

# 1 Towards uncertainty quantification 2 in high-throughput biology

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## 8 Abstract

9 Advances in biological technologies have facilitated the generation of large-scale datasets,  
10 profiling biological systems at an unprecedented scale and resolution. The high dimensionality  
11 and complexity of these datasets require machine learning (ML) approaches to extract  
12 meaningful and actionable biological insights. However, current ML models often lack  
13 interpretability for human practitioners, and their predictions typically do not include a measure  
14 of uncertainty. In this study, we apply Gaussian Process (GP) to two high-throughput biological  
15 datasets, single cell RNA sequencing (scRNA-seq) and mass spectrometry-based metabolomics  
16 (MS) for classification tasks. We further utilize the uncertainty estimates to propose new  
17 experiments aimed at improving the model's confidence. Collectively, our results indicate that GP  
18 is widely applicable in various stages of biological data analysis.

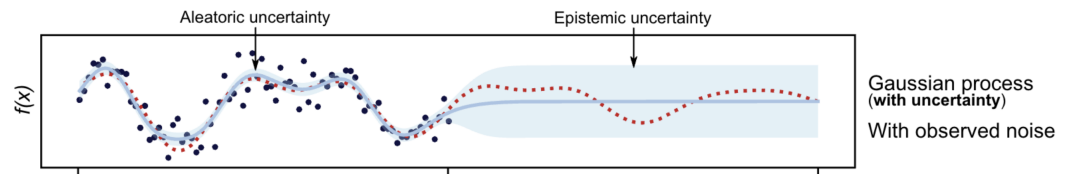
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## 20 Introduction

21 High-throughput biology, with its capacity to generate large-scale datasets of biological systems, is  
22 a powerful tool that aids in understanding complex human diseases. However, this avalanche of  
23 data introduces new challenges in data analysis. To extract biological signals from these datasets,  
24 numerous groups have devised machine learning (ML) approaches for data analysis and low-dimensional  
25 representation (McInnes et al.)(family=Maaten and Hinton)(Maćkiewicz and Ratajczak). Large-scale  
26 datasets inherently exhibit variability due to biological, technical, and stochastic factors, making it  
27 critical to quantify the uncertainty in the data analysis process.

28 Uncertainty quantification assists in assessing whether a model prediction is under- or over-  
29 confident and provides a measure of the reliability of model predictions. This is particularly crucial  
30 in high-stakes applications like medical diagnosis, where prediction reliability is vital (Unc). In ex-  
31 perimental biology, uncertainty quantification can help guide the design of new experiments. For  
32 instance, single-cell RNA sequencing (scRNA-seq), a powerful tool for profiling the transcriptome  
33 of individual cells, is prone to technical noise (Sin). While this noise can be reduced by increasing  
34 the sequencing depth, doing so is often infeasible due to cost and time constraints. In such cases,  
35 uncertainty quantification can help guide the design of new experiments by suggesting which cell  
36 types to prioritize next to maximize information gain.

37 In this study, we aim to quantify uncertainty in high-throughput biological datasets. We choose  
38 to focus on two primary data modalities: single-cell RNA sequencing (scRNA-seq) and mass spectrometry-  
39 based metabolomics (MS). Both data modalities are commonly used in biological research and are  
40 subject to technical noise and batch effects. We demonstrate that Gaussian Processes (GP) can be  
41 employed to quantify both uncertainties from data noise (aleatoric uncertainty) and from model  
42 uncertainty (epistemic uncertainty). We then utilize the uncertainty estimates to suggest new ex-



**Figure 1.** Two types of uncertainty in machine learning. Aleatoric uncertainty is due to noise in the data, while epistemic uncertainty is due to uncertainty in the model.

43 periments that enhance the model's confidence. Collectively, our results suggest that GP can be  
 44 widely applicable in various types of biological data.

45 This manuscript is organized as follows: We first introduce the concept of uncertainty quantifi-  
 46 cation and the Gaussian Process. We then apply the Gaussian Process to a real-world dataset and  
 47 compare its performance with other machine learning methods. Finally, we discuss the advantages  
 48 and disadvantages of using the Gaussian Process in omics data analysis.

## 49 Results

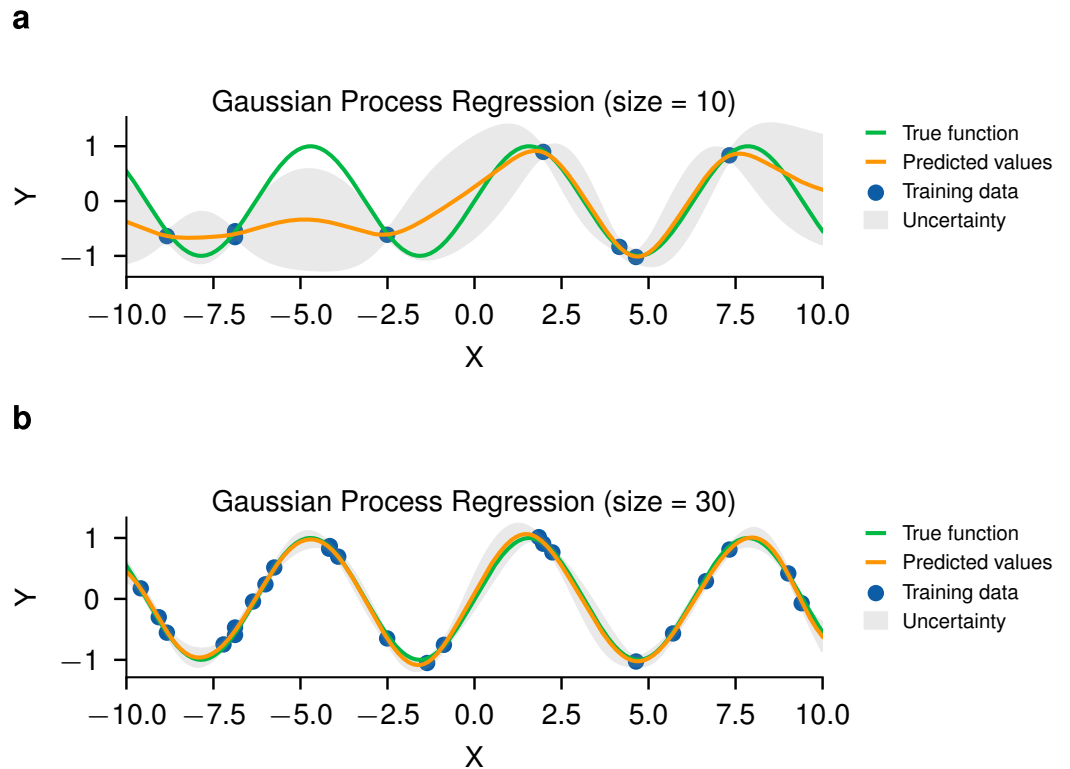
### 50 Gaussian Processes

51 Gaussian processes (GPs) are powerful and flexible tools in machine learning and statistics for  
 52 modeling complex relationships between variables and predicting uncertain outcomes. They have  
 53 been employed in a wide range of applications, including computer vision, robotics, bioinformatics,  
 54 and finance (Kalaitzis and Lawrence)(Simek et al.). Fundamentally, GPs are a collection of random  
 55 variables, any finite number of which have a joint Gaussian distribution. They can be used to model  
 56 functions that map inputs to outputs, such as the relationship between the features of a dataset  
 57 and their associated labels. Unlike traditional regression techniques that assume a fixed functional  
 58 form for the relationship between input and output variables, GPs allow for a more flexible mod-  
 59 eling approach capable of capturing complex nonlinear relationships.

60 One of the key strengths of GPs is their ability to provide uncertainty estimates alongside their  
 61 predictions. This is particularly useful in situations where data is noisy or there is a high level of  
 62 uncertainty in the underlying model. Figure 2 shows an example of a GP regression model with  
 63 uncertainty estimates. The shaded region represents the 95% confidence interval, which is wider  
 64 in regions with fewer data points and narrower in regions with more data points. The uncertainty  
 65 estimates can be used to inform decision-making and improve prediction reliability.

66 Uncertainty quantification is crucial in numerous applications, such as medical diagnosis. Here,  
 67 prediction accuracy is of utmost importance and must be accompanied by a measure of confidence  
 68 in the results. To estimate uncertainty, GPs model the output variable as a Gaussian distribution  
 69 with a mean and variance. The variance represents the uncertainty in the prediction, with larger  
 70 variances indicating greater uncertainty. GPs can also be used to perform Bayesian inference, al-  
 71 lowing for the incorporation of prior knowledge and the updating of beliefs as new data becomes  
 72 available. Bayesian inference proves especially useful in situations with limited data or when the  
 73 model is complex and challenging to estimate using traditional techniques.

74 GPs offer several advantages over other machine learning techniques, such as neural networks  
 75 and support vector machines (Işık and Alptekin). They are non-parametric and do not assume a  
 76 fixed functional form for the relationship between input and output variables, thereby allowing for  
 77 greater flexibility and the modeling of complex relationships. GPs can also manage missing data  
 78 and noisy measurements, which are common in biological sciences. Additionally, they provide a  
 79 measure of uncertainty that can inform decision-making and enhance prediction reliability.



**Figure 2.** Gaussian Processes Regression on different number of test points.

80 **Active learning**

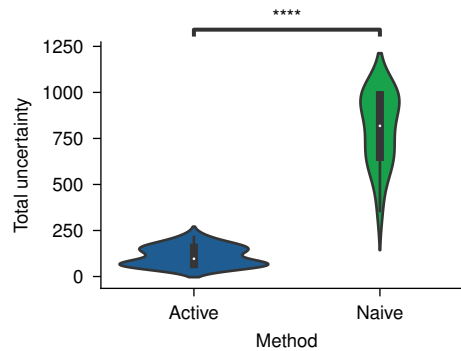
81 Active learning is a machine learning technique in which the algorithm determines which data to  
 82 label for training, instead of passively receiving labeled data. This approach can be beneficial in  
 83 scenarios where obtaining labeled data is expensive or time-consuming (Hemmer et al.). The goal  
 84 is to minimize the amount of labeled data needed to train a model by intelligently choosing which  
 85 data points to label next.

86 Active learning algorithms operate by iteratively selecting the most informative samples to la-  
 87 bel, based on a predefined criterion. Several criteria can be utilized for this purpose, including  
 88 uncertainty sampling, query by committee, and expected model change. Uncertainty sampling  
 89 involves choosing the data points for which the model is most uncertain about the correct label.  
 90 Query by committee involves selecting the data points that are most controversial among a group  
 91 of models. Expected model change involves selecting data points expected to have the greatest  
 92 impact on the model's performance.

93 One method of implementing active learning is through Gaussian processes (Riis et al.). Gaus-  
 94 sian processes are a flexible and potent probabilistic modeling technique that can be used for  
 95 regression and classification tasks. In the context of active learning, Gaussian processes can be  
 96 employed to model the uncertainty of the model's predictions (Riis et al.).

97 In Gaussian process-based active learning, the algorithm begins with an initial set of labeled  
 98 data points and fits a Gaussian process model to these points. It then selects the data point with  
 99 the highest uncertainty according to the Gaussian process model and requests its label from an  
 100 oracle (i.e., a human expert). This labeled data point is then added to the training set, and the  
 101 Gaussian process model is updated. The process is repeated iteratively until the desired level of  
 102 accuracy is achieved or the labeling budget is exhausted.

103 One advantage of Gaussian process-based active learning is that it facilitates a principled ap-  
 104 proach to modeling uncertainty [10]. The Gaussian process model can be used to compute the



**Figure 3.** Active learning improves model confidence.

105 uncertainty of the model's predictions, which can then guide the selection of the most informative  
 106 data points for labeling. Another advantage is that Gaussian processes are non-parametric mod-  
 107 els, meaning they can adapt to complex patterns in the data without making strong assumptions  
 108 about the underlying distribution.

109 However, Gaussian process-based active learning also has some limitations. One limitation is  
 110 that Gaussian processes can be computationally expensive to train and evaluate, especially for  
 111 large datasets. Another limitation is that the performance of the Gaussian process model de-  
 112 pends on the choice of kernel function and hyperparameters, which can be challenging to optimize  
 113 (Krauth et al.).

### 114 **Active learning in improving confidence**

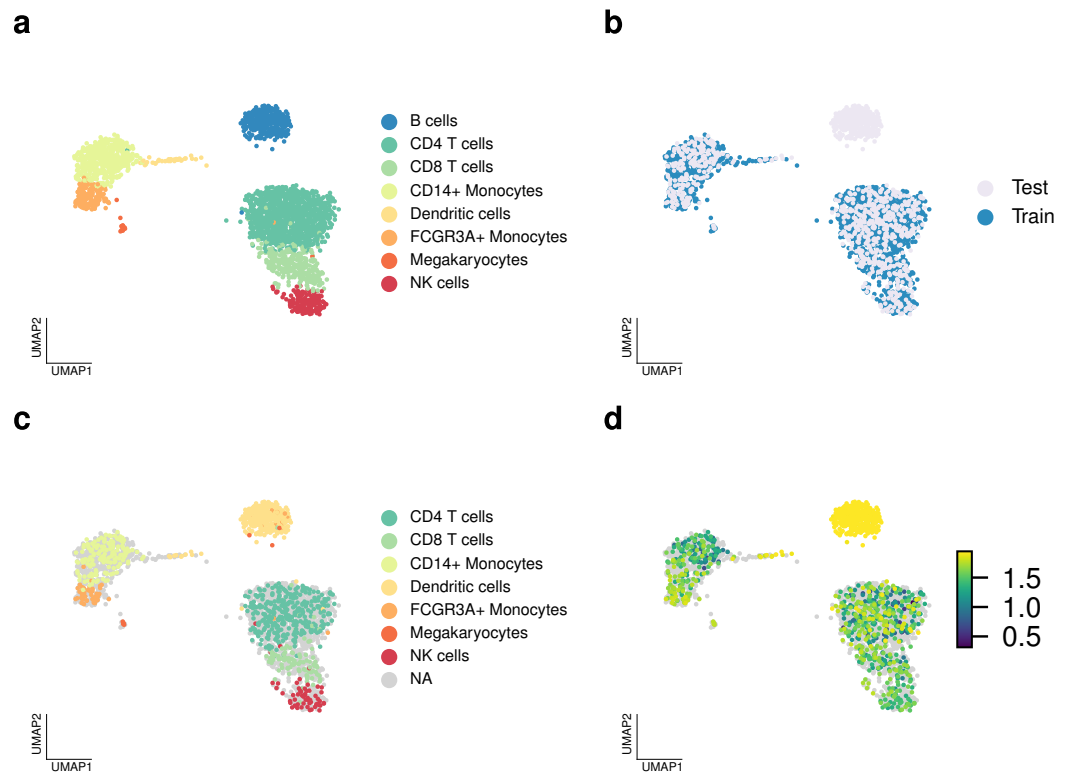
115 Following the example in Figure 2, an uncertainty-guided active learning approach can be deployed  
 116 to acquire new data points that enhance the model's confidence. Figure 3 illustrates an instance  
 117 of active learning in a GP regression model. The model is initialized with a small number of data  
 118 points and iteratively selects new data points to acquire based on the uncertainty estimates. These  
 119 estimates are used to select data points that are likely to boost the model's confidence. This ap-  
 120 proach is contrasted with a random sampling method, where data points are selected randomly.  
 121 The results demonstrate that the active learning approach is capable of improving the model's con-  
 122 fidence more rapidly than the random sampling method, given the same number of data points.

### 123 **Application in scRNA-seq data**

124 We then sought to apply the Gaussian Process (GP) to a single-cell RNA sequencing (scRNA-seq)  
 125 dataset derived from peripheral blood mononuclear cells (PBMCs). PBMCs represent a heteroge-  
 126 neous population of immune cells, including T cells, B cells, natural killer (NK) cells, monocytes, and  
 127 dendritic cells, among others. In a typical scRNA-seq experiment, PBMCs are first isolated from the  
 128 blood and then subjected to droplet-based or plate-based single-cell capture. Here, individual cells  
 129 are encapsulated into microfluidic droplets or wells. The cells are then lysed, and the RNA is re-  
 130 verse transcribed into complementary DNA (cDNA). This cDNA is subsequently amplified, and the  
 131 resulting library is sequenced using high-throughput sequencing technologies, typically Illumina se-  
 132 quencing. The sequencing data obtained provides information about the gene expression profiles  
 133 of each individual cell.

### 134 **Gaussian Processes reveal epistemic uncertainty in cell type classification**

135 We applied Gaussian Processes to a classification problem to predict cell types based on gene  
 136 expression (Figure 4a). We set aside 50% of the data for testing and used the remaining 50% for  
 137 training. To simulate a real-world scenario where the training data has limited coverage compared  
 138 to a clinical query dataset, we additionally held out all B cells (Figure 4b). Encouragingly, the GP  
 139 model was able to accurately predict the cell type of the test data with high accuracy ( $AUC \approx 1$ )

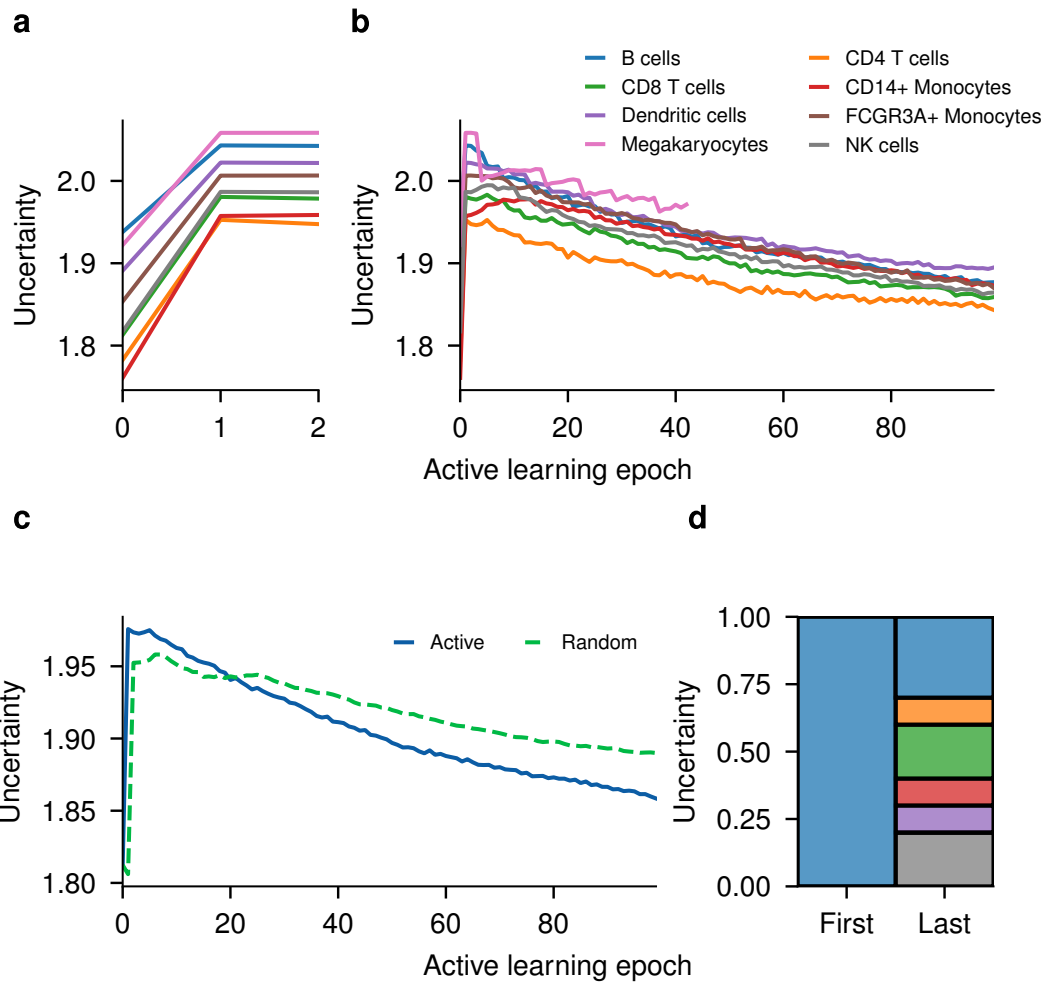


**Figure 4.** scRNA-seq dataset of 3000 peripheral blood mononuclear cell.

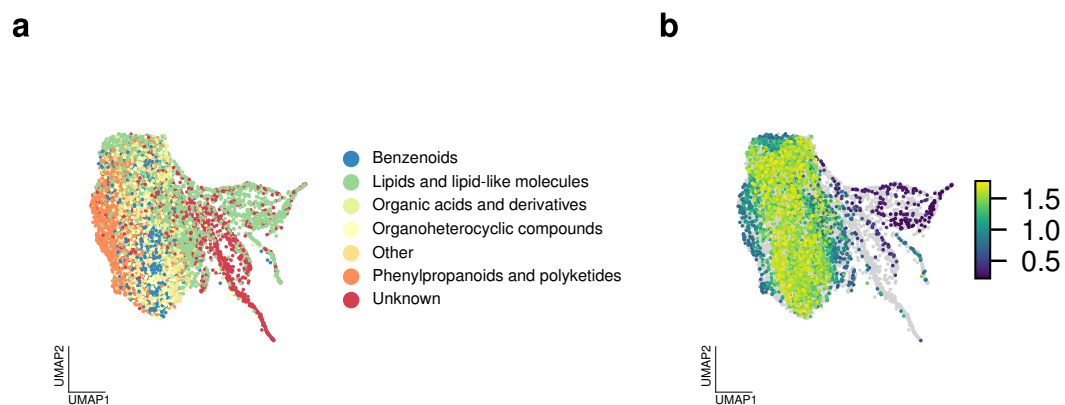
140 (Figure 4c). As anticipated, the model was unable to predict the held-out B cells and incorrectly  
 141 classified the cells as dendritic cells, likely due to their biological similarity. This type of error is  
 142 known as epistemic uncertainty, which arises from the lack of training data, leading to uncertainty  
 143 in the model. This kind of classification is known as a label transfer task, and it's widely used in  
 144 annotating scRNA-seq data. Misannotation is, therefore, a common pitfall in scRNA-seq analysis  
 145 and can lead to erroneous biological conclusions.

146 In the application of GP, we calculated the uncertainty of the prediction as the Shannon entropy  
 147 of the prediction probability associated with each classification (Figure 4d). As expected, the uncer-  
 148 tainty of the prediction was highest among the held-out B cells. Surprisingly, GP was also able to  
 149 assign high uncertainty accurately to rare cell types such as Megakaryocytes and Dendritic cells.

150 Analogous to the sinusoidal function example above, we again used active learning to iteratively  
 151 select new data points to acquire based on the uncertainty estimates. In the earliest iteration,  
 152 model uncertainty appears to increase across cell types (Figure 5a). This is due to the introduc-  
 153 tion of B cells into the new training set, which increases the alphabet size for entropy calculation.  
 154 Initially, B cells exhibited the highest uncertainty, but as more data points were acquired, the un-  
 155 certainty decreased (Figure 5b). In the final iterations, all cell types had lower uncertainty than in  
 156 the initial iterations. This is because the model, having seen more data points, had increased con-  
 157 fidence in its predictions, even for cell types initially included in the training set. Upon examining  
 158 the acquired data, we observed that the model prioritized acquiring B cells in the beginning. As the  
 159 uncertainty of B cells became comparable to other cell types, the model began to acquire other  
 160 cell types (Figure 5d). As a comparison, we also performed a random data acquisition approach,  
 161 analogous to generating the same scRNA-seq dataset experimentally. While the random approach  
 162 also improved the model's confidence, it did so at a much slower rate than the uncertainty-guided  
 163 approach (Figure 5c). This is because the random approach does not consider the model's uncer-  
 164 tainty and is thus unable to prioritize data points likely to improve the model's confidence.



**Figure 5.** Active learning increase model confidence across cell types.



**Figure 6.** Gaussian processes reveal aleatoric uncertainty in molecular class prediction.

165 In summary, our use of GP for label transfer in single-cell RNA-seq data demonstrated high ac-  
 166 curacy and the ability to quantify prediction uncertainty. From the example dataset, we highlighted  
 167 uncertainty associated with both held-out cell types and rare cell types. This understanding can  
 168 help guide future experimental design to selectively enrich for these cell types, in order to improve  
 169 representation in the model.

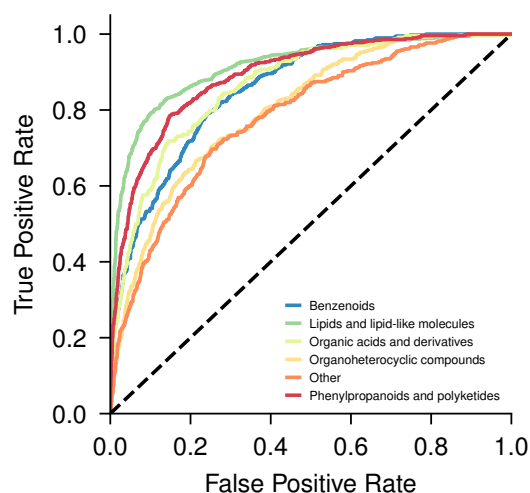
#### 170 **Application in metabolomic data**

171 Mass spectrometry (MS) is a powerful analytical technique used to identify and characterize molecules  
 172 based on their mass-to-charge ratio. The basic principle of mass spectrometry is the generation  
 173 of ions from a sample, which are then separated based on their mass-to-charge ratio using a com-  
 174 bination of electric and magnetic fields. The ions are then detected and the resulting signal is  
 175 analyzed to determine the mass and abundance of the ions present in the sample. The chemical  
 176 structure of the molecule can be identified by analyzing the mass spectrum of the molecular ion  
 177 peak, which represents the intact molecule without any fragmentation. The mass of the molecular  
 178 ion provides information about the molecular weight of the compound. The fragmentation pat-  
 179 terns of the molecule provide information about the chemical bonds within the molecule and can  
 180 be used to reconstruct the molecular structure. These fragmentation patterns can be analyzed  
 181 using software tools that compare the observed mass spectrum to a database of known spectra,  
 182 such as the MassBank database. This comparison can then help identify potential molecular struc-  
 183 tures that match the observed fragmentation pattern. However, identifying unknown compounds  
 184 or compounds from a mixture of samples using mass spectra remains challenging. Over 90% of  
 185 compounds from a typical sample are unknown, and quantifying uncertainty in the identification  
 186 of these compounds is critical, both for the curation of a representative database and for future  
 187 experimental design.

#### 188 **Gaussian Processes reveal aleatoric uncertainty in molecular class prediction**

189 We transformed spectral intensities into feature vectors using a spectral featurizer, and trained a  
 190 GP classifier with 50% of the data, withholding the remaining 50% for testing. Interestingly, even  
 191 without any held-out molecular class, there was substantial uncertainty in the prediction (Figure  
 192 6a, b). Comparing the average uncertainty between molecular classes, we observed that in a UMAP  
 193 representation of the data, the uncertainty is highest where there is greater label mixing within a  
 194 neighborhood. Molecular classes such as lipids exhibited the lowest uncertainty, as they were well  
 195 separated from other classes. This shows that GP is able to capture intrinsic uncertainty due to  
 196 noise in the data, known as aleatoric uncertainty.

197 In order to be considered a viable option for routine MS analysis, GP must demonstrate a clas-  
 198 sification performance that is on par with or superior to state-of-the-art methods. We conducted



**Figure 7.** Gaussian processes classifier has good classification performance relative to state of art methods.

2199 a comparison of the performance of GP to a variety of these methods, including random forest  
 2200 and support vector machine algorithms. Our findings revealed that GP offers comparable perfor-  
 2201 mance to these methods (Figure 7, Data not shown). This suggests that GP could serve as a suitable  
 2202 replacement for these methods, with the added advantage of being able to quantify the level of  
 2203 uncertainty in the prediction. This ability to estimate uncertainty could be particularly beneficial in  
 2204 areas such as metabolomics, where a large proportion of compounds remain unidentified and the  
 2205 ability to quantify the confidence in compound identification can provide valuable information for  
 2206 future experimental design and database curation.

## 2207 Discussion

2208 Our study has demonstrated the utility of Gaussian Processes (GP) in quantifying prediction un-  
 2209 certainty in both single-cell RNA-seq and metabolomic data. For the single-cell RNA-seq analysis,  
 2210 we were able to identify the levels of uncertainty in predictions for held-out cell types as well as  
 2211 rare cell types. With metabolomic data, GP was able to quantify uncertainty within the prediction  
 2212 of molecular classes. Importantly, in both scenarios, we showed that GP can be used to measure  
 2213 the uncertainty of predictions for held-out data points, which can provide valuable insights to help  
 2214 guide the design of future experiments, with the aim of enriching data for these particular points  
 2215 and thus enhancing their representation within the model.

2216 In the current version of our implementation, we largely utilize a paired cosine similarity kernel  
 2217 for GP. However, there is potential for enhancing performance and scalability by exploring other  
 2218 kernels, such as the Radial Basis Function (RBF) kernel. While this study focused mainly on the  
 2219 application of GP as a drop-in replacement for standard analysis tasks within biological datasets,  
 2220 future research could benefit from integrating uncertainty quantification into earlier stages of the  
 2221 analysis process, such as spectral featurization.

2222 Given the often large scale of data in high throughput biology, it can present challenges for  
 2223 the application of GP. To address this, it may be beneficial to explore performance engineering  
 2224 techniques in Julia, such as parallelization and GPU computing, to enhance the performance of  
 2225 GP.

2226 Despite these challenges, we have clearly demonstrated the value of GP in quantifying predic-  
 2227 tion uncertainty within biological datasets. We anticipate that GP will prove to be a valuable tool  
 2228 in perturbation experiments, such as perturb-seq and chemical perturbation experiments, where  
 2229 experiments are not easily scalable and can be expensive to conduct. By quantifying uncertainty,  
 2230 we can more effectively guide the design of future experiments to selectively enrich for the most



231 informative data, ultimately improving our understanding of complex biological systems.

## 232 **Methods and Materials**

### 233 **Computational analysis**

234 Code used in this study is available at [https://github.com/nhcheng/Xavier\\_MS\\_Active\\_Learning\\_](https://github.com/nhcheng/Xavier_MS_Active_Learning_notebook)  
235 [notebook](#). All analyses is performed using both the Julia programming language and Python. Unless  
236 otherwise specified, all gaussian processes analyses is performed using the *scikit-learn* package in  
237 Python. All analysis is performed on a 44-core Intel Xeon CPU computer with 88 GB of RAM.

### 238 **Single-cell analysis**

239 The *pbmc-3k* standard processed dataset is used throughout. All single-cell analysis is performed  
240 using the *scanpy* package in Python.

### 241 **Metabolomic analysis**

242 A pre-trained Siamese neural network MS2DeepScore to predict the structural similarity between  
243 a pair of spectra. For molecular class prediction, Classyfire is used for automated chemical classi-  
244 fication of the molecules.

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