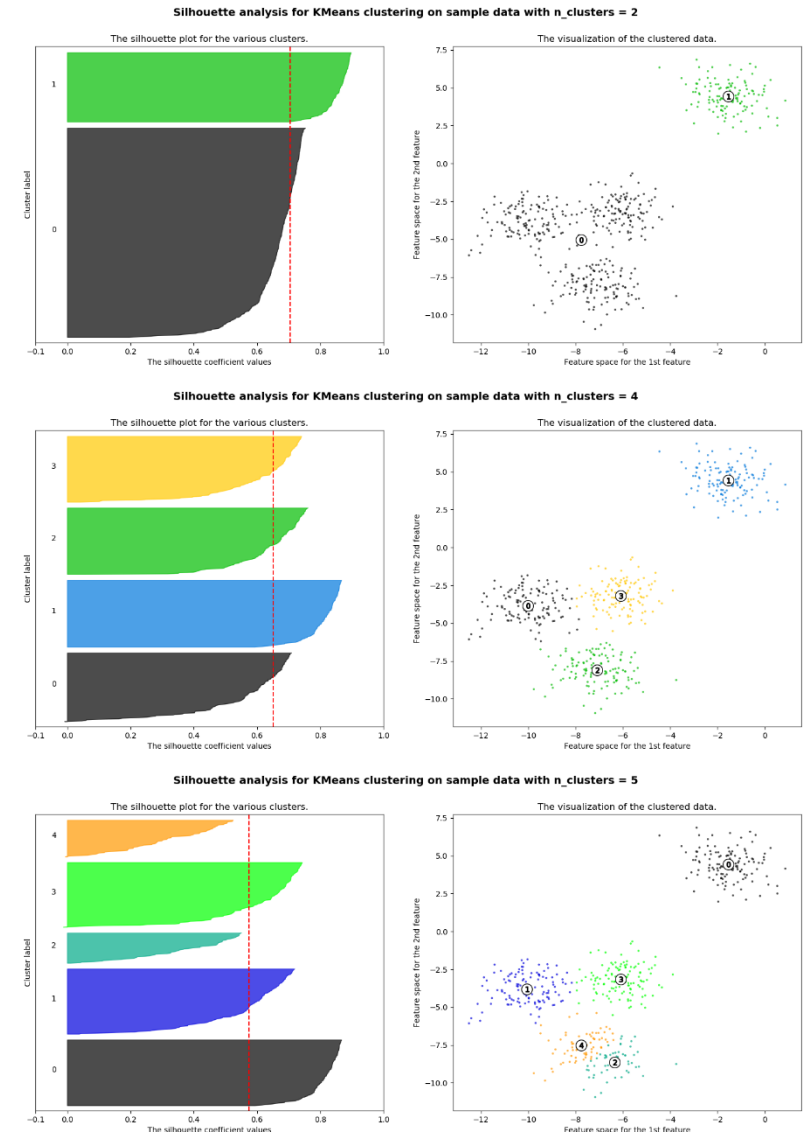
# How to choose the best number of clusters $k$ ?

- Elbow method (for sum of squared distances to cluster centers)
  - How find the “elbow”? By eye or <https://github.com/arvkevi/kneed>
- Silhouette analysis
- BIC and AIC



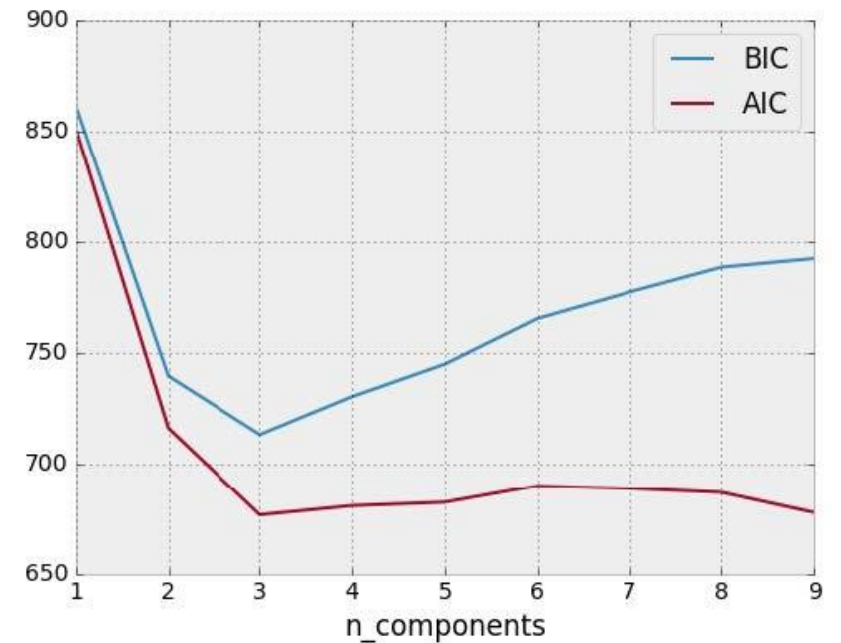
# How to choose the best number of clusters k?

- Elbow method
- Silhouette analysis
  - Plots silhouette score: a measure of how close each point in one cluster is to points in the neighboring clusters, in  $[-1, 1]$ .
  - Values near +1 indicate that the sample is far away from the neighboring clusters. A value of 0 indicates that the sample is on very close to the decision boundary between two neighboring clusters and negative values indicate that those samples might have been assigned to the wrong cluster.



# How to choose the best number of clusters k?

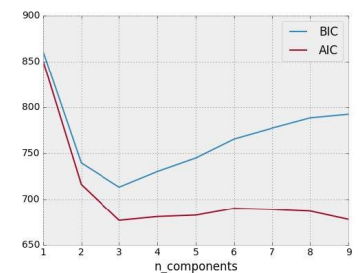
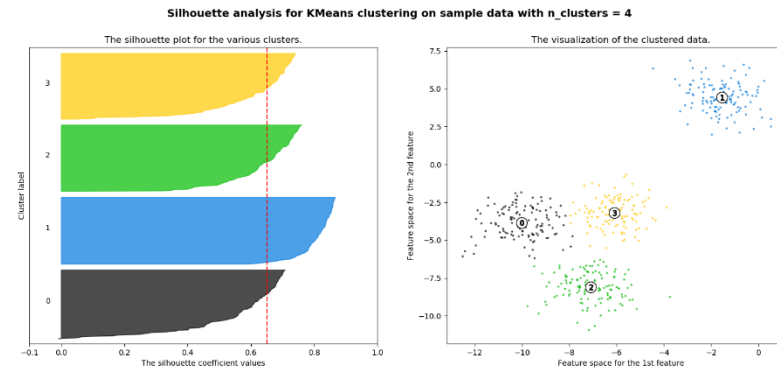
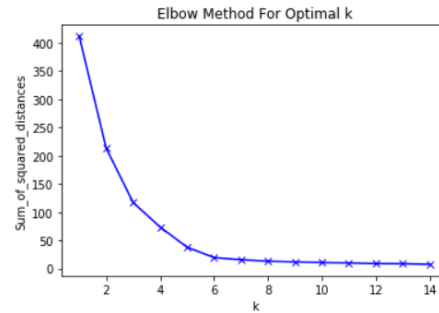
- Elbow method
- Silhouette analysis
- Akaike information criterion (AIC) or Bayesian information criterion (BIC)
  - The BIC generally penalizes free parameters more strongly than the AIC



From <https://sites.northwestern.edu/msia/2016/12/08/k-means-shouldnt-be-our-only-choice/>

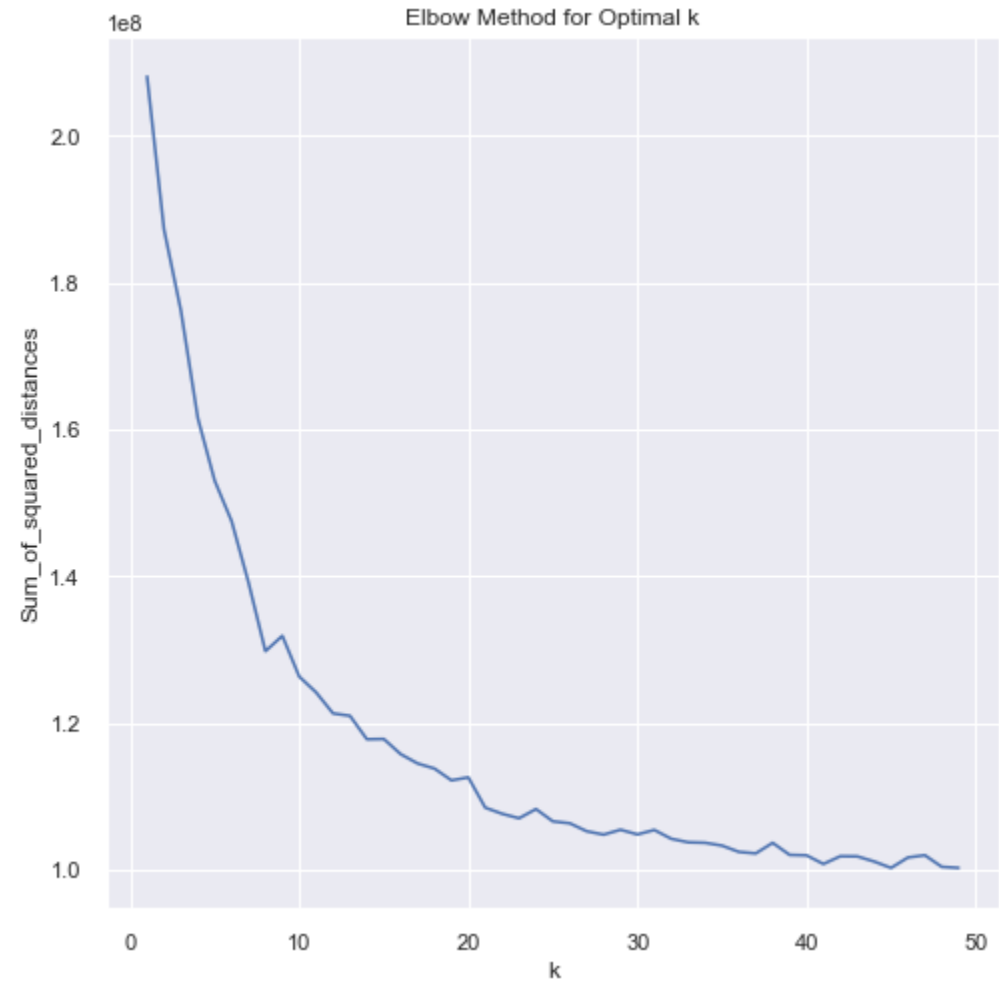
# How to choose the best number of clusters k?

- Elbow method
- Silhouette analysis
- BIC and AIC



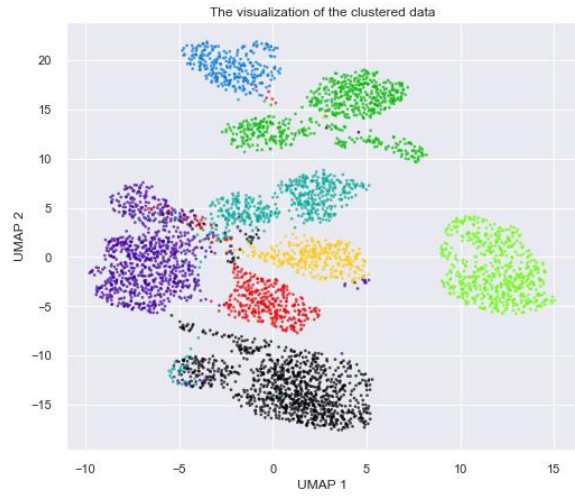
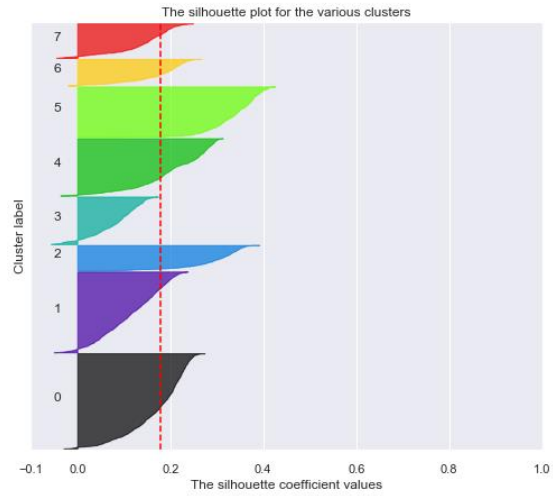
Let's go to our hands on exercise to see how it works!

Which value of k would you choose?  
(Mini-batch k-means clustering method)



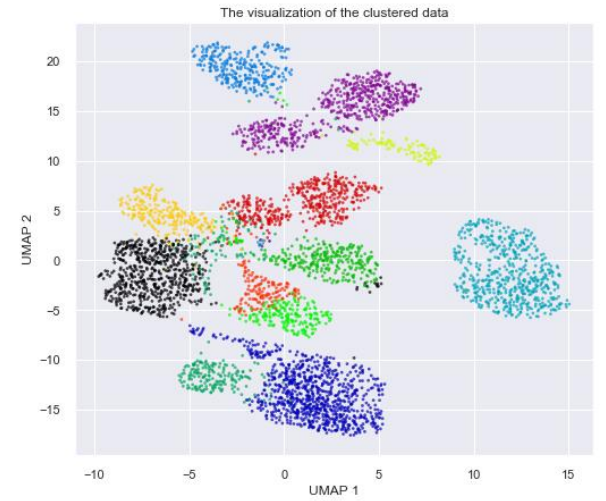
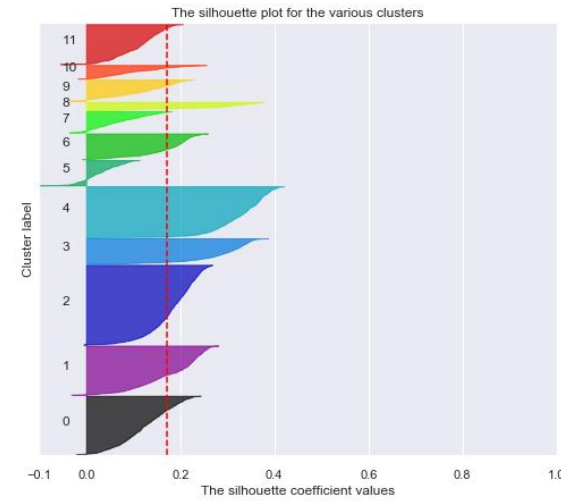
# Which value of k would you choose?

Silhouette analysis for KMeans clustering on sample data with n\_clusters = 8

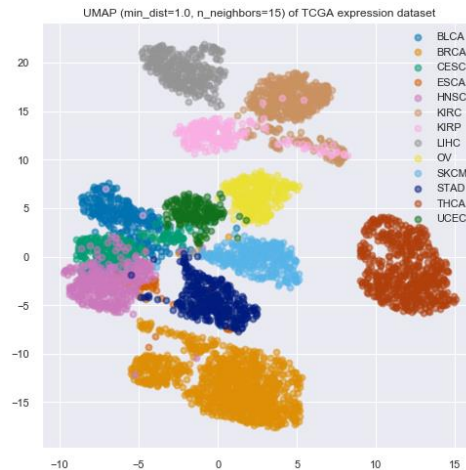


$k = 8$

Silhouette analysis for KMeans clustering on sample data with n\_clusters = 12



$k = 12$

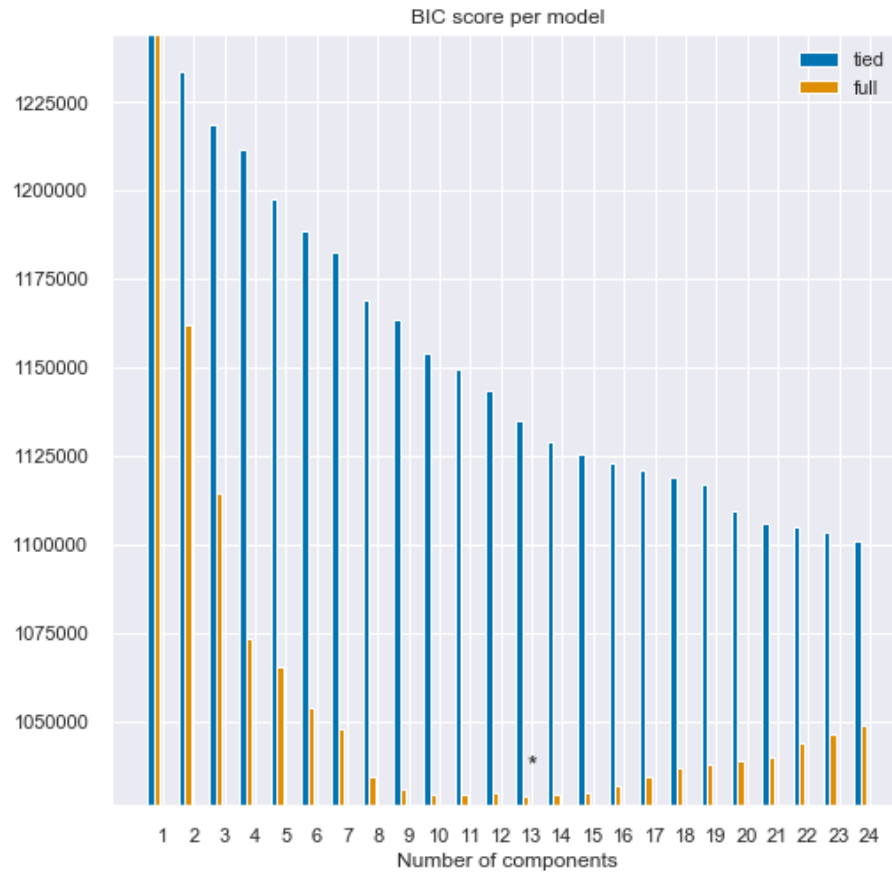


- BLCA Bladder
- BRCA Breast
- CESC Cervical squamous cell
- ESCA Esophageal
- HNSC Head and Neck squamous cell
- KIRC Kidney clear cell
- KIRP Kidney papillary
- LIHC Liver
- OV Ovarian
- SKCM Skin Cutaneous Melanoma
- STAD Stomach
- THCA Thyroid
- UCEC Uterine Corpus Endometrial

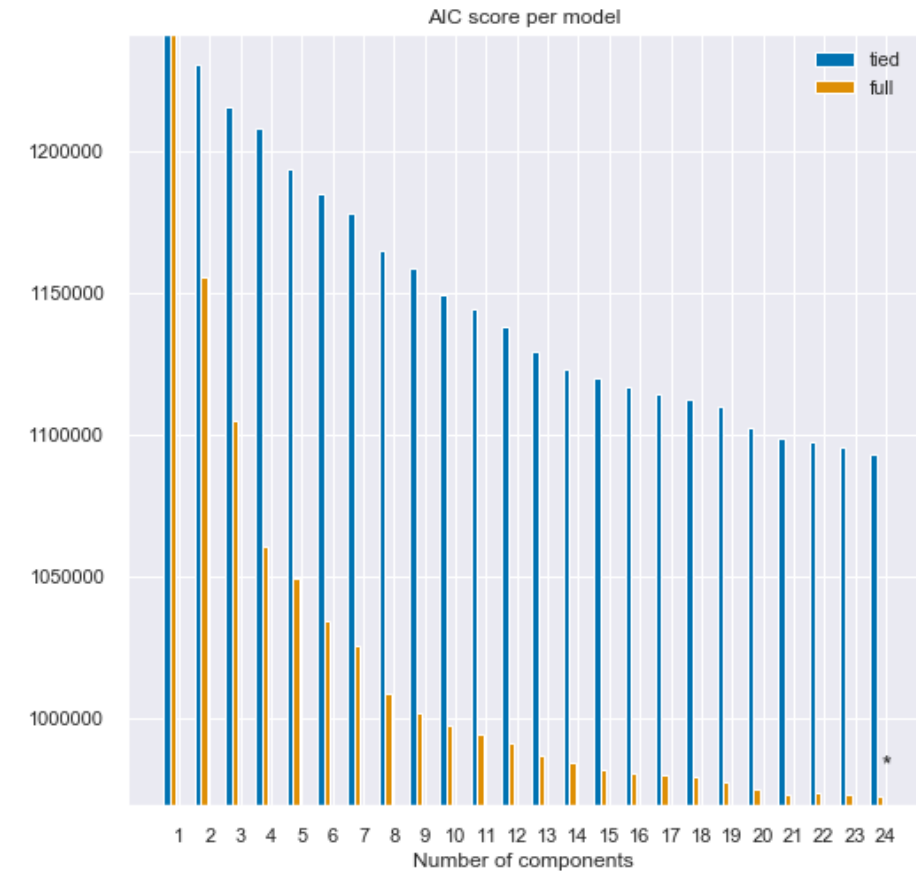
True types

# Which value of k would you choose? (Gaussian mixture model)

## BIC



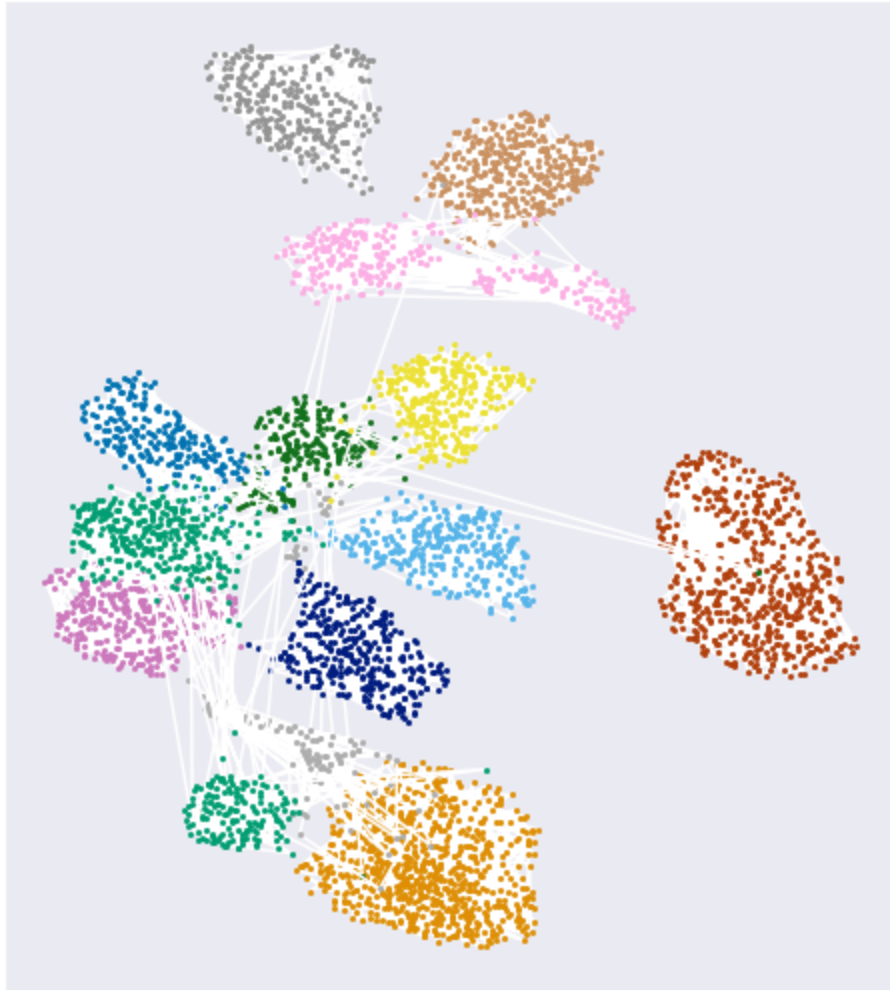
## AIC



# Which value of k would you choose? (Gaussian mixture model)

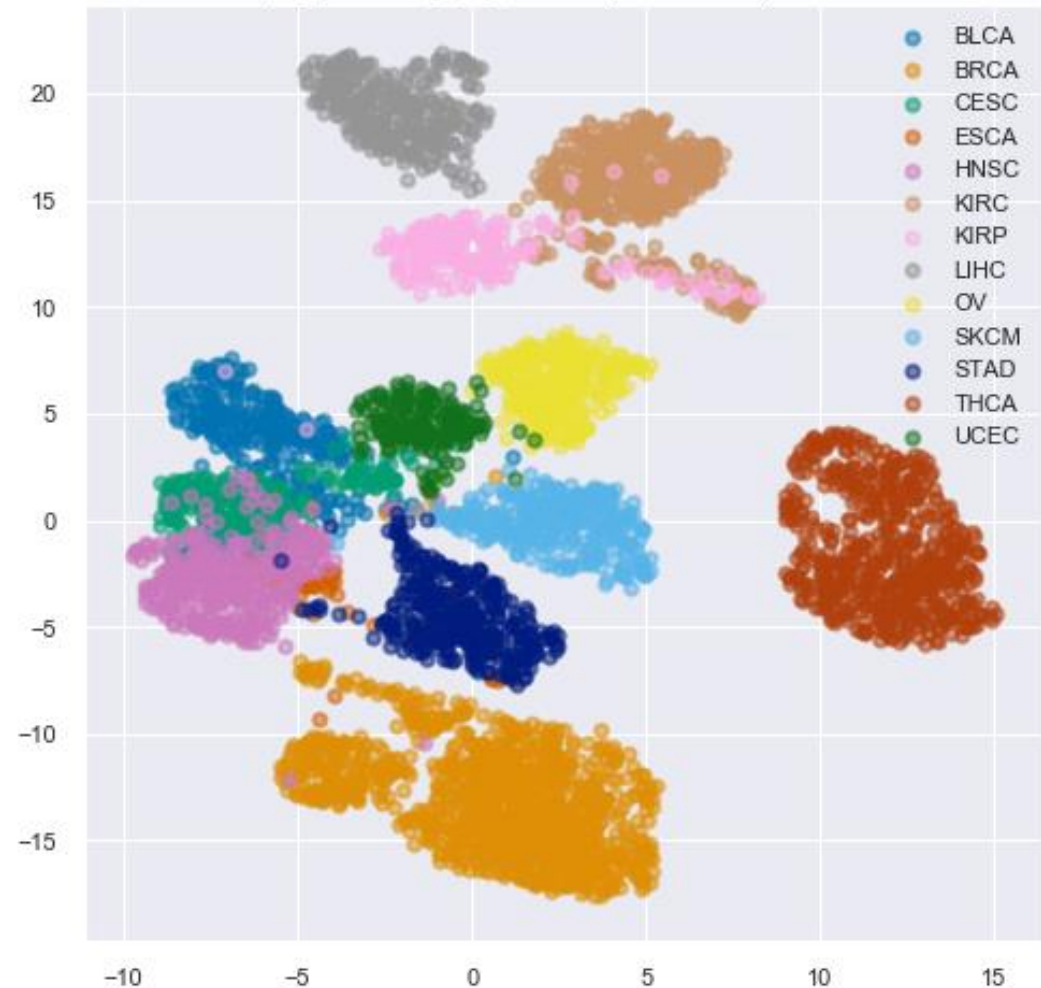
Best solution according to BIC:  $k=13$

Gaussian mixture models for clustering TCGA expression dataset



True labels

UMAP (min\_dist=1.0, n\_neighbors=15) of TCGA expression dataset





# NMI and ARI allows to see which clustering model is the best on our data set

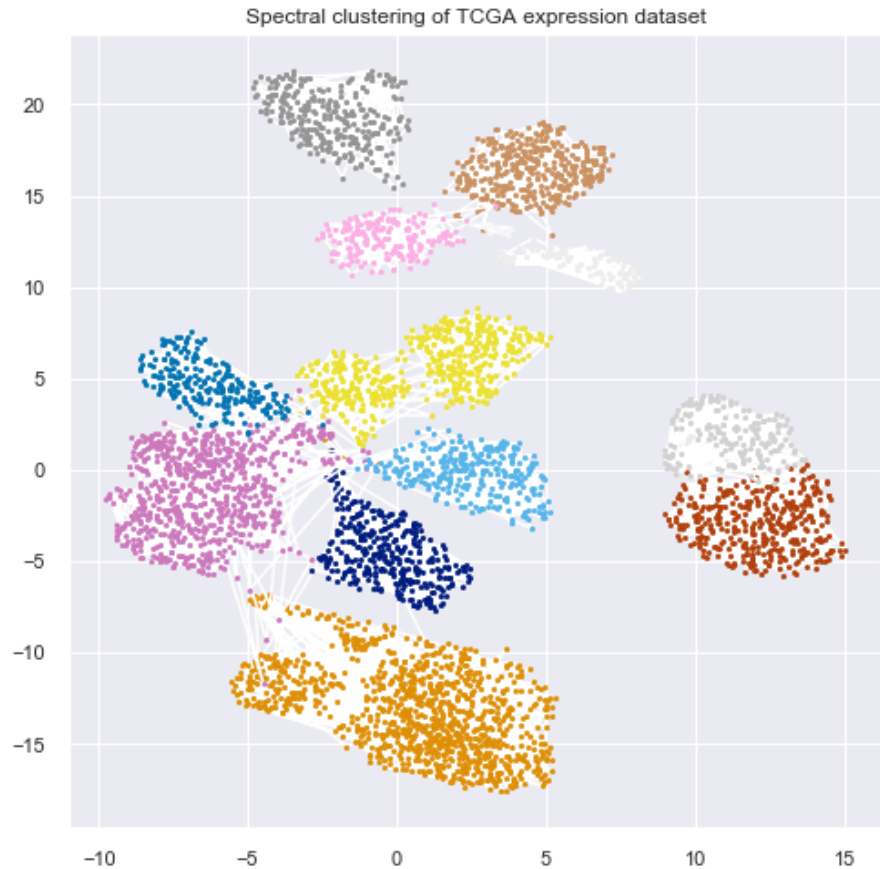
When true labels are available (rarely the case though):

NMI = Normalized Mutual Information }  
ARI = Adjusted rand index } measure mutual dependence between the true and predicted labels  
(can be also used as a measure of similarity between two clusterings)

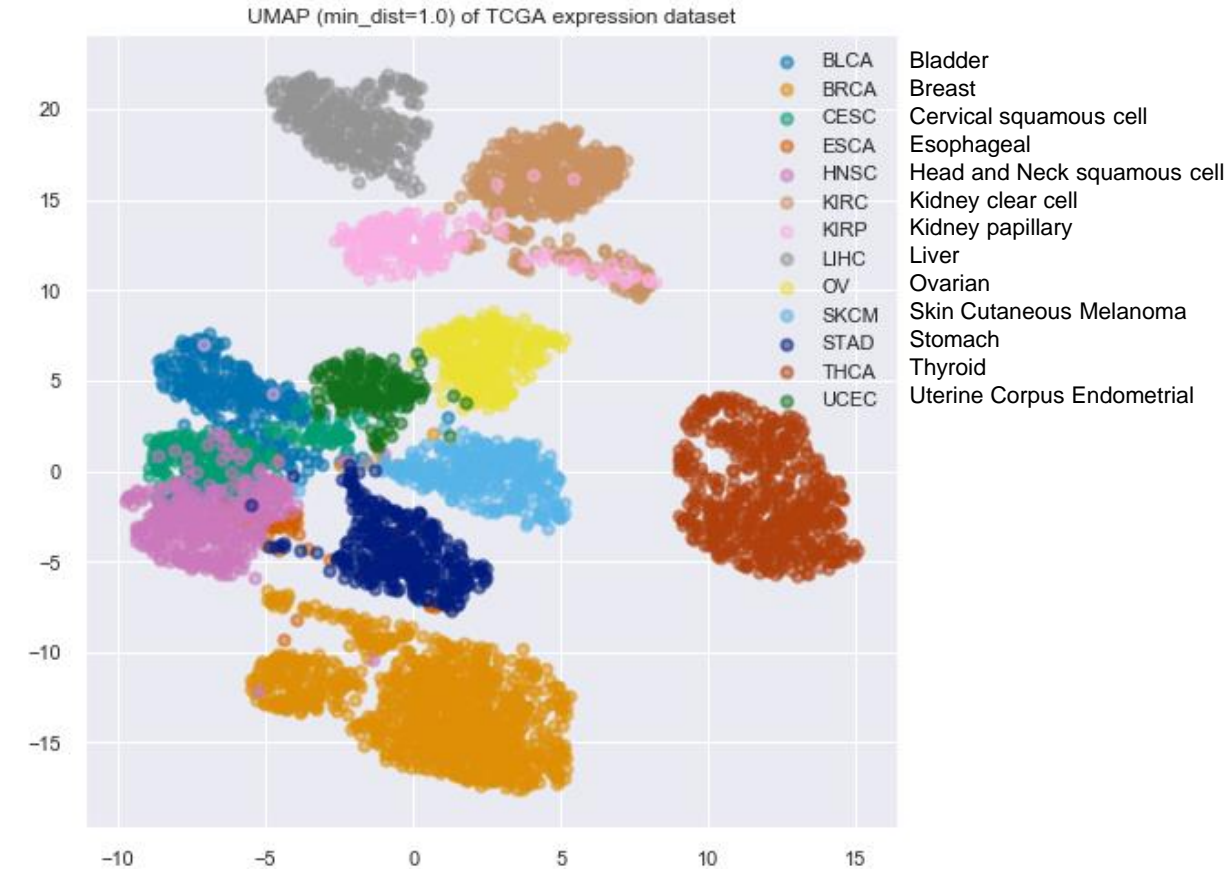
Method	<i>k</i>	ARI	NMI
Mini-batch k-means	8	0.7479	0.8102
Mini-batch k-means	19	0.7426	0.8179
Gaussian mixture model	13	0.7600	0.8479
<b>Spectral clustering</b>	<b>13</b>	<b>0.8148</b>	<b>0.8671</b>
Hierarchical clustering (linkage='ward')	13	0.7394	0.8379

# Spectral clustering result (k=13)

## Best solution according to NMI



## True labels



# Take home message: clustering

- The choice of the clustering method should be advised by data structure
  - Visualize the data first
  - E.g, ellipsoids => GMM; sophisticated connected components => structural clustering
- Choosing the number of clusters can be done using:
  - Elbow method
  - Silhouette method
  - BIC or AIC
- Different methods for the estimation of the number of clusters provide different results
  - E.g., BIC is more conservative than AIC

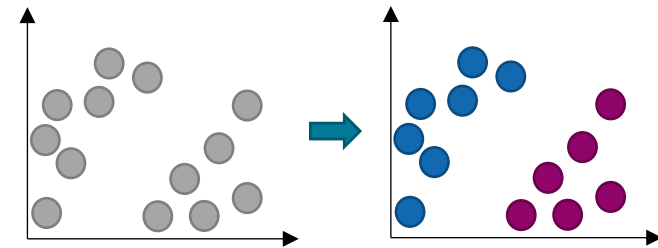
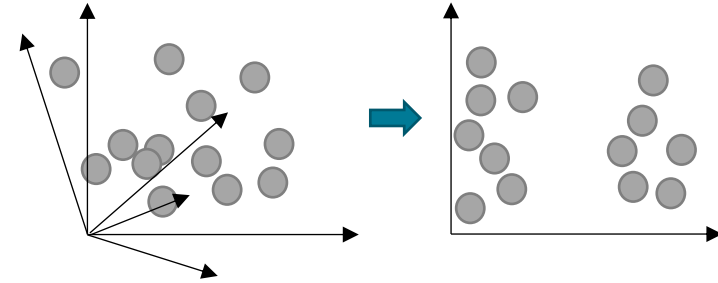
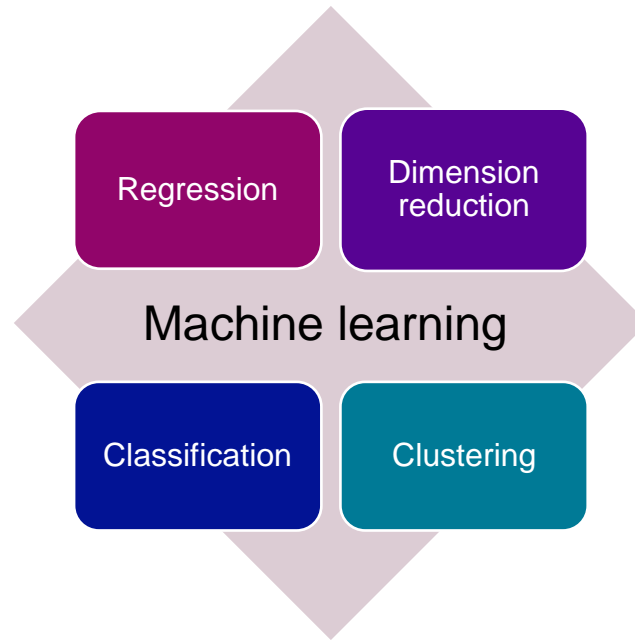
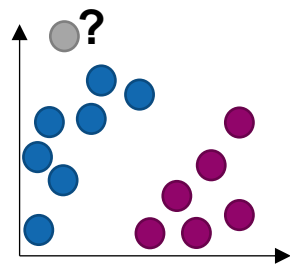
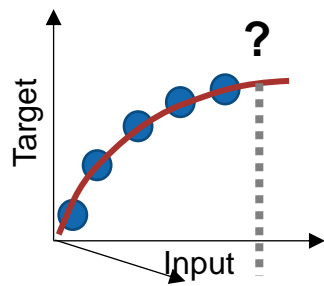
# What we did not cover today

- Graph-based clustering methods (Leiden and Louvain algorithms, commonly used on scRNA-seq data)
- Topic models/LDA
- Autoencoder-based methods for dimensionality reduction
- Integration of different data types (e.g., clusters of clusters)

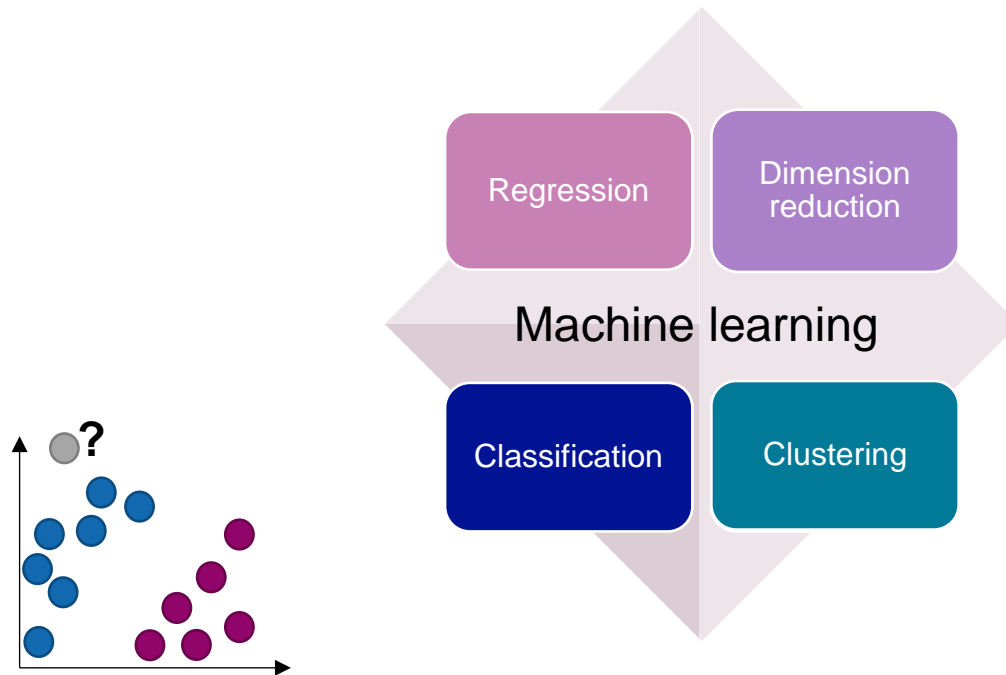
# Take home message

- Dimensionality reduction can be used as a step prior to clustering
- One usually tries different dimensionality reduction techniques to choose the one that fits the expectations
- The choice of clustering method should match the data structure

# Map of classical machine learning methods

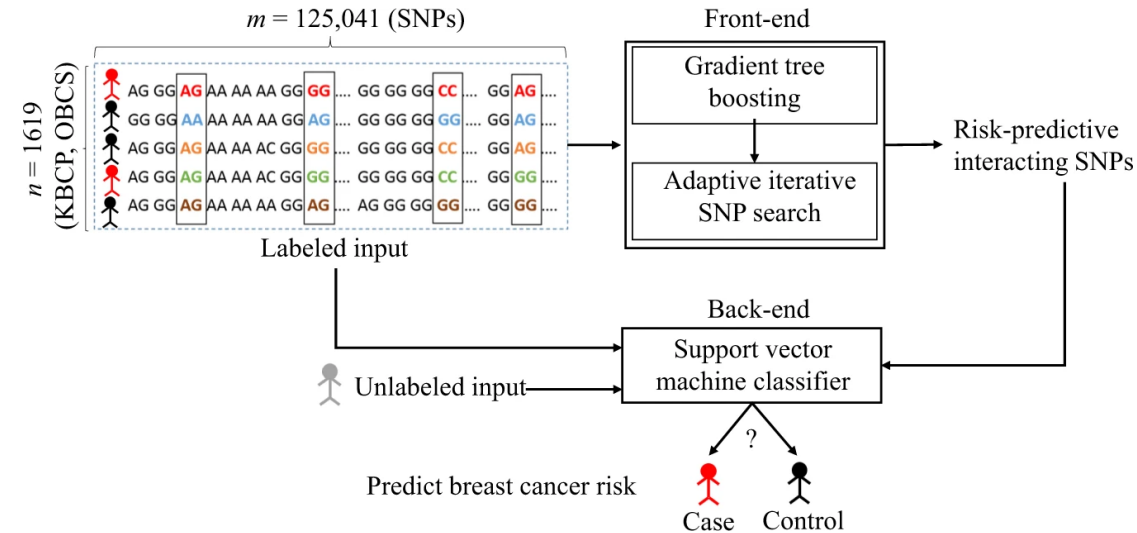


# Map of classical machine learning methods



# Classification: Biological examples

- Prediction of risk groups
  - **Machine learning identifies interacting genetic variants contributing to breast cancer risk [...]** Behravan et al., *Sci Rep.* 2018
- Primary diagnosis
  - **DNA methylation-based classification of central nervous system tumours.** Capper et al. *Nature* 2018

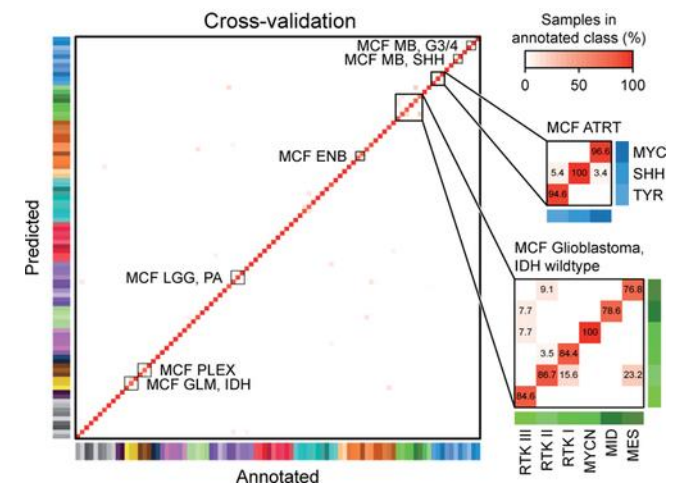
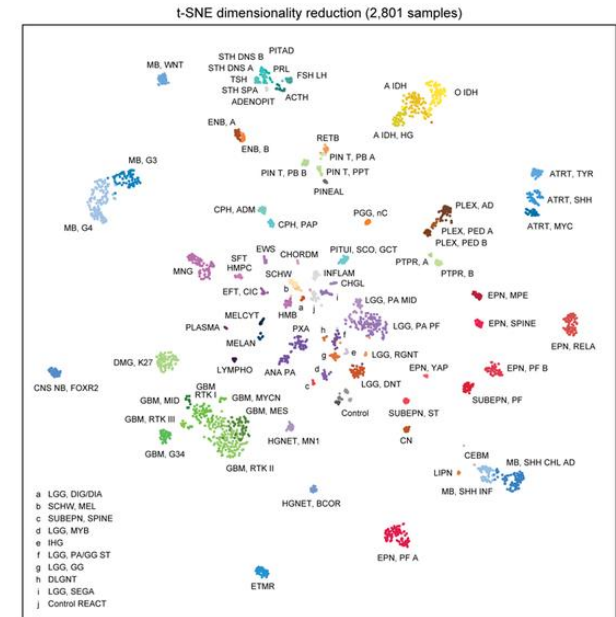


Proposed breast cancer risk prediction approach using identified risk-predictive interacting SNPs



# Classification: Biological examples

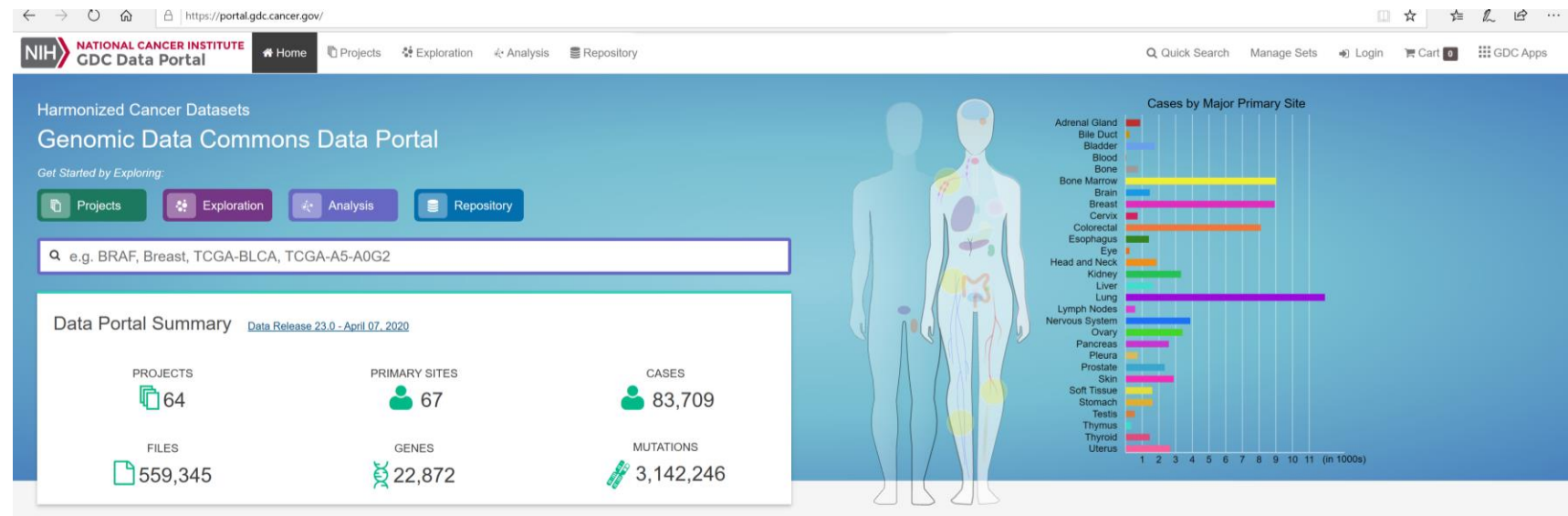
- Prediction of risk groups
  - Machine learning identifies interacting genetic variants contributing to breast cancer risk [...] Behravan et al., *Sci Rep.* 2018
- Primary diagnosis
  - DNA methylation-based classification of central nervous system tumours. Capper et al. *Nature* 2018



Hands-on:

1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression

- **Input:** The Cancer Genome Atlas (TCGA) mRNA expression data

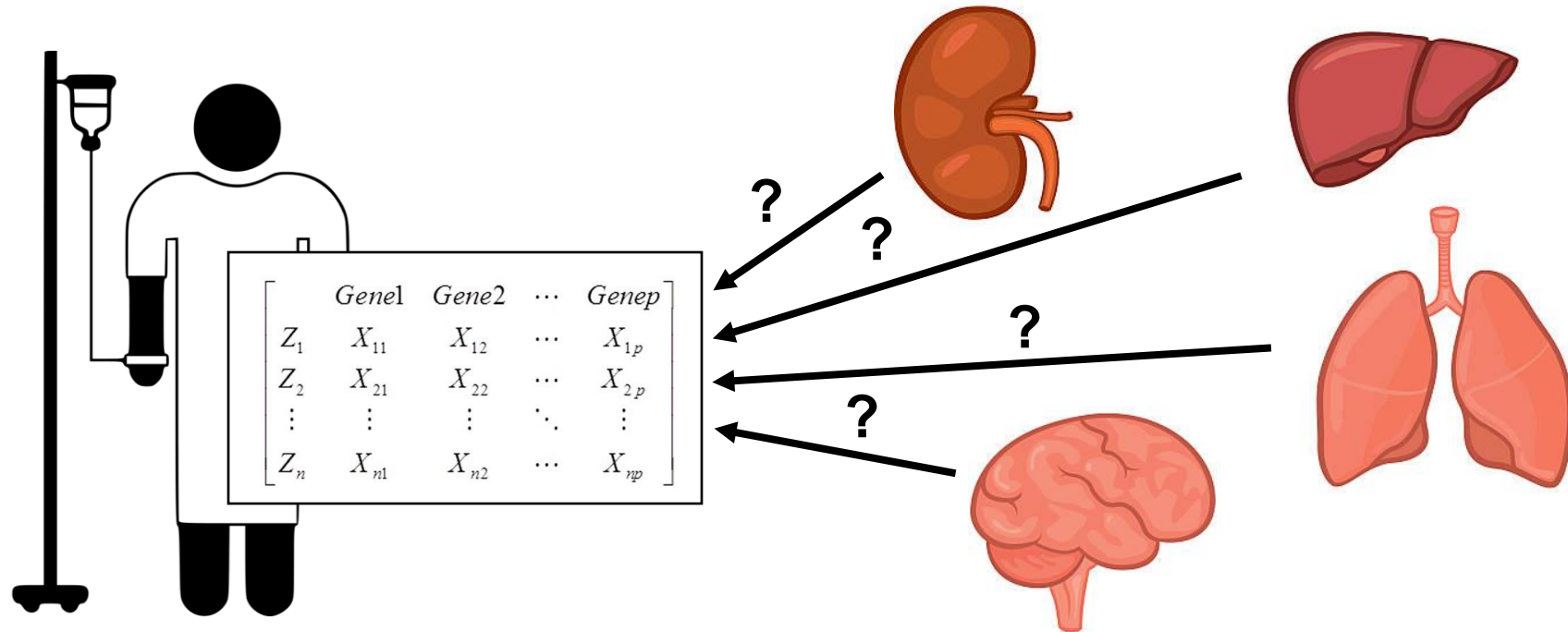


Hands-on:

1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression

- **TASK 1:**

Given mRNA expression, predict cancer type



Hands-on:

1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression

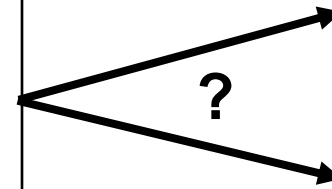
- **TASK 2:**

Given mRNA expression (and clinical data: stage, age), stratify patients according to good and bad prognosis



	<i>Gene1</i>	<i>Gene2</i>	...	<i>Gene<sub>p</sub></i>
$Z_1$	$X_{11}$	$X_{12}$	...	$X_{1p}$
$Z_2$	$X_{21}$	$X_{22}$	...	$X_{2p}$
$\vdots$	$\vdots$	$\vdots$	$\ddots$	$\vdots$
$Z_n$	$X_{n1}$	$X_{n2}$	...	$X_{np}$

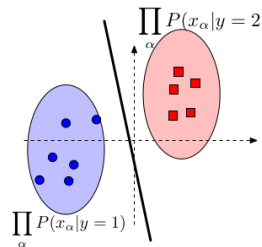
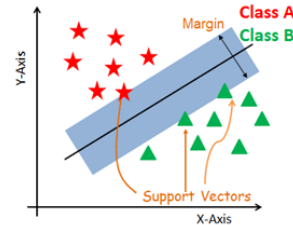
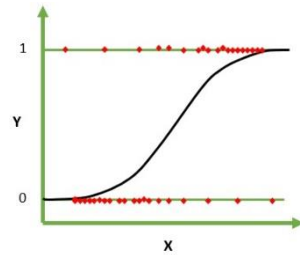
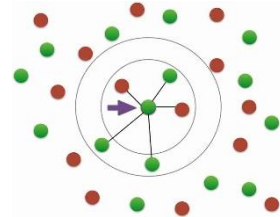
+ clinical stage + age



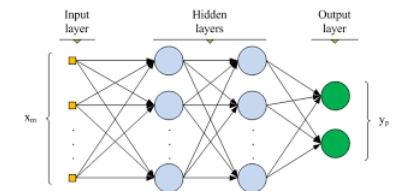
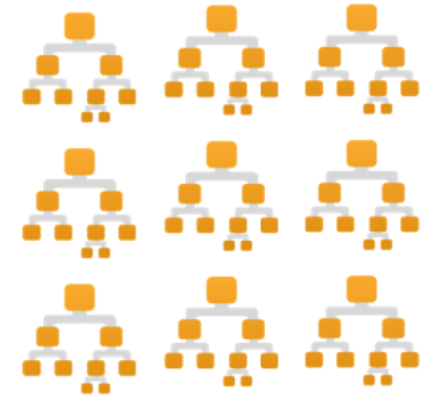
Aggressive treatment to be applied

# Standard approaches for classification

- k Nearest Neighbors (k-NN)
- Logistic Regression
- Logistic Regression with L1+L2 (Elastic Net) penalty
- Support Vector Machines (SVM)
- Naive Bayes (Gaussian)



- Random Forest
- AdaBoost
- Gradient Tree Boosting (gradient boosting machine, GBM)
- Multi-layer perceptron (MLP)



# Hands-on:

1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression

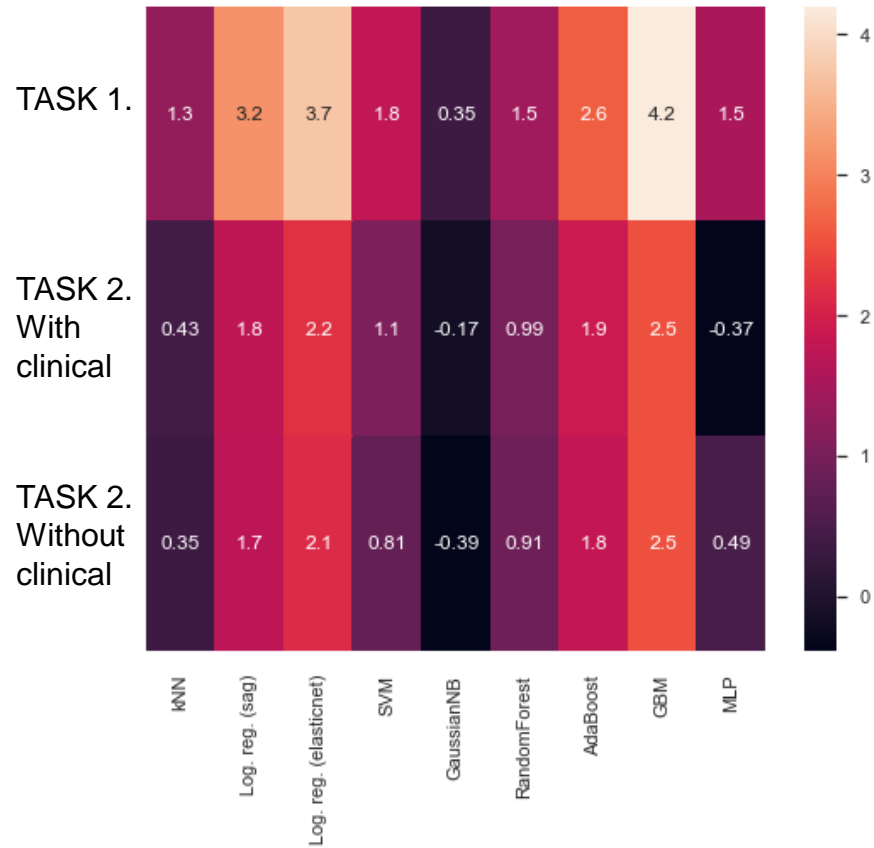
$$\text{Accuracy} = \frac{\text{TP} + \text{TN}}{\text{TP} + \text{TN} + \text{FP} + \text{FN}}$$

Fraction predicted correctly

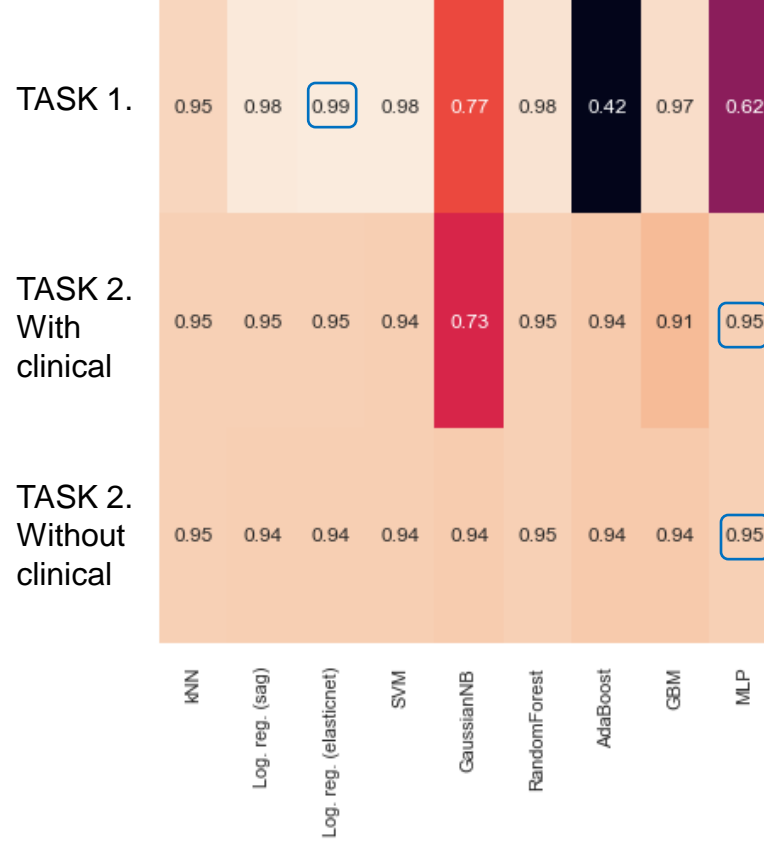
F1 score: harmonic mean of recall and precision

$$F1 = \frac{2}{\frac{1}{\text{precision}} + \frac{1}{\text{recall}}} = \frac{2 * (\text{precision} * \text{recall})}{\text{precision} + \text{recall}}$$

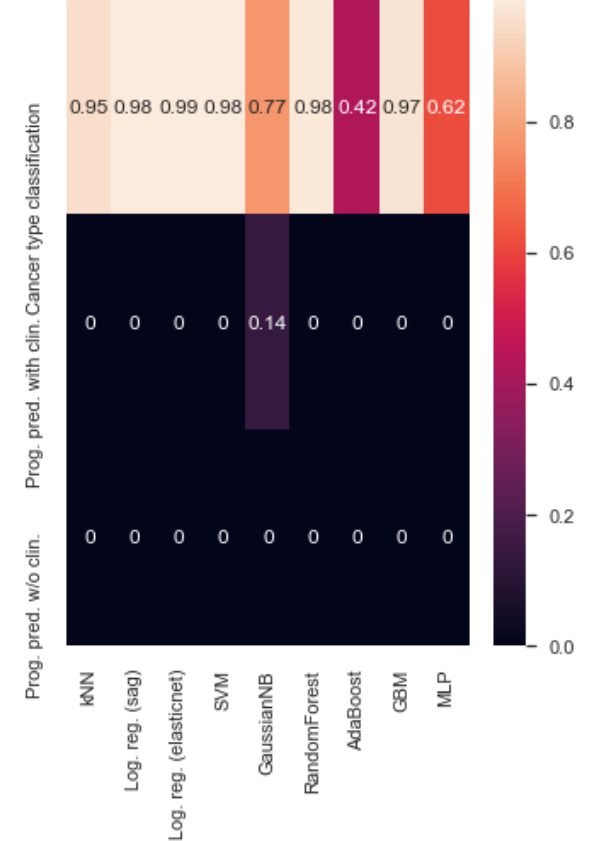
Time to train the model  
Log10 seconds



Model accuracy on the test set



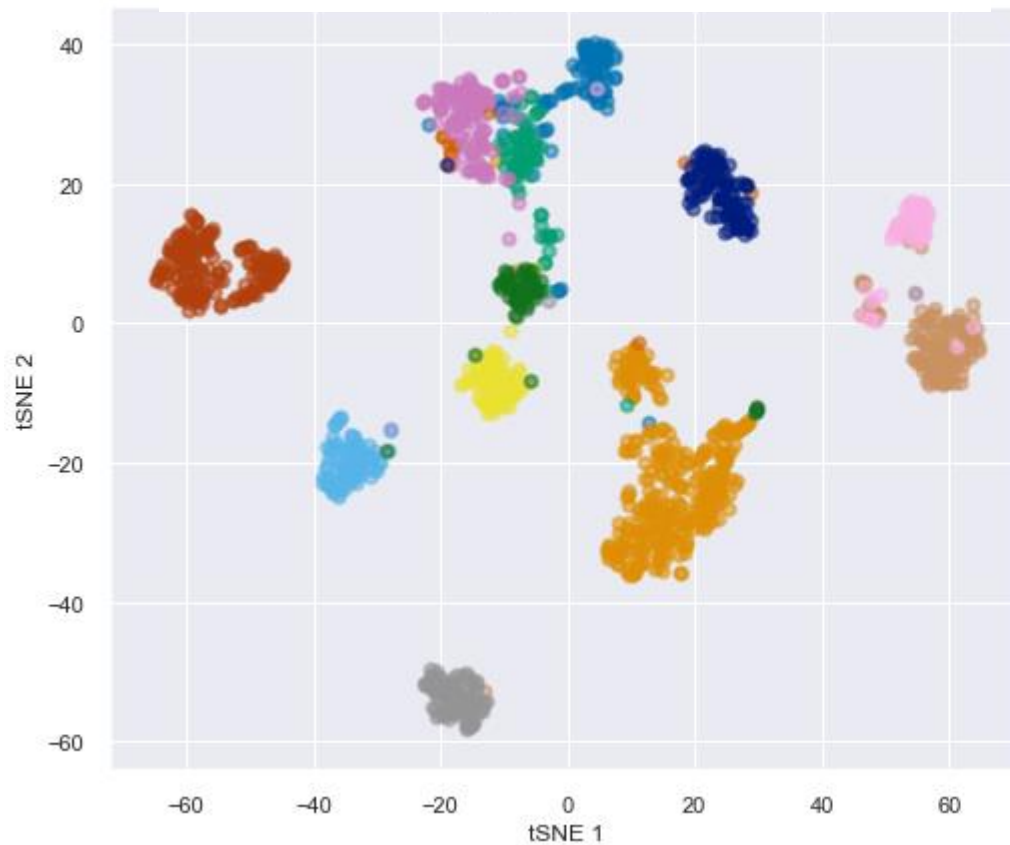
F1-score on the test set



Hands-on:

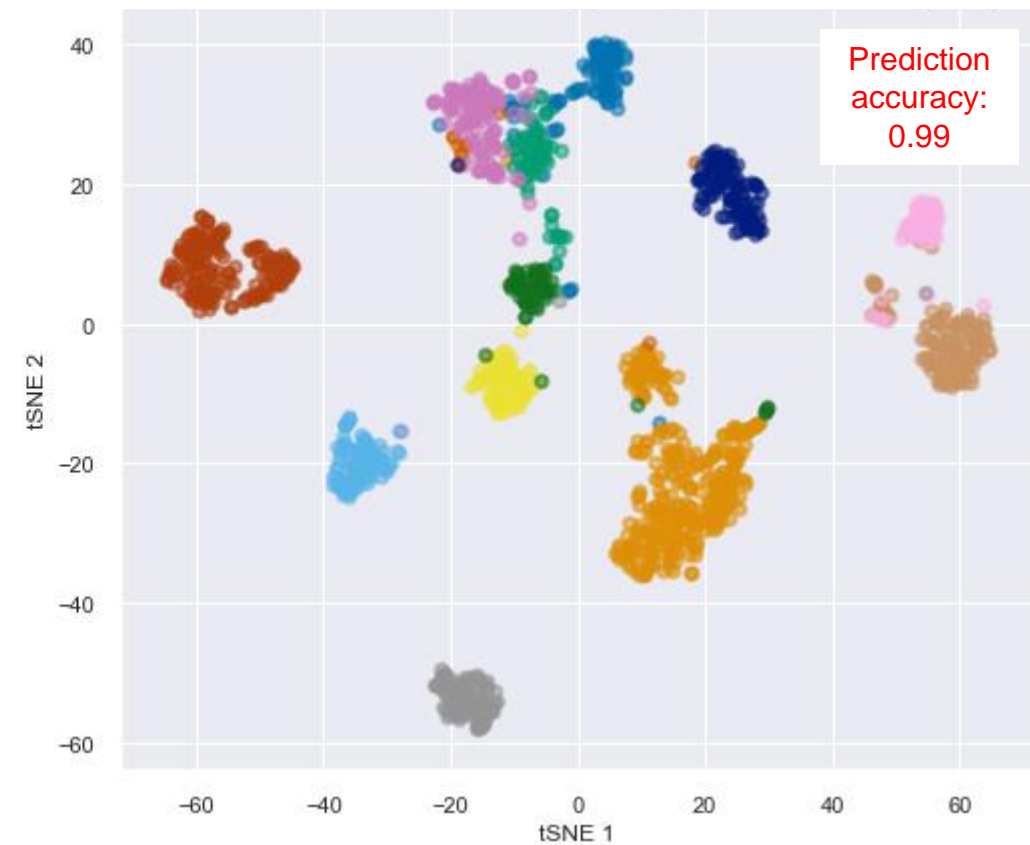
1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression

True values, tumor classification task



Validation set (colors correspond to true cancer types)

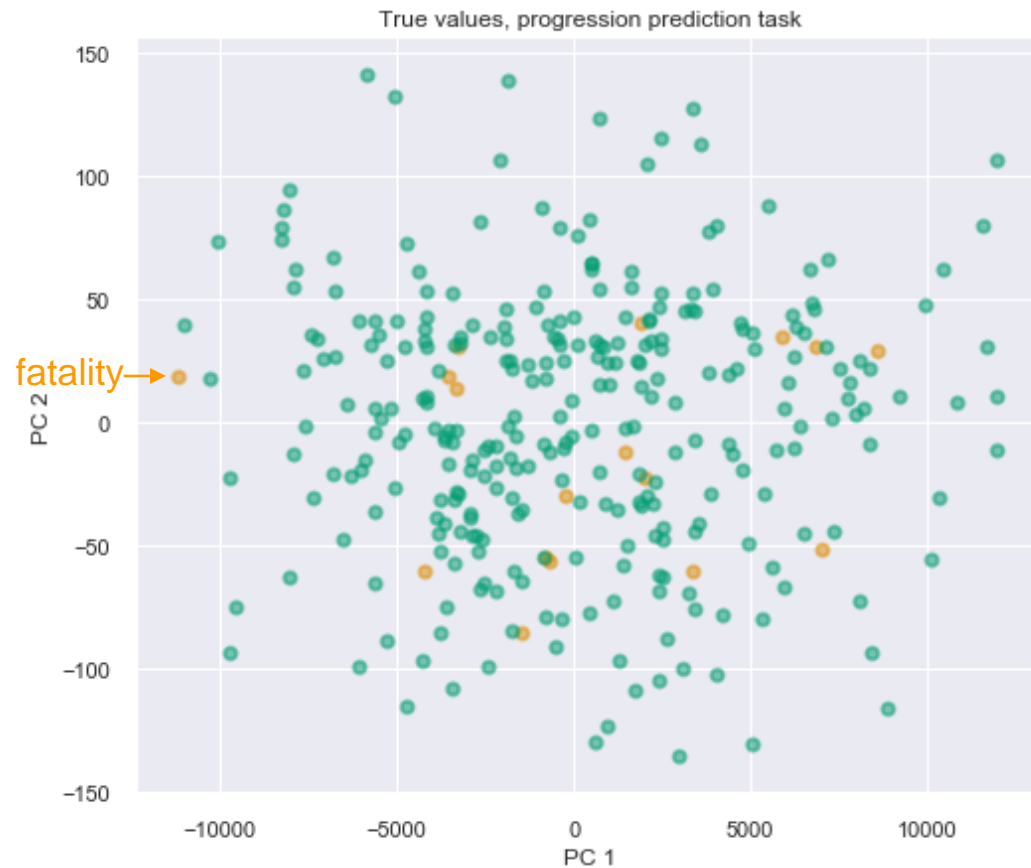
Best solution: Elastic net logistic regression



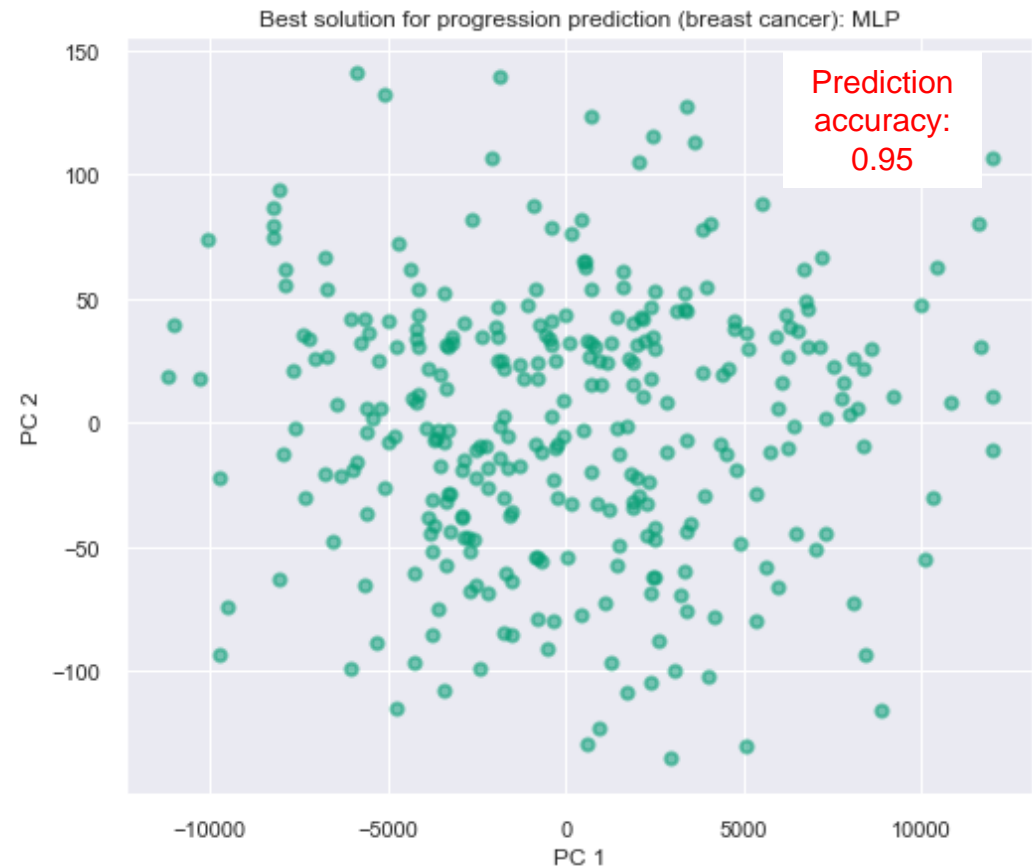
Elastic net on the validation set (colors correspond to predictions)

Hands-on:

1. Primary cancer diagnosis from gene expression
2. Breast cancer patients' stratification based on gene expression



Validation set (true colors)



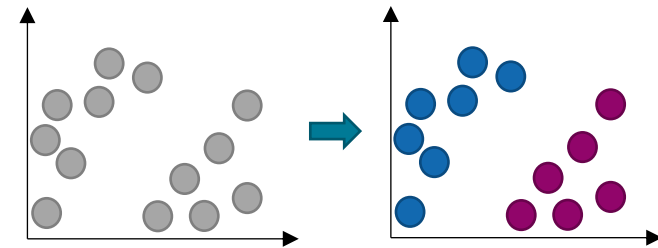
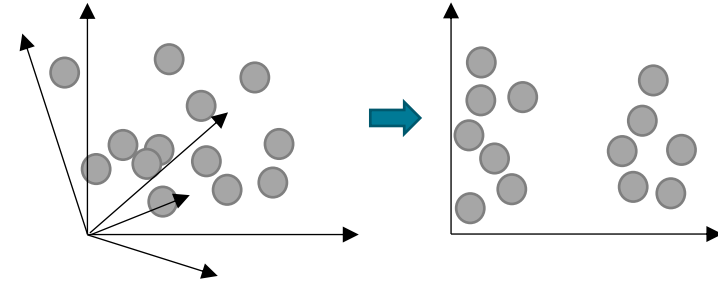
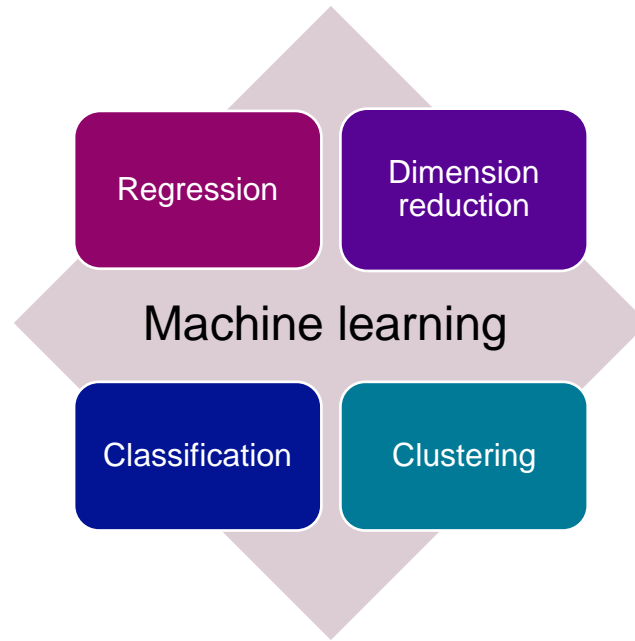
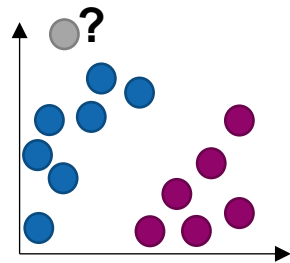
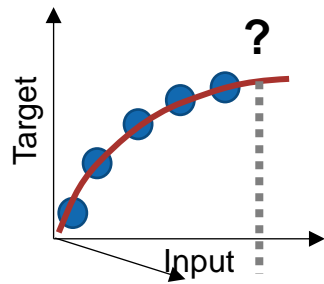
multi-layer perceptron on the validation set



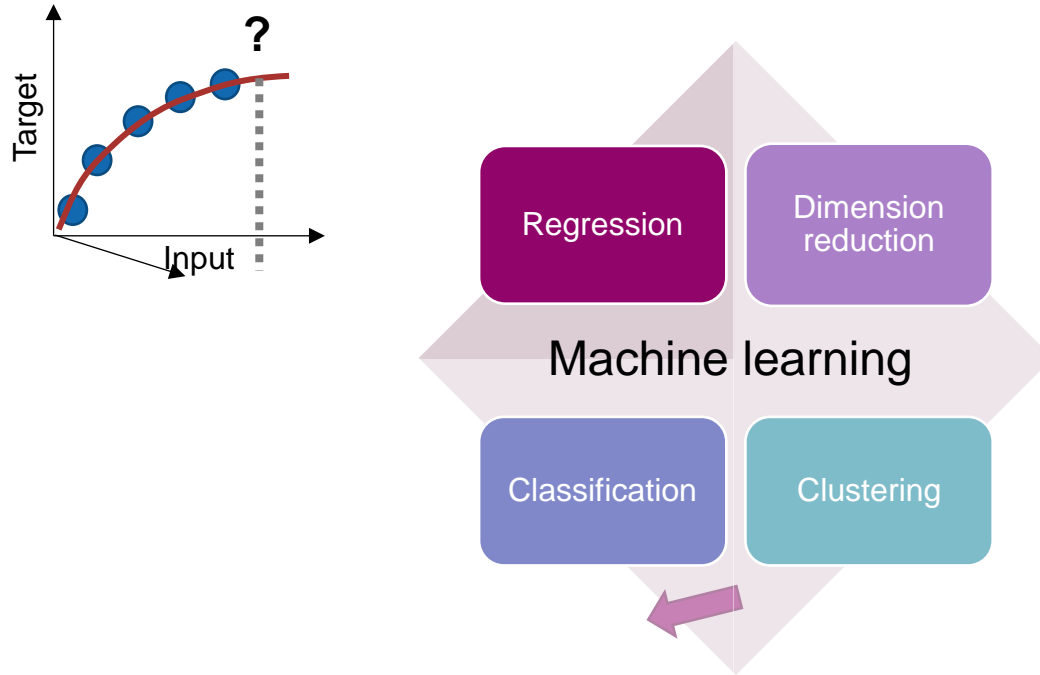
# Take home message: Classification

- Linear and non-linear models can provide similar prediction accuracy (TASK1)
- Classification on imbalanced groups with low information content may fail (TASK2)
  - Study your data first
    - Check data summary
    - Visualize your data
  - Use the right evaluation metrics (e.g., precision and recall)
  - Consider redesigning your task

# Map of classical machine learning methods



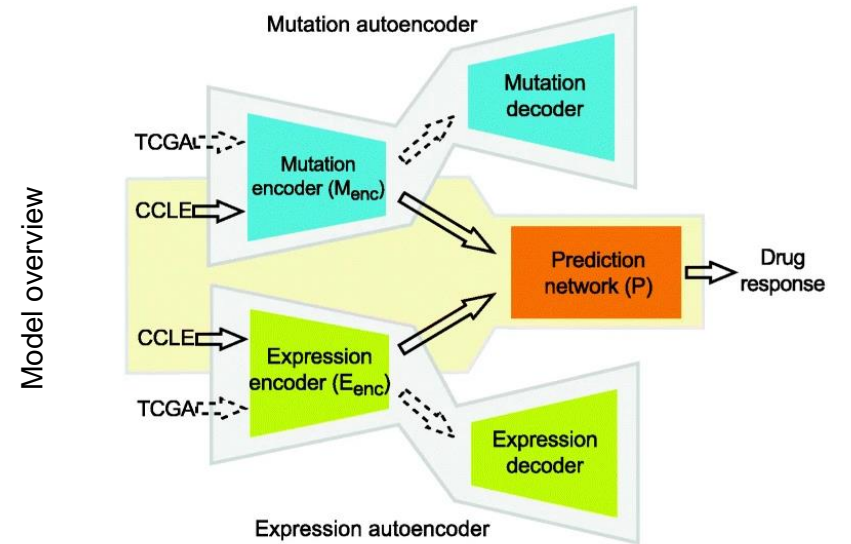
# Map of classical machine learning methods



# Regression: Biological examples

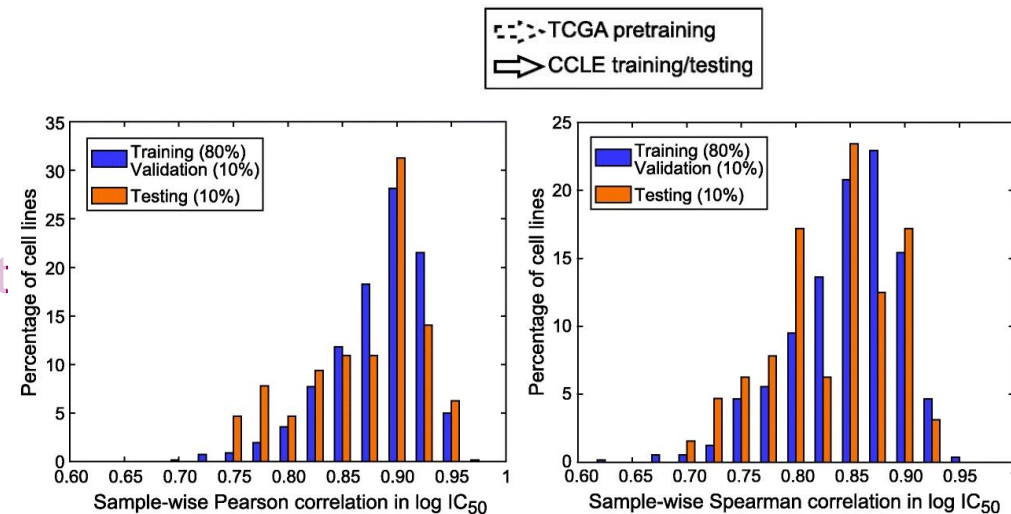
- Prediction of treatment efficiency / drug response

- **Predicting drug response of tumors from integrated genomic profiles by deep neural networks.** Chiu et al., *BMC Med. Genomic*, 2019



- Prediction of molecular/cellular properties (e.g., protein-DNA binding affinities)

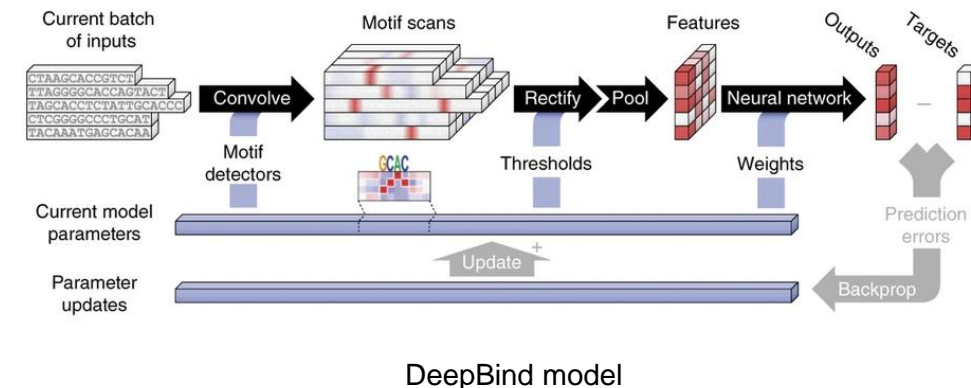
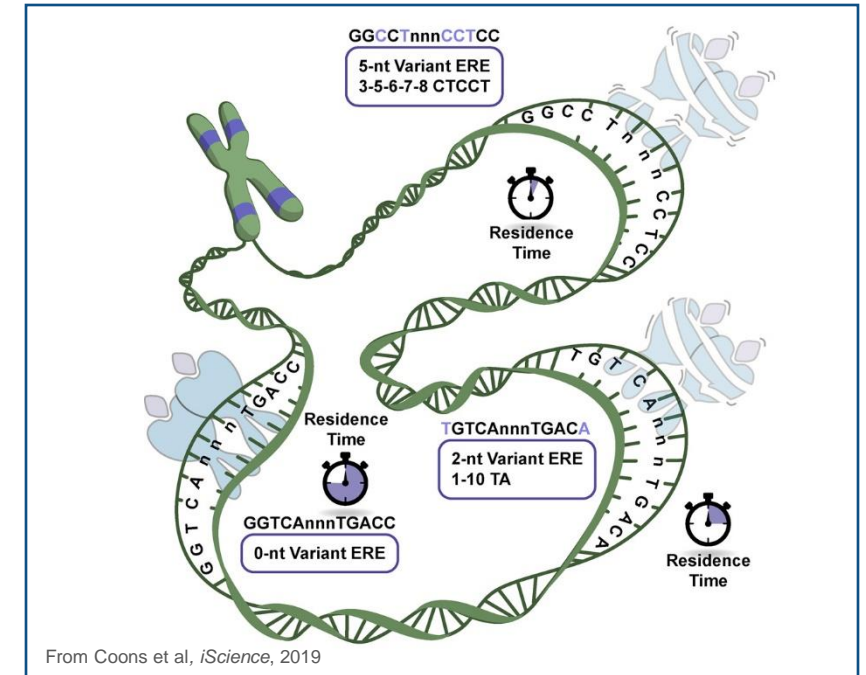
- **Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning.** Alipanahi et al., *Nature Biotech.* 2015



Sample-wise Pearson and Spearman correlation between imputed and predicted  $IC_{50}$  data of CCGE samples

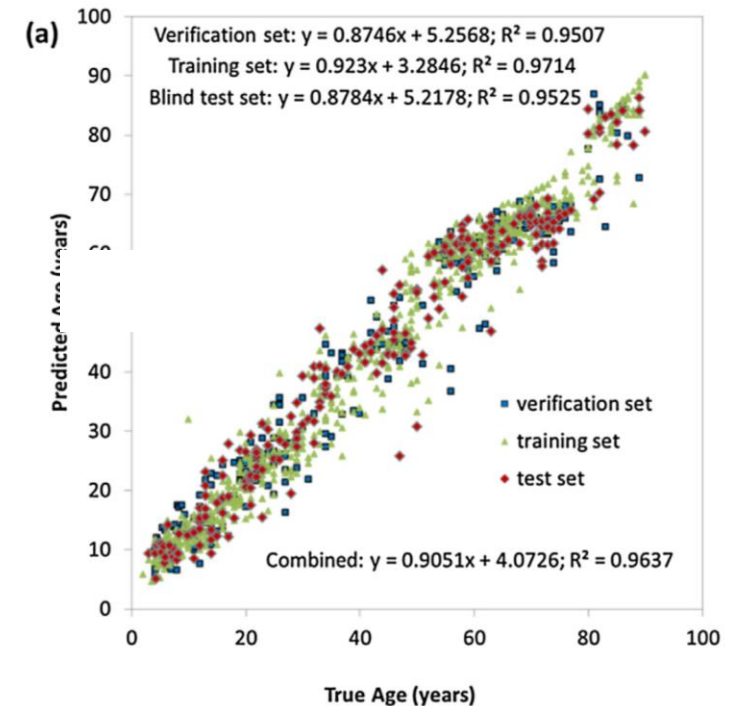
# Regression: Biological examples

- Prediction of treatment efficiency / drug response
  - Predicting drug response of tumors from integrated genomic profiles by deep neural networks. Chiu et al., *BMC Med. Genomic*, 2019
- Prediction of molecular/cellular properties (e.g., protein-DNA binding affinities)
  - Predicting the sequence specificities of DNA- and RNA-binding proteins by deep learning. Alipanahi et al., *Nature Biotech.* 2015



# Regression: Biological examples

- Age prediction from DNA methylation (e.g., for forensics)
  - **DNA methylation-based forensic age prediction using artificial neural networks and next generation sequencing.** Vidaki et al. *Forensic Sci Int Genet.* 2017



# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

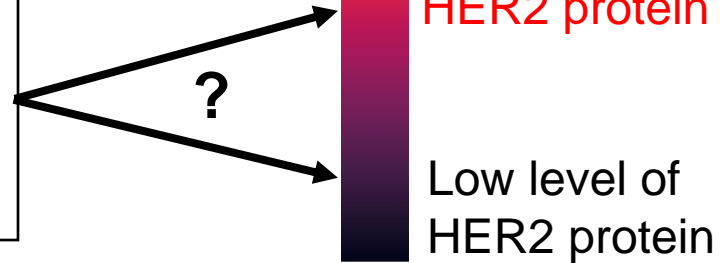
<https://github.com/BoevaLab/Teaching>

## TASK 1:



	<i>Gene1</i>	<i>Gene2</i>	...	<i>Gene<sub>p</sub></i>
$Z_1$	$X_{11}$	$X_{12}$	...	$X_{1p}$
$Z_2$	$X_{21}$	$X_{22}$	...	$X_{2p}$
$\vdots$	$\vdots$	$\vdots$	$\ddots$	$\vdots$
$Z_n$	$X_{n1}$	$X_{n2}$	...	$X_{np}$

+ clinical stage + age



Treatment with HER2 inhibitors: Herceptin (trastuzumab) or Tykerb (lapatinib)

The **HER2** protein is coded by the **ERBB2** gene, frequently amplified in human breast cancer

**Target variable (y):** Reverse Phase Protein Array (RPPA) value of HER2 presence in tumor cells

# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

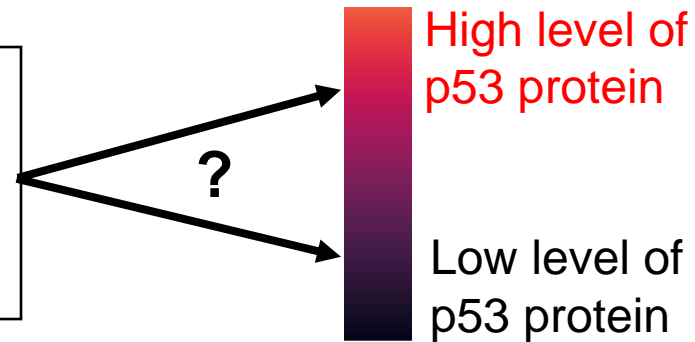
<https://github.com/BoevaLab/Teaching>

## TASK 2:



	Gene1	Gene2	...	Gene $p$
$Z_1$	$X_{11}$	$X_{12}$	...	$X_{1p}$
$Z_2$	$X_{21}$	$X_{22}$	...	$X_{2p}$
$\vdots$	$\vdots$	$\vdots$	$\ddots$	$\vdots$
$Z_n$	$X_{n1}$	$X_{n2}$	...	$X_{np}$

+ clinical stage + age



Standard chemotherapy, e.g., cisplatin

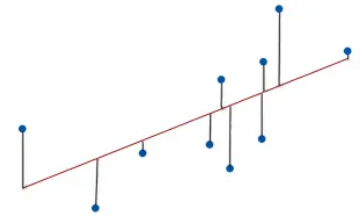
The **p53** protein is coded by the **TP53 gene**, frequently deleted, mutated or repressed in human cancers

**Target variable (y):** Reverse Phase Protein Array (*RPPA*) value of p53 presence in tumor cells



# Classic regression models

- Ordinary Least Squares
- Lasso (L1 penalty on model coefficients)
- Ridge (L2 penalty on model coefficients)
- Elastic Net (L1 and L2 penalty on model coefficients)

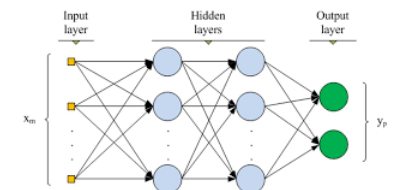
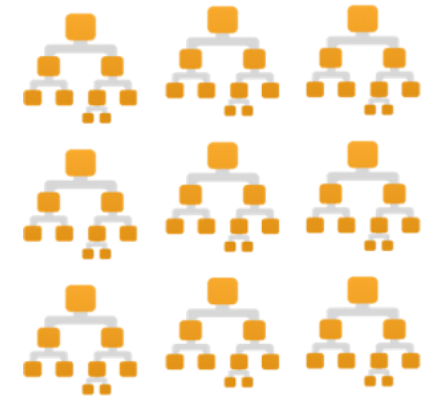


$$+ \lambda \sum_{j=1}^p |\beta_j|$$

$$+ \lambda \sum_{j=1}^p |\beta_j|^2$$

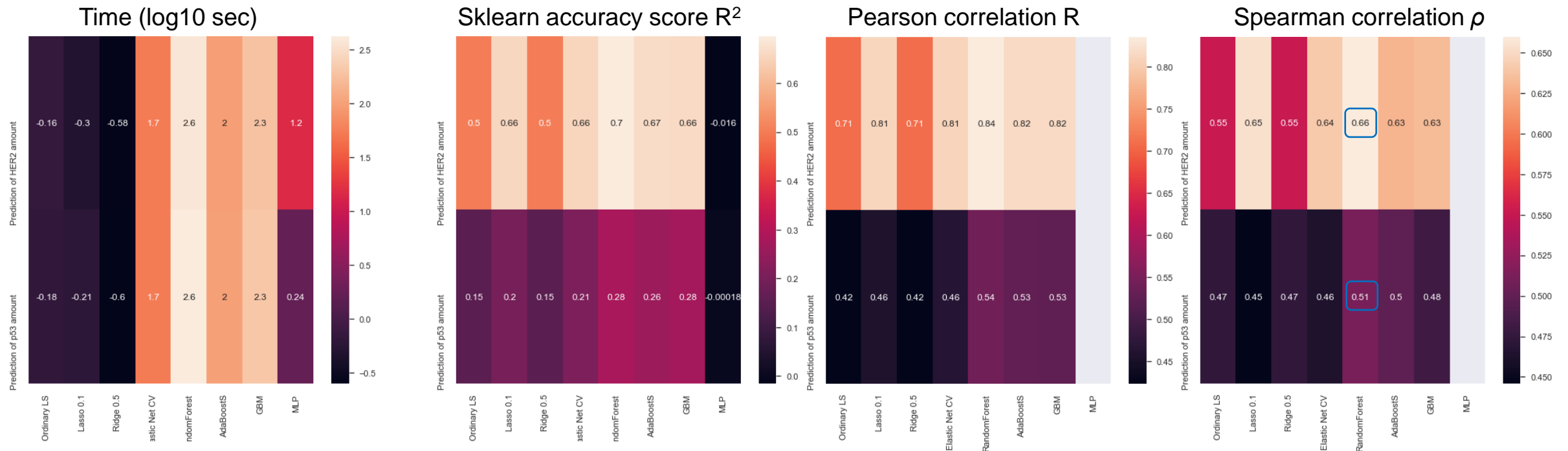
$$+ \lambda_1 \sum_{j=1}^p |\beta_j| + \lambda_2 \sum_{j=1}^p |\beta_j|^2$$

- Random Forest
- AdaBoost
- Gradient Tree Boosting (gradient boosting machine, GBM)
- Multi-layer perceptron (MLP)



# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)



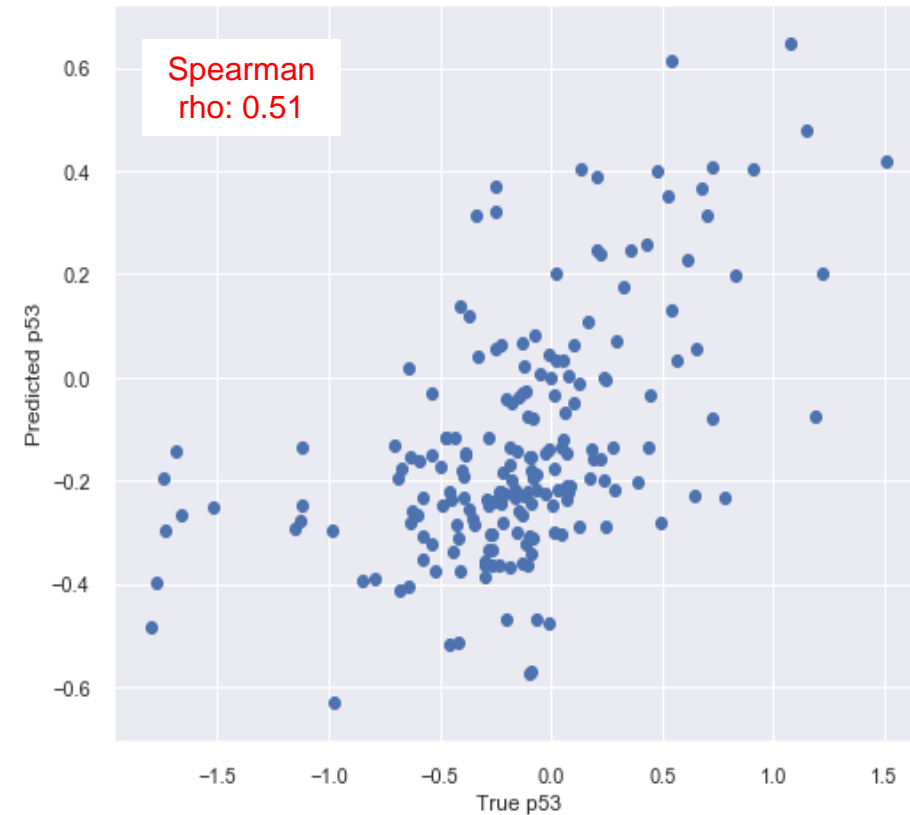
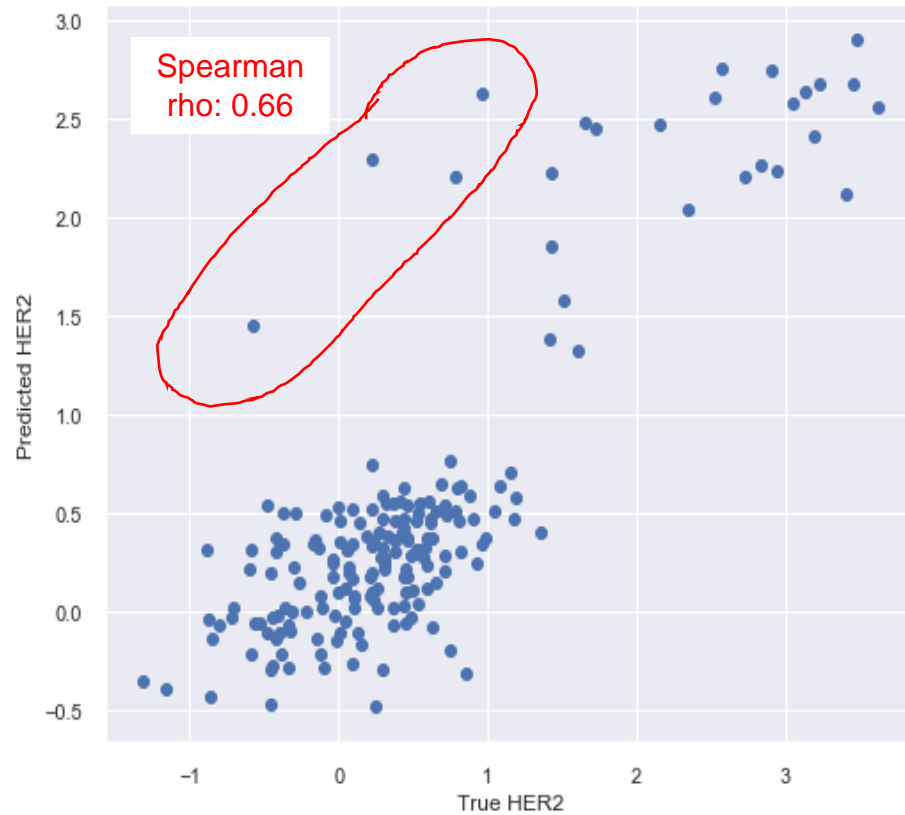
Coefficient of determination  $R^2 = \left(1 - \frac{RSS}{\sum(y_i - \bar{y})^2}\right)$

$Spearman \rho (HER2, ERBB2) = 0.63$

$Spearman \rho (p53, TP53) = 0.27$

# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

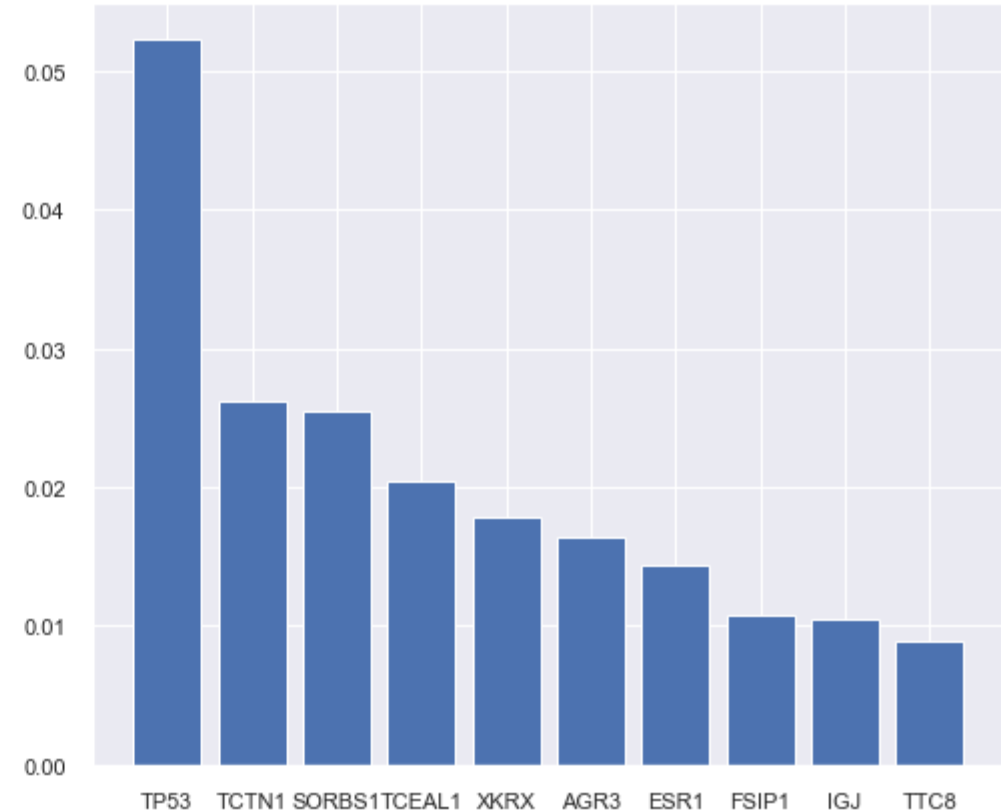
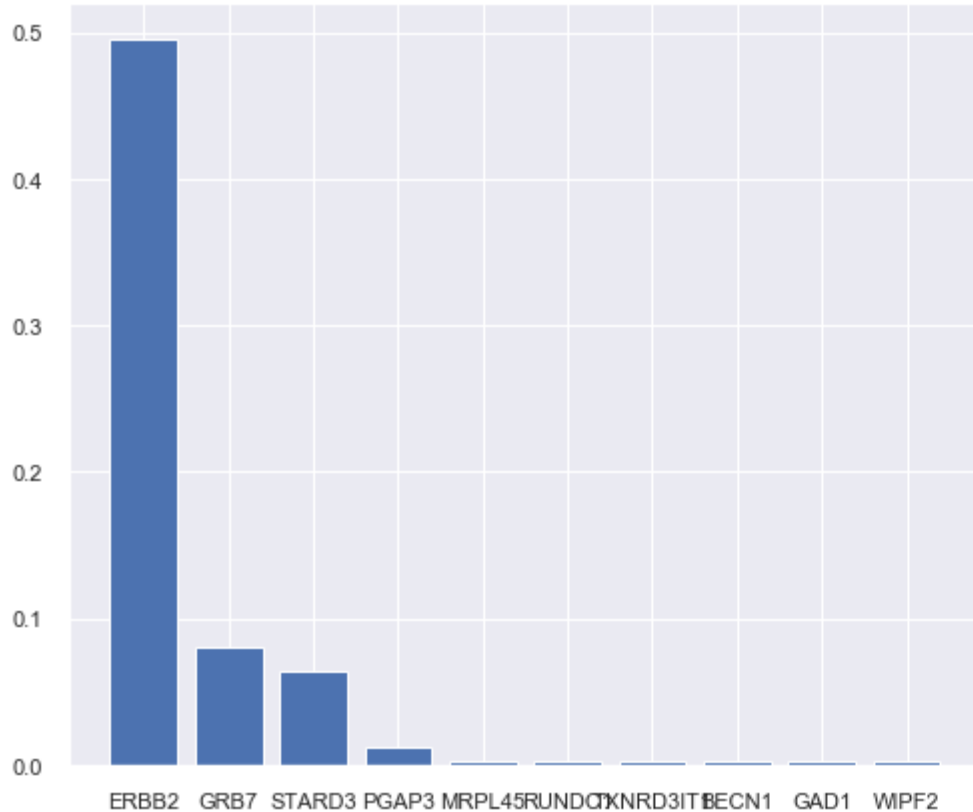


Random Forest predictions

# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

## FEATURE IMPORTANCE:

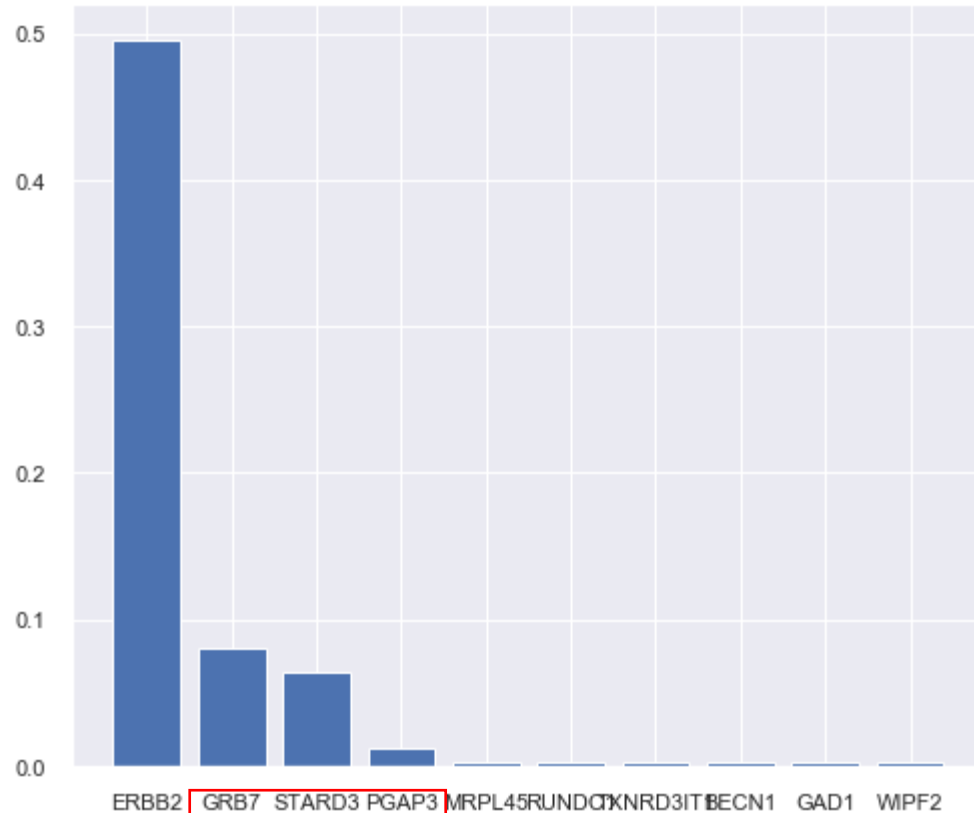


Random Forest predictions

# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

## FEATURE IMPORTANCE:



chr17 chr17:39,644,374-39,754,484 Go

109 kb

39,660 kb 39,680 kb 39,700 kb 39,720 kb 39,740 kb

STARD3 PGAP3 ERBB2 MIR4728 GRB7

Appl Immunohistochem Mol Morphol, 2017 Sep;25(8):553-558. doi: 10.1097/PAI.0000000000000349.

### GRB7 Expression and Correlation With HER2 Amplification in Invasive Breast Carcinoma.

Bivin WW<sup>1</sup>, Yergiyev O, Bunker ML, Silverman JF, Krishnamurti U.

Author information

#### Abstract

Growth factor receptor-bound protein 7 (GRB7) gene is located adjacent to the HER2 gene on the 17q12-21 amplicon, is often coamplified with HER2 in a subset of breast cancers, and has been implicated in resistance to anti-HER2 and antiestrogen therapy. This study investigated the correlation of GRB7 expression by immunohistochemistry with HER2 expression, HER2 amplification, increased chromosome 17 copy number, and other prognostic and predictive factors in invasive breast cancer, including histologic grade, pathologic stage, and ER, PR, and p53 status. Paraffin-embedded samples of 188 invasive breast carcinomas with documented HER2, ER, and PR testing were collected and divided into 3 groups: cases positive for HER2 overexpression/gene amplification (n=60), negative for HER2 overexpression (n=97), and cases with increased chromosome 17 copy number without HER2 amplification (n=31). GRB7 expression was evaluated on all 188 cases. In addition, p53 immunohistochemistry was performed on 13 HER2+/GRB7+ cases and 39 HER2+/GRB7- cases. GRB7 expression correlated strongly with HER2 overexpression. GRB7 expression was present in 20/60 (33.33%) of HER2+ cases, compared with 1/97 (1.03%) HER2- cases, and 1/31 (3.22%) increased chromosome 17 copy number cases (P<0.0001). In HER2+ cases, GRB7 expression was found to correlate significantly with a greater degree of HER2 amplification. The mean±SEM HER2 copy number was 21.14±2.59 in GRB7+ cases, compared with 9.8±1.38 in GRB7- cases (P=0.0001). GRB7 expression correlated significantly with ER negativity (P=0.012) and p53 positivity (P=0.03). GRB7 expression did not correlate with histologic grade, pathologic stage, or PR expression. Our data shows that GRB7 expression in invasive breast cancer correlates with markers of a more aggressive phenotype, including HER2 overexpression, a greater degree of HER2 amplification, ER negativity, and p53 positivity.

PMID: 26945445 DOI: 10.1097/PAI.0000000000000349

# Hands-on: Prediction of protein concentration based on mRNA data (Breast cancer samples)

1. Concentration of HER2 (coded by the *ERBB2* gene)
2. Concentration of p53 (coded by the *TP53* gene)

## FEATURE IMPORTANCE:



Review

### The p53 Pathway in Glioblastoma

Ying Zhang<sup>1,†</sup>, Collin Dube<sup>1,†</sup>, Myron Gibert Jr.<sup>1,†</sup>, Nichola Cruickshanks<sup>1</sup>, Baomin Maeve Coughlan<sup>1</sup>, Yanzi Yang<sup>1</sup>, Initha Setiady<sup>1</sup>, Ciana Deveau<sup>1</sup>, Karim Saoud<sup>1</sup>, Cassandra Grello<sup>1</sup>, Madison Oxford<sup>1</sup>, Fang Yuan<sup>1</sup> and Roger Abounader<sup>1,2,3,\*</sup>

“ARF deletion [from p53 pathway] is correlated with overexpression of tectonic family member 1 (TCTN1), a protein involved in a diverse range of cellular processes, including promotion of GBM cell proliferation”.

www.impactjournals.com/oncotarget/

Oncotarget, 2017, Vol. 8, (No. 6), pp: 9108-9122

Research Paper

### SORBS1 suppresses tumor metastasis and improves the sensitivity of cancer to chemotherapy drug

Lele Song<sup>1,2</sup>, Renxu Chang<sup>1,2</sup>, Cheng Dai<sup>1,2</sup>, Yanjun Wu<sup>1,2</sup>, Jingyu Guo<sup>1,2</sup>, Meiyuan Qi<sup>1</sup>, Wu Zhou<sup>3</sup>, Lixing Zhan<sup>1</sup>

“Silencing of SORBS1 [...] attenuates chemical drug sensitivity especially that to cisplatin, by inhibition of p53 in **breast cancer cells.**”

Meng et al. *Journal of Translational Medicine* 2014, 12:288  
<http://www.translational-medicine.com/content/12/1/288>

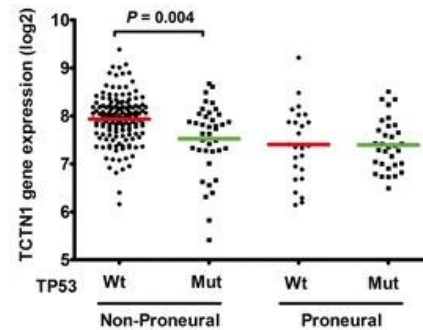


RESEARCH

Open Access

### Expression and prognostic significance of TCTN1 in human glioblastoma

Delong Meng<sup>1†</sup>, Yuanyuan Chen<sup>1†</sup>, Yingjie Zhao<sup>1</sup>, Jingkun Wang<sup>1</sup>, Dapeng Yun<sup>1</sup>, Song Yang<sup>2</sup>, Juxiang Chen<sup>3</sup>, Hongyan Chen<sup>1</sup> and Daru Lu<sup>1\*</sup>



0.05

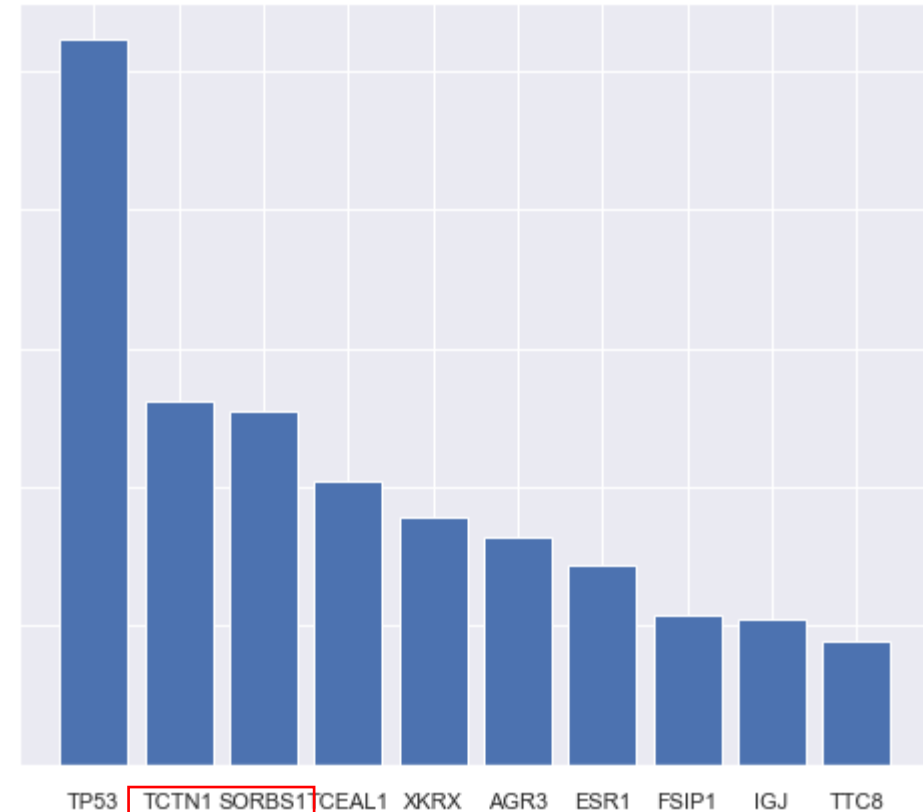
0.04

0.03

0.02

0.01

0.00

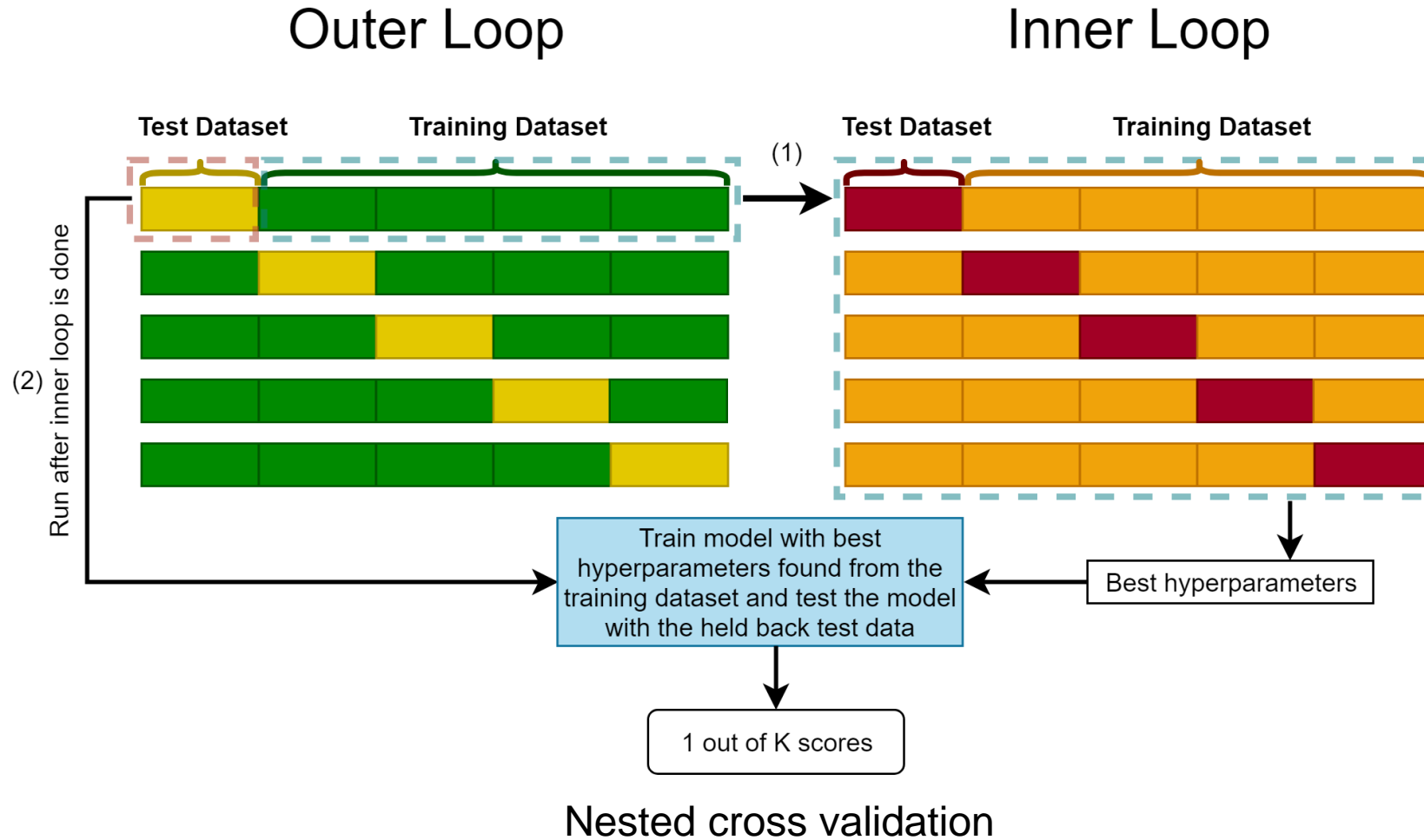


# Take home message: Regression

- Regularized methods generally work better
  - Regularization may prevent over-fitting and select “important” features
- Regularized linear methods may provide accuracy similar to non-linear methods
- Neural networks do not always win
- Checking the feature importance may provide insights into biological mechanisms

# Selection of hyperparameters via cross validation

What we did not do today, but in real life we should do it:





# Take home message

- Classification and regression are extremely widely used in biology and medicine to automatize decisions of clinicians (diagnosis, choice of treatment) and predict treatment response and side effects
- The accuracy of predictions depends a lot on the information present in the data rather than on the ML method used
  - In our hands-on exercises the difference in accuracy between linear and non-linear methods varied between 0.5% and 15%
- There is no method that works the best in any situation
- Cross validation should be always applied to select the best hyperparameters
- In real life, one should compare a model built on omics data (+ clinical) with a model built using clinical variables only

**ETH** zürich

Thank you for your attention!

Professor Valentina Boeva  
valentina.boeva@inf.ethz.ch

ETH Zurich  
Dept of Computer Science  
CAB F51.2  
Universitatstrasse 6  
8092 Zurich, Switerland

[www.boevalab.com](http://www.boevalab.com)

