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Sustainable Routes for the Synthesis of Renewable Heteroatom-**Containing Chemicals**

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ABSTRACT: One of the biggest discrepancies between the structure of many utilized chemicals and petrochemicals is the ubiquity of heteroatoms in the former and the lack thereof in the latter. Many commodity chemicals and almost all specialty chemicals and pharmaceuticals contain one or more heteroatoms, but introducing functionalities containing oxygen, nitrogen, sulfur or phosphorus into crude oil-derived chemicals is often a very energy- and resource-intensive endeavor. This and the inevitable depletion of fossil resources in the not too distant future are the main reasons for the



development of sustainable ways to produce compounds bearing heteroatoms. Synthesis of oxygen-containing compounds from renewable resources such as starch, cellulose and hemicellulose is already well-known, and the production of phenolic compounds from lignin is garnering significant attention recently. In the meantime, there is a surge in the valorization of chitin from waste crustacean and insect shells for the production of various nitrogen-containing compounds. Much less explored is the valorization of sulfur- and phosphorus-containing biomass components, although they find some high market value applications. Catalysis plays a central role to enable the conversion of biomass into value-added products with high activity and selectivity. Further developments made by chemical engineers and process technologists will be required to make those processes economically feasible and competitive with current synthetic schemes from fossil resources. This perspective highlights the most recent advances and the upcoming challenges in the development of renewable and sustainable routes toward heteroatomcontaining chemicals.

KEYWORDS: Biomass, Renewable chemicals, Heteroatom-containing compound, Catalysis, Green processes, Chitin, Lignin

INTRODUCTION

C-H bond activation remains one of the biggest challenges in the current chemical industry because of the stable and unpolarized nature of the C-H bond.¹ Although the high energy density renders hydrocarbons ideal transportation fuels, almost all chemicals in everyday life contain one or more elements other than carbon and hydrogen. For instance, all top 10 pharmaceuticals² contain nitrogen elements whereas nine of them contain oxygen as well. Analysis into top 10 pesticides³ reveals a similar situation (Figure 1). Introducing heteroatom elements into hydrocarbons remains chemically challenging, and current technologies put a strain on our environment due to the formation of chemical waste and high energy consumption correlated with the emission of greenhouse gases. Fuels produced from biomass ideally would have a zero carbon footprint, but for various reasons including the necessity for the removal of high amounts of heteroatoms within biomass shift this approach far from carbon neutral.⁴ Direct ways of transforming renewable resources into heteroatom-containing chemicals may circumvent those problems and represent an attractive alternative to conventional processes.^{5,6} One example is lignin, one major component of lignocellulosic biomass, which is composed of an extended phenolic network. Its current predominant use is as a heat

source in the pulp and paper industry, while the potential for generating valuable, oxygenated aromatic building blocks is yet to be fully exploited. Another underutilized material is chitin, the second most abundant biopolymer on earth after cellulose, a major component of crustacean waste, the exoskeleton of insects and the cell walls of fungi.^{8,9} Despite it probably being the only copious renewable source of N-containing chemicals and materials, the overwhelming majority is currently dumped into landfills and in the ocean.¹⁰ Although S- and P-containing biomass sources are much less abundant, some examples exist and their full potential should be considered and assessed. In parallel, alternative strategies including the conversion of renewable O-containing chemicals into their sulfur- and phosphorus-analogues can offer viable pathways for a more sustainable chemical industry (Figure 2).

O-CONTAINING CHEMICALS

Oxygen is the most common heteroatom in all organic matter on this planet. Plant biomass predominantly consisting of cellulose, hemicellulose and lignin makes up a major part of it.

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Perspective



Figure 1. Chemical structures of the top 10 pharmaceutical products by prescription in 2016 and top 10 conventional agricultural pesticide active ingredients in 2012 with heteroatoms (except halogens and alkali metals) marked in color.

The valorization of plant biomass components for the production of biofuels,^{4,5,11–22} various oxygen-containing platform^{23–28} or even higher value chemicals^{11,13,16,17,29–34} have been either achieved or are currently under investigation. Homogeneous and heterogeneous catalysts are among the most common tools to convert lignocellulosic biomass into more useful chemical products. Until relatively recently, the potential of lignin, the only abundant aromatic oxygenated biopolymer has been underestimated and thus far, large-scale industrial processes solely focus on the conversion of polysaccharides. The reason might be the heterogeneity and recalcitrance of lignin due to its extended C-C and C-O linking network or the change of the native lignin structure during the processing of lignocellulosic biomass under harsh conditions. Along this line, lignin was either actively stabilized before further treatment of the sugar components $^{35-37}$ or lignin was extracted from the biomass resource first.^{38,39} Gaining a more fundamental understanding on the processes involved in biomass fractionation,⁴⁰⁻⁴³ lignin depolymerization^{40,41,44} and

its partial or complete defunctionalization^{45–50} may facilitate the development of more efficient catalyst, reactor and process designs. A certain emphasis has been put on the production of biorenewable polymers^{51–53} because they are desirable products and are made from simple repeating units.⁵⁴ In this Perspective, we will limit the discussion on alternative pathways toward three dicarboxylic acids as polymer precursors, to highlight how some of the most important synthetic polymers or substitutes thereof are achieved from renewable sources.

Adipic acid, arguably the most important dicarboxylic acid in industry, is a major component of nylon-66 alongside its diamine pendant. The majority of this precursor is currently produced from crude oil by the nitric acid oxidation of KA oil (a mixture of cyclohexanol and cyclohexanone) made from benzene. Another process starts from cyclohexene utilizing hydrogen peroxide as milder and slightly more environmentally benign oxidant but the significantly higher substrate price prevents large-scale industrial applications.



Figure 2. Overview of the suggested pathways for the production of heteroatom-containing chemicals from renewable resources, contrasted to their current production from crude oil.

In the past two decades, various processes have been developed to employ lignocellulosic biomass or sugar resources for the sustainable production of adipic acid, avoiding the use of carcinogenic benzene (Figure 3).55 Drivers of this technology are biotechnology/biocatalysis, chemical catalysis advances and often a combination of both.^{56,57} Glucose has been converted into glucaric acid on a heterogeneous catalyst using molecular oxygen followed by a hydrogenation reaction to the dicarboxylic acid.⁵⁸ Alternatively, mucic acid derived from galactose was converted into an adipic acid ester in a homogeneous rhenium-catalyzed deoxydehydration reaction and a subsequent platinum-catalyzed transfer hydrogenation.⁵⁵ Two of the most prevalent sugar-derived platform chemicals, levulinic acid and 5-hydroxymethylfurfural (5-HMF), have been shown to be convertible into adipic acid. Levulinic acid needs to be transformed into γ -valerolactone⁶⁰ and subsequently into a mixture of several pentenoic acid derivatives by reactive distillation in the presence of a solid acid catalyst. A homogeneous palladium-catalyzed carbonylation reaction then produces adipic acid in good yields.⁶¹ Multiple pathways exist for the production of adipic acid from 5-HMF. For instance, 5-HMF can be converted into hexanediol by reduction and ringopening over heterogeneous catalysts, followed by oxidation to adipic acid using a biocatalyst.^{62,63} Another pathway that relies solely on chemical catalysts includes the conversion of 5-HMF into 2,5-furandicarboxylic acid (FDCA),⁶⁴ which can be subsequently converted into adipic acid via a metal-free

process.⁶⁵ Via dearomatization, deoxygenation and dealkylation, several lignin-derived monomers could possibly be funneled into a KA oil which can then be further oxidized into adipic acid relying on the conventional process.^{66,67} Despite its reliance on a renewable feedstock, the harsh oxidative conditions are environmentally harmful. In another approach, depolymerized lignin was converted to muconate using an engineered strain of *Pseudomonas putida* KT2440, the product subsequently purified and hydrogenated to adipic acid using a conventional platinum catalyst supported on activated carbon.^{68–71} The electrocatalytic reduction of muconic acid at room temperature under atmospheric pressure has been considered as an alternative and possibly more sustainable pathway toward several products such as adipic acid.⁷²

Another very common monomer is terephthalic acid, the precursor of polyethylene terephthalate (PET), especially useful for the production of synthetic fibers and drinking bottles. The conventional synthesis starts from oil reformates. Various purification steps including crystallization, distillation and freezing are necessary to obtain purified *p*-xylene, which then needs to be oxidized to the respective dicarboxylic acid in the Amoco process employing molecular oxygen as oxidant and acetic acid as solvent.⁷³ Particularly relevant for the synthesis of terephthalic acid from biorenewable resources is the Diels–Alder reaction of various alkenes and dienes (Figure 4).⁷⁴ *p*-Xylene is accessible from biomass via several pathways. In one recent example, 5-HMF was converted into dimethylfuran and



Figure 3. Conventional crude oil-based and new biomass-based pathways for the production of adipic acid.



Figure 4. Conventional crude oil-based and new biomass-based pathways for the production of terephthalic acid.

then reacted with ethylene to yield the cycloadduct which can then be transformed into *p*-xylene by dehydration. Surprisingly, a phosphorus-containing zeolite leads to yields close to 100% due to the superior selectivity toward the desired product.⁷⁵ Alternatively, acrolein and isoprene, two common products from renewable resources, were converted into an aldehyde intermediate which underwent a dehydroaromatization-hydrodeoxygenation cascade reaction on a supported tungsten carbide catalyst to yield *p*-xylene without the necessity to add redoxactive components.⁷⁶ Besides that, the direct conversion of Diels-Alder reactants to terephthalic acid or its esters has been shown.⁷⁷ Upon selective oxidation of 5-HMF to its monocarboxylic acid derivative, a reaction with ethylene in the presence of a Lewis acid catalyst was successful, a reaction otherwise impossible with FDCA.⁷⁸ Similar to the production of adipic acid, muconic acid can serve as a starting materials for diethyl terephthalate. Only upon esterification the cycloadduct formed over a silicotungstic acid catalyst and then further dehydrogenated in the presence of a palladium catalyst reaching yields of above 80% based on muconic acid.⁷⁹ A different pathway to obtain muconates from renewable resources is the cross metathesis of sorbates and acrylates employing common ruthenium-based catalysts.⁸⁰ Due to its aromaticity and high oxygen content, lignin seems to be an ideal starting point for the production of terephthalic acid. Indeed, a synthesis starting from lignin-derived phenolic acids had been proposed previously. Vanillic and syringic acid were selectively demethoxylated and carboxylated using a two-step process in a fixed-bed reactor.⁸¹ We envisage that lignin can be directly converted into terephthalic acid by appropriate funneling of the various obtained aromatic monomers.

FDCA is a component of polyethylene furanoate (PEF) that is considered a potential substitute for PET for certain applications where a higher gas barrier and glass transition temperature or a lower melting point are required. Due to its heteroaromatic structure, FDCA is not easily achieved from crude oil but is synthesized from biomass resources. The most common process for the production of FDCA relies on the selective oxidation of 5-HMF through a range of well-known intermediates (Figure 5). The reaction has been achieved by using homo- or heterogeneous catalysts, photo- or electro-chemistry or biological processes with various oxidants.⁸²⁻⁸⁴ Most homogeneous and heterogeneous catalytic processes are based on noble metals such as platinum,^{85,86} palladium,^{86–90} gold,^{87,89,91,92} and ruthenium^{93,94} or non-noble metals includ-ing iron,^{94,95} nickel,⁸⁸ cobalt,^{93,96} manganese,^{93,96} zirconium⁹⁵ or zinc.⁹⁴ Some examples for the oxidation of 5-HMF to FDCA avoiding the use of metals exist⁹⁷⁻¹⁰¹ but are limited by the requirement of stoichiometric oxidants other than air or low catalyst recyclability.

Besides those conventional catalytic methods, photo- and electrocatalytic routes have been investigated. Cobalt-thioporphyrazine-moieties bound to graphitic carbon nitride afforded FDCA in up to 96% yield from 5-HMF using a simulated sunlight-mediated room temperature and ambient pressure aerobic oxidation. The interaction between the cobalt complex and C₃N₄ promotes the formation of singlet oxygen and disables the generation of less selective hydroxyl radicals as evidenced by active spin trapping and EPR.¹⁰² In 2016, a system was reported to couple water splitting with the electrochemical oxidation of 5-HMF to FDCA. The hydrogen evolution reaction occurred at around 200 mV lower potential at the same current density if the biomass-derived alcohol was oxidized instead of the formation of molecular oxygen. Ni₂P nanoparticles supported on metallic nickel, hierarchically porous nickel- or nanocrystalline copper foam all achieved faradaic efficiencies close to unity and were robust for more than 10 h.¹⁰³⁻¹⁰⁵



Figure 5. Possible pathways for the synthesis of 2,5-furandicarboxylic acid (FDCA) from plant biomass. Intermediates are hydroxymethylfurfural (5-HMF), 2,5-diformylfuran (DFF), 5-hydroymethyl-2-furancarboxylic acid (HFCA) and 5-formyl-2-furancarboxylic acid (FFCA).



Figure 6. Chemical structures of chitin, chitosan and several derived N-containing compounds.

In most reports, the oxidation of the aldehyde group in the S-HMF substrate is faster than the alcohol oxidation. With this knowledge, the research community should focus more on the development of efficient catalysts for the relatively mild oxidation of alcohols to aldehydes using green reaction conditions such as water as solvent, oxygen as oxidant and a recyclable catalyst possibly based on non-noble metals. Photoand electrocatalytic processes can possibly provide more sustainable pathways but much more effort needs to be devoted to improve catalyst, reactor and finally process optimization.

N-Containing Chemicals. As mentioned earlier, chitin might be the most promising biomass source for the production of renewable N-containing chemicals due to its shear abundance and high nitrogen content of around 7% by weight. It has been estimated that 6–8 million tons of crustacean shell waste are produced annually, whereas bioproduction of insect shells are several orders of magnitude higher. Consequently, the concept of shell biorefinery, targeting the fractionation and utilization of various components in shells, was proposed and developed.¹⁰

Chitin is responsible for the high rigidity of insects' exoskeletons and crustaceans' shells and is structurally similar to cellulose despite the fact that the former has *N*-acetyl side chains on the C-2 position instead of a hydroxyl group. Normally, chitin occurs together with calcium carbonate and proteins and thus, fractionation is the inevitable first step for the generation of chitin-derived products.^{106,107} Tremendous efforts have been devoted to the utilization of chitin- and

chitosan-derived materials for a range of applications such as bioplastics,¹⁰⁸ biomedical applications^{109–111} and adsorbent materials for wastewater treatment^{112,113} but besides that, chitin is an ideal starting material for a range of heteroatom-containing chemicals. In the following, we will focus on N-containing chemicals from chitin, some of which are depicted in Figure 6, although many compounds that do not contain nitrogen are also available, such as acetic acid, several polyols, 5-HMF and other deaminated furan derivatives.^{114–120}

The most apparent product from chitin is N-acetylglucosamine via hydrolysis, but the harsh reaction conditions lead to a significant deacetylation.¹²¹ An enhanced process was developed to depolymerize chitin under retention of the acetyl group by a mechanochemical process with the addition of catalytic amounts of sulfuric acid. The ball-milled chitin-sulfuric acid mixture can then be hydrolyzed or methanolysed to yield two different products.¹²² Alternatively, the conversion of chitin into glucosamine can be selectively enhanced by using a mixture of aprotic polar solvents with water and acid amounts that are lower than previously reported. Cosolvents increased the glucosamine yield from less than 1% in pure water to up to 80% in a mixture of water and diethylene glycol diethyl ether.¹²³ A simple three-step procedure including a mechanocatalytic hydrolysis in the presence of sulfuric acid followed by a short high-temperature hydrolytic treatment and a hydrogenation with a heterogeneous ruthenium catalyst at pH 3.0 without any intermediate purification yielded the monomer in up to 52% yield.¹²⁴ Other sugar derivatives such as hydroxyethyl-2-amino-2-deoxyhexopyranoside (HADP) and 2Among the obtained N-containing compounds, high yields of N-acetylmonoethanolamine (NMEA), a possibly useful agent for CO₂ absorption among other applications, were achieved by reductive treatment over a noble metal catalyst. Although the conversion could not be enhanced by adding base such as sodium hydroxide, the selectivity toward certain products could be influenced significantly.¹²⁰ Besides aliphatic N-containing compounds, several aromatic heterocycles are available from chitin. One example is pyrrole, which was the main product (alongside pyrazine and pyridine) obtained after a short treatment of chitin at 325 °C using a basic aqueous solution. The addition of ammonia solution led to yields of up to 12 mol % from chitin within 1 min.¹¹⁸

Special emphasis should be placed on a very unique furanderivative: 3-acetamido-5-acetylfuran (3A5AF) available both from chitin and its monomeric sugar.^{127–131} The uncommon substitution pattern on the furan ring cannot be easily achieved by conventional synthetic methods from petrochemicals but is an essential structural motif in compounds such as proximicin antibiotics,¹³² pyrrolosine¹³³ and hyrtioseragamine A and B,¹³⁴ all of which are molecules with biological activity. Since the development of one-step production of 3A5AF from chitin and its monomer, many pathways for the synthesis of derived compounds have been proposed and some of them were realized recently, creating a chemical space with a range of Ncontaining chemicals¹³⁵ (Figure 7). Upon cleavage of the



Figure 7. Nitrogen-bearing chemical space derived from 3A5AF.

amide, a renewable amine (1) can be obtained whereas the selective reduction of the acetyl-group over the amide group can be achieved by reduction with sodium borohydride or catalyzed transfer hydrogenation affording product 2. Under certain conditions, the spontaneous formation of alkene 3 was observed probably due to the elimination of the formed alcohol. Upon the use of a stronger reductant such as lithium aluminum hydride, both carbonyls were reduced and the cyclic structure of the furan ring was opened.¹³⁶ The reaction of 3ASAF with several ketones such as acetone, butanone or

benzophenone surprisingly yielded a novel dihydrofuropyridine scaffold **6**, and produced **4** with formaldyhyde via hydroxymethylation of the furan ring. Reaction with acetaldehyde under the same conditions led to the formation of the cross-linked furan derivative **5**.¹³⁷

Another step toward shell biorefinery has been achieved recently by showing the complete synthesis of proximicin A, an antibiotic and cytostatic agent, from chitin as starting material. The currently existing synthetic procedure from crude oil contains a plethora of chemical reactions and multiple separation processes to even obtain the substituted furan derivative. Subsequently, another seven steps are required, which involves the use of dangerous, environmentally harmful reagents and solvents, and generates significant amounts of chemical waste due to the employment of protecting groups.^{132,138} In stark contrast, 3A5AF was obtained from chitin in one step with a single purification. In the following, six reactions are sufficient to synthesize proximicin A in a sustainable way utilizing green solvents and reagents that can be obtained from renewable resources (Figure 8).¹³⁹ Despite those growing efforts, more studies should be devoted to explore the full potential of this unique platform chemical.



Figure 8. Comparison of the synthesis of proximicin A from crude oil and chitin.

A polymer that is closely related to chitin is chitosan, obtained after deacetylation of 50% of the sugar units.¹⁴⁰ Due to the free amine functionality, the properties are very different from chitin regarding the basicity, solubility in water and chemical reactivity. Processes for the synthesis of N-containing chemicals from chitosan are so far limited to the depolymerization and formation of ethanolamine¹²⁰ and glucosamine¹⁴¹ (Figure 6) using a catalytic reductive or a microwave-assisted method, respectively.

The fractionation and pretreatment of plant biomass for cellulose, hemicellulose and lignin have already been well-developed.³² In contrast, much less has been achieved for the fractionation of shells. Current practice heavily relies on a two-step process, a first demineralization step using strong mineral acids followed by a deproteinization step employing strong bases, resulting in huge amounts of chemical waste and high cost of the product.^{142–144} The efficient use of chitin as starting material for the synthesis of N-containing chemicals is therefore

limited by the detrimental environmental impact of the fractionation process. There is an urgent need for the development of more environmentally benign and cost-efficient processes. One example has been reported recently, where a simple pretreatment of prawn shells in hot glycerol improved the removal of both the protein and the mineral fraction by citric acid.¹⁴⁵ Although this method might already prove more favorable than the conventional industrial method, a procedure employing nonstoichiometric amounts of organic or even mineral acids is highly desirable in the future.

Amines from Renewable Oxygenates. Almost all kinds of O-containing functional groups have been shown to be convertible into their N-containing counterparts using ammonia as sole nitrogen source and possibly oxygen as abundant and atom-economic oxidant.¹⁴⁶ Previous developments in the synthesis of biobased amines have been reviewed in detail¹⁴⁷ and thus only selected most recent developments will be discussed here (Figure 9).



Figure 9. Recent developments in the synthesis of organic nitrogencontaining compounds from oxygenates using ammonia; oxygen atoms that are removed or replaced during the process are depicted in red.

Due to the versatility and abundance of renewable alcohols and the attractiveness of primary mono- and diamines as organic building blocks or for polymer applications, efficient interconversions need to be developed. It is commonly postulated that the amination of alcohols occurs via a "hydrogen borrowing" mechanism where a carbonyl compound is formed as an intermediate which is then converted into the imine and subsequently reduced to the amine.^{148,149} Therefore, it seems to be attractive to start with a carbonyl compound and employ a reducing agent to produce alcohols.^{150,151} One example is the production of ethanolamine from cellulose in a two-step process with a yield above 10% using hydrogen gas as reducing agent and a heterogeneous ruthenium catalyst for the reductive amination.¹⁵² Inspired by the naturally occurring aldolase enzyme, the direct reductive aminolysis of glucose toward several short-chained amines with carbon yields up to 87% have been achieved.^{153,154} Likewise, the amination of isomannide, a renewable sugar-derivative, in water using a

heterogeneous ruthenium catalyst has been demonstrated. This process can be extended to the conversion of other biogenic alcohols such as ethanol or hexane-1,6-diol.¹⁵⁵

Most recently, a process has been developed for the production of acrylonitrile, a valuable substrate for the synthesis of a range of resins, polymers and carbon fibers, from renewable resources including lignocellulosic biomass. A fermentation process was used to produce an organic acid from sugars, which was then converted into acrylic acid, esterified and converted into the respective nitrile with ammonia using an inexpensive catalyst. Life cycle assessment showed that on a very large scale, the produced acrylonitrile reached a price below 1\$/lb, avoiding the link with sometimes volatile fossil fuel prices. It also represents a safer process and could thus become a realistic alternative to the prevailing Sohio process. Extending the substrate scope to more expensive nitriles might render the process economically competitive even at smaller scales.¹⁵⁶

The synthesis of another N-containing aromatic heterocyclic compound, 2-methylpyrazine, has been realized from different renewable sugars and ammonia using catalytic amounts of tungsten clusters. Glucose, fructose and xylose all turned out to be valuable starting materials. Optimization studies showed that close to 30 mol % yield can be achieved, rendering the process attractive compared to conventional routes starting from petrochemicals.¹⁵⁷ A similar product (2-hydroxymethyl-5-methylpyrazine) was synthesized from dihydroxyacetone and diammonium hydrogen phosphate without the use of a catalyst with up to 72% yield.¹⁵⁸

Vanillic acid has been shown to be convertible into two isomeric pyridine dicarboxylic acids using ammonium chloride as nitrogen source by modifying the protocatechuate pathway of *Rhodococcus jostii* RHA1. Wheat straw biomass can be used directly together with a minimal growth medium for the synthesis of the diacids. The close resemblance of the products to terephthalic acid hint at the potential use for polymer applications although the changed electronic structure in the aromatic ring and the enhanced coordination abilities of the pyridine moiety should lead to different properties.¹⁵⁹

One crucial aspect for the conversion of O-containing functional groups into N-containing ones is atom efficiency. Ideally, ammonia can be used leading to the formation of water as the only byproduct. Furthermore, robust and general procedures that apply to the widest possible range of oxygenates preferably employing a recyclable and cheap catalyst need to be developed and employed for products that are already in use and manufactured from fossil fuel resources.

S-CONTAINING CHEMICALS

Sulfur is a ubiquitous element in chemical industry. Natural resources for sulfur-containing chemicals are widely unexplored even though some plant biomass, especially algae, carry high amounts of sulfated polysaccharides. In the current endeavor to produce biofuels from algal biomass, harnessing a part of that biomass for the synthesis of renewable sulfur-containing chemicals could become attractive. Some of those sulfated polysugars are used widely as thickening agent, and as anticoagulants, anticancer or anti-inflammatory agents.¹⁶⁰ Carrageenans and fucoidan belong to the more commonly used polysaccharides but their full chemical potential has yet to be developed and evaluated (Figure 10). Fucoidan, for example, is primarily found in brown algae and brown seaweed making up around 10–20 wt % of the dry algae weight.¹⁶¹ Despite its perceived health benefits, any large scale applications are



Figure 10. Proposed sulfur-bearing chemical space derived from sulfated polysaccharides.

currently limited by the horrendous price. Based on the structural similarity to previously mentioned polysaccharides, the synthesis of similar products including oligomeric or monomeric sugars and sugar-derivatives and different heterocyclic chemicals such as thiophenes and sulfated furans are envisioned from both carrageenans and fucoidan. Some of those compounds can be expected to find applications in the synthesis of organic electronics,¹⁶² as surfactants¹⁶³ or for biomedical applications.¹⁶⁴ Any work along this line will motivate the development of more efficient processes for the isolation and purification of sulfur-containing polysaccharides which are essential first steps to increase the economic viability.

Other strategies to convert renewable feedstock into Scontaining chemicals either employ organic compounds containing sulfur, gaseous SO₃ or elemental sulfur. Most studies on renewable S-containing products emphasize the production of polymers^{165–167} or sorbent materials,^{168,169} which lie beyond the scope of this perspective. It has been shown that lubricating oils can be synthesized from different vegetable oils in a two-step process using a photochemical approach. Thiyl radicals bind to unsaturated bonds in the fatty acids and a transesterification step yields the sulfur-containing esters (Figure 11).^{170,171} In principle, renewable alcohols can be used for the transesterification step and the development of procedures yielding thiols from biogenic alcohols would be beneficial.

It was shown recently that a sulfonated surfactant based on furan and fatty acids, two completely renewable components, show excellent performance in forming micelles in hard water and at low temperatures (Figure 12). Several common fatty acids ranging from seven to 18 carbon atoms were linked to furan by a heterogeneously catalyzed Friedel-Crafts acylation, which then underwent sulfonation under mild conditions. Common surfactant mixtures contain 6-22% of chelating agents depending on water property, to avoid the inhibition effect of divalent metal cations in micelle formation. This fraction is no longer necessary for the new material, due to the extraordinary micelle formation ability of oleo-furan-based surfactants even in the presence of metal cations. The exceptional quality of the renewable surfactants and the low price of the feedstock could possibly render the process economically competitive and enable the use of ecologically more benign common products.¹⁷²

P-CONTAINING CHEMICALS

Phosphorus is an essential element in living organisms, but abundant sources of *P*-containing biopolymers are widely unknown. Nevertheless, biological membranes are known to contain a high fraction of phosphorus as part of their



Figure 11. Synthesis of sulfur-containing lubricating oils from mostly renewable resources.



Figure 12. Structure and possible utilization of surfactants almost entirely based on renewable feedstock. (Reprinted with permission from ref 172.)

membrane lipids such as phosphatidylcholine (such as shown in Figure 2). Some studies have been devoted to the hydrolysis of those lipids into various phosphorus-containing chemicals.^{173–175} Although organophosphorus compounds do not belong to the most common intermediates and products in chemical industries, they are used as pesticides,¹⁷⁶ herbicides¹⁷⁷ or in certain medical applications.¹⁷⁸ To find biorenewable alternatives for those fossil-fuel-derived compounds, novel sources of phosphorus from biomass need to be explored and conversion strategies have to be developed.

CONCLUSION

In summary, processes for the synthesis of heteroatomcontaining chemicals from renewable resources are in steady development with many promising systems been presented. Though renewable oxygen-containing chemicals have been most extensively studied, more efforts are required to be

devoted to other heteroatom-containing chemicals to explore potential new platform chemicals and associated further transformation into fine chemicals. Generally, we need to make use of chemical functionalities within biobased feedstock as much as possible, as any further conversions inevitably require energy, resources and generate waste. If unavoidable, any additional treatments need to be done in accordance with the 12 principles of green chemistry especially considering the use of catalytic reagents, atom economic procedures and energy efficiency. Along this line, novel procedures employing electroor photochemistry can be beneficial and should be developed further for a broader range of biorenewable compounds.^{179,180}

For many heteroatom-containing biopolymers, more efficient and environmentally benign fractionation methods to extract them from the native state, and enhanced strategies for the transformation of separated components into a variety of valueadded products are needed. In the case of lignocellulosic biomass, various strategies to stabilize the different components during biomass treatment such as the lignin-first approach emerged. Similarly, mild methods have to be developed for the fractionation of shell biomass into its components. The same challenges await S- and P-containing biopolymers once good sources have been discovered and the range of obtainable molecules has been extended.

As highlighted recently, the biorenewable feedstock has to be utilized entirely in order to make biorefinery competitive with current fossil fuel-based refinery schemes.¹⁸¹ All those processes should then be assessed based on a thorough technological, economic and environmental evaluation before they are ready for scaling up development.¹⁸² All current technological advances and all available products from a biomass resource need to be considered in an integrated manner estimating their global warming potential, the cumulative energy demand and operating costs. In most cases, a particular feed split ratio for several products probably has the highest feasibility potential from different aspects.¹⁸³ The interdependence between several products of a biorefinery has already become very clear in some cases, such as biodiesel production, in which it is only economically viable and environmentally beneficial if processes for glycerol upgrading can be integrated. Beyond that, computational tools should be developed that enable decision-makers acting in the most reasonable way possible given a certain set of conditions that inevitably will deviate massively throughout different countries and regions.

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REFERENCES

(1) Schwach, P.; Pan, X.; Bao, X. Direct Conversion of Methane to Value-Added Chemicals over Heterogeneous Catalysts: Challenges and Prospects. *Chem. Rev.* 2017, *117* (13), 8497–8520.

(2) McGrath, N. A.; Brichacek, M.; Njardarson, J. T. A graphical journey of innovative organic architectures that have improved our lives. *J. Chem. Educ.* **2010**, *87* (12), 1348–1349.

(3) Atwood, D.; Paisley-Jones, C. *Pesticides industry sales and usage* 2008–2012 market estimates; U.S. Environmental Protection Agency: Washington, DC, 2017.

(4) Huber, G. W.; Iborra, S.; Corma, A. Synthesis of Transportation Fuels from Biomass: Chemistry, Catalysts, and Engineering. *Chem. Rev.* **2006**, *106* (9), 4044–4098.

(5) Li, C.; Zhao, X.; Wang, A.; Huber, G. W.; Zhang, T. Catalytic Transformation of Lignin for the Production of Chemicals and Fuels. *Chem. Rev.* **2015**, *115* (21), 11559–11624.

(6) Upton, B. M.; Kasko, A. M. Strategies for the Conversion of Lignin to High-Value Polymeric Materials: Review and Perspective. *Chem. Rev.* **2016**, *116* (4), 2275–2306.

(7) Ragauskas, A. J.; Beckham, G. T.; Biddy, M. J.; Chandra, R.; Chen, F.; Davis, M. F.; Davison, B. H.; Dixon, R. A.; Gilna, P.; Keller, M.; Langan, P.; Naskar, A. K.; Saddler, J. N.; Tschaplinski, T. J.; Tuskan, G. A.; Wyman, C. E. Lignin valorization: improving lignin processing in the biorefinery. *Science* **2014**, *344* (6185), 709–719.

(8) Kumar, M. N. V. R. A review of chitin and chitosan applications. *React. Funct. Polym.* **2000**, *46* (1), 1–27.

(9) Chen, X.; Yang, H.; Yan, N. Shell Biorefinery: Dream or Reality? *Chem.—Eur. J.* **2016**, 47 (46), 1–21.

(10) Yan, N.; Chen, X. Don't waste seafood waste: Turning cast-off shells into nitrogen-rich chemicals would benefit economies and the environment. *Nature* **2015**, *524* (7564), 155–158.

(11) Chheda, J. N.; Huber, G. W.; Dumesic, J. A. Liquid-phase catalytic processing of biomass-derived oxygenated hydrocarbons to fuels and chemicals. *Angew. Chem., Int. Ed.* **2007**, *46* (38), 7164–7183.

(12) Alonso, D. M.; Bond, J. Q.; Dumesic, J. A. Catalytic conversion of biomass to biofuels. *Green Chem.* **2010**, *12* (9), 1493–1513.

(13) Gallezot, P. Conversion of biomass to selected chemical products. *Chem. Soc. Rev.* **2012**, *41* (4), 1538–1558.

(14) Huber, G. W.; Corma, A. Synergies between Bio-and Oil Refineries for the Production of Fuels from Biomass. *Angew. Chem., Int. Ed.* **2007**, *46* (38), 7184–7201.

(15) Mortensen, P. M.; Grunwaldt, J.-D.; Jensen, P. A.; Knudsen, K. G.; Jensen, A. D. A review of catalytic upgrading of bio-oil to engine fuels. *Appl. Catal., A* **2011**, *407* (1), 1–19.

(16) Xu, C.; Arancon, R. A. D.; Labidi, J.; Luque, R. Lignin depolymerisation strategies: towards valuable chemicals and fuels. *Chem. Soc. Rev.* **2014**, 43 (22), 7485–7500.

(17) Li, H.; Riisager, A.; Saravanamurugan, S.; Pandey, A.; Sangwan, R. S.; Yang, S.; Luque, R. Carbon-Increasing Catalytic Strategies for Upgrading Biomass into Energy-Intensive Fuels and Chemicals. *ACS Catal.* **2018**, *8*, 148–187.

(18) Zhang, J.; Zhao, C. Development of a Bimetallic Pd-Ni/HZSM-5 Catalyst for the Tandem Limonene Dehydrogenation and Fatty Acid Deoxygenation to Alkanes and Arenes for Use as Biojet Fuel. ACS Catal. 2016, 6 (7), 4512–4525.

(19) Song, W.; Liu, Y.; Barath, E.; Zhao, C.; Lercher, J. A. Synergistic effects of Ni and acid sites for hydrogenation and C-O bond cleavage of substituted phenols. *Green Chem.* **2015**, *17* (2), 1204–1218.

(20) Kong, J.; He, M.; Lercher, J. A.; Zhao, C. Direct production of naphthenes and paraffins from lignin. *Chem. Commun.* **2015**, *51* (99), 17580–17583.

(21) Che, P.; Lu, F.; Zhang, J.; Huang, Y.; Nie, X.; Gao, J.; Xu, J. Catalytic selective etherification of hydroxyl groups in 5-hydroxymethylfurfural over H4SiW12O40/MCM-41 nanospheres for liquid fuel production. *Bioresour. Technol.* **2012**, *119*, 433–436.

(22) Puga, A. V. Photocatalytic production of hydrogen from biomass-derived feedstocks. *Coord. Chem. Rev.* 2016, 315, 1–66.

(23) Serrano-Ruiz, J. C.; Luque, R.; Sepúlveda-Escribano, A. Transformations of biomass-derived platform molecules: from high added-value chemicals to fuels via aqueous-phase processing. *Chem. Soc. Rev.* **2011**, *40* (11), 5266–5281.

(24) Alonso, D. M.; Wettstein, S. G.; Dumesic, J. A. Gammavalerolactone, a sustainable platform molecule derived from lignocellulosic biomass. *Green Chem.* **2013**, *15* (3), 584–595.

(25) Sheldon, R. A. Green and sustainable manufacture of chemicals from biomass: state of the art. *Green Chem.* **2014**, *16* (3), 950–963.

(26) Sun, Z.; Fridrich, B.; de Santi, A.; Elangovan, S.; Barta, K. Bright Side of Lignin Depolymerization: Toward New Platform Chemicals. *Chem. Rev.* **2018**, *118* (2), 614–678.

(27) Mika, L. T.; Cséfalvay, E.; Németh, A. Catalytic Conversion of Carbohydrates to Initial Platform Chemicals: Chemistry and Sustainability. *Chem. Rev.* **2018**, *118* (2), 505–613.

(28) Chen, J.; Lu, F.; Zhang, J.; Yu, W.; Wang, F.; Gao, J.; Xu, J. Immobilized Ru Clusters in Nanosized Mesoporous Zirconium Silica for the Aqueous Hydrogenation of Furan Derivatives at Room Temperature. *ChemCatChem* **2013**, 5 (10), 2822–2826.

(29) Corma, A.; Iborra, S.; Velty, A. Chemical routes for the transformation of biomass into chemicals. *Chem. Rev.* 2007, 107 (6), 2411–2502.

(30) Zakzeski, J.; Bruijnincx, P. C.; Jongerius, A. L.; Weckhuysen, B. M. The catalytic valorization of lignin for the production of renewable chemicals. *Chem. Rev.* **2010**, *110* (6), 3552–3599.

(31) Vispute, T. P.; Zhang, H.; Sanna, A.; Xiao, R.; Huber, G. W. Renewable chemical commodity feedstocks from integrated catalytic processing of pyrolysis oils. *Science* **2010**, 330 (6008), 1222–1227.

(32) Schutyser, W.; Renders, T.; Van den Bosch, S.; Koelewijn, S.-F.; Beckham, G. T.; Sels, B. F. Chemicals from lignin: an interplay of lignocellulose fractionation, depolymerisation, and upgrading. *Chem. Soc. Rev.* **2018**, *47*, 852–908.

(33) Gall, D. L.; Ralph, J.; Donohue, T. J.; Noguera, D. R. Biochemical transformation of lignin for deriving valued commodities from lignocellulose. *Curr. Opin. Biotechnol.* **2017**, *45*, 120–126.

(34) Beckham, G. T.; Johnson, C. W.; Karp, E. M.; Salvachúa, D.; Vardon, D. R. Opportunities and challenges in biological lignin valorization. *Curr. Opin. Biotechnol.* **2016**, *42*, 40–53.

(35) Renders, T.; Van den Bosch, S.; Koelewijn, S.-F.; Schutyser, W.; Sels, B. F. Lignin-first biomass fractionation: the advent of active stabilisation strategies. *Energy Environ. Sci.* **2017**, *10* (7), 1551–1557.

(36) Shuai, L.; Amiri, M. T.; Questell-Santiago, Y. M.; Héroguel, F.; Li, Y.; Kim, H.; Meilan, R.; Chapple, C.; Ralph, J.; Luterbacher, J. S. Formaldehyde stabilization facilitates lignin monomer production during biomass depolymerization. *Science* **2016**, *354* (6310), *329*–333.

(37) Lan, W.; Amiri, M. T.; Hunston, C. M.; Luterbacher, J. Protection Group Effects during α , γ -Diol Lignin Stabilization Promote High-Selectivity Monomer Production. *Angew. Chem., Int. Ed.* **2018**, *57* (5), 1356–1360.

(38) Chen, X.; Kuhn, E.; Jennings, E. W.; Nelson, R.; Tao, L.; Zhang, M.; Tucker, M. P. DMR (deacetylation and mechanical refining) processing of corn stover achieves high monomeric sugar concentrations (230 g L- 1) during enzymatic hydrolysis and high ethanol concentrations (> 10% v/v) during fermentation without hydrolysate

purification or concentration. *Energy Environ. Sci.* 2016, 9 (4), 1237–1245.

(39) Chen, X.; Wang, W.; Ciesielski, P.; Trass, O.; Park, S.; Tao, L.; Tucker, M. P. Improving sugar yields and reducing enzyme loadings in the deacetylation and mechanical refining (DMR) process through multistage disk and Szego refining and corresponding technoeconomic analysis. ACS Sustainable Chem. Eng. 2016, 4 (1), 324–333.

(40) Anderson, E. M.; Stone, M. L.; Katahira, R.; Reed, M.; Beckham, G. T.; Román-Leshkov, Y. Flowthrough Reductive Catalytic Fractionation of Biomass. *Joule* **2017**, *1* (3), 613–622.

(41) Rinaldi, R. A Tandem for Lignin-First Biorefinery. *Joule* 2017, 1 (3), 427–428.

(42) Ferrini, P.; Rezende, C. A.; Rinaldi, R. Catalytic Upstream Biorefining through Hydrogen Transfer Reactions: Understanding the Process from the Pulp Perspective. *ChemSusChem* **2016**, 9 (22), 3171–3180.

(43) Shuai, L.; Saha, B. Towards high-yield lignin monomer production. *Green Chem.* 2017, *19* (16), 3752–3758.

(44) Kessler, M.; Woodward, R.; Wong, N.; Rinaldi, R. Kinematic Modelling of Mechanocatalytic Depolymerization of α -Cellulose and Beechwood. *ChemSusChem* **2018**, *11* (3), 552–561.

(45) Shetty, M.; Buesser, B.; Román-Leshkov, Y.; Green, W. H. Computational Investigation on Hydrodeoxygenation (HDO) of Acetone to Propylene on α -MoO3 (010) Surface. *J. Phys. Chem. C* **2017**, *121* (33), 17848–17855.

(46) Ferguson, G. A.; Vorotnikov, V.; Wunder, N.; Clark, J.; Gruchalla, K.; Bartholomew, T.; Robichaud, D. J.; Beckham, G. T. Ab Initio Surface Phase Diagrams for Coadsorption of Aromatics and Hydrogen on the Pt(111) Surface. *J. Phys. Chem. C* **2016**, *120* (46), 26249–26258.

(47) Gu, G. H.; Mullen, C. A.; Boateng, A. A.; Vlachos, D. G. Mechanism of dehydration of phenols on noble metals via first-principles microkinetic modeling. *ACS Catal.* **2016**, *6* (5), 3047–3055.

(48) Réocreux, R.; Hamou, C. A. O.; Michel, C.; Giorgi, J. B.; Sautet, P. Decomposition Mechanism of Anisole on Pt (111): Combining Single-Crystal Experiments and First-Principles Calculations. *ACS Catal.* **2016**, *6* (12), 8166–8178.

(49) Kennema, M.; de Castro, I. B. D.; Meemken, F.; Rinaldi, R. Liquid-Phase H-Transfer from 2-Propanol to Phenol on Raney Ni: Surface Processes and Inhibition. *ACS Catal.* **2017**, *7* (4), 2437–2445.

(50) Shetty, M.; Murugappan, K.; Green, W. H.; Román-Leshkov, Y. Structural properties and reactivity trends of molybdenum oxide catalysts supported on zirconia for the hydrodeoxygenation of anisole. *ACS Sustainable Chem. Eng.* **2017**, *5* (6), 5293–5301.

(51) Meier, M. A. R.; Metzger, J. O.; Schubert, U. S. Plant oil renewable resources as green alternatives in polymer science. *Chem. Soc. Rev.* **2007**, *36* (11), 1788–1802.

(52) Gandini, A. Polymers from renewable resources: a challenge for the future of macromolecular materials. *Macromolecules* **2008**, *41* (24), 9491–9504.

(53) Koelewijn, S. F.; Van den Bosch, S.; Renders, T.; Schutyser, W.; Lagrain, B.; Smet, M.; Thomas, J.; Dehaen, W.; Van Puyvelde, P.; Witters, H.; Sels, B. F. Sustainable bisphenols from renewable softwood lignin feedstock for polycarbonates and cyanate ester resins. *Green Chem.* **2017**, *19* (11), 2561–2570.

(54) Zhu, Y.; Romain, C.; Williams, C. K. Sustainable polymers from renewable resources. *Nature* **2016**, *540* (7633), 354–362.

(55) Niu, W.; Draths, K. M.; Frost, J. W. Benzene-Free Synthesis of Adipic Acid. *Biotechnol. Prog.* 2002, 18 (2), 201–211.

(56) Beerthuis, R.; Rothenberg, G.; Shiju, N. R. Catalytic routes towards acrylic acid, adipic acid and *e*-caprolactam starting from biorenewables. *Green Chem.* **2015**, *17* (3), 1341–1361.

(57) Deng, Y.; Ma, L.; Mao, Y. Biological production of adipic acid from renewable substrates: Current and future methods. *Biochem. Eng. J.* **2016**, *105*, 16–26.

(58) Lee, J.; Saha, B.; Vlachos, D. G. Pt catalysts for efficient aerobic oxidation of glucose to glucaric acid in water. *Green Chem.* **2016**, *18* (13), 3815–3822.

(59) Li, X.; Wu, D.; Lu, T.; Yi, G.; Su, H.; Zhang, Y. Highly Efficient Chemical Process To Convert Mucic Acid into Adipic Acid and DFT Studies of the Mechanism of the Rhenium-Catalyzed Deoxydehydration. *Angew. Chem.* **2014**, *126* (16), 4284–4288.

(60) Wright, W. R.; Palkovits, R. Development of heterogeneous catalysts for the conversion of levulinic acid to γ -valerolactone. *ChemSusChem* **2012**, 5 (9), 1657–1667.

(61) Wong, P. K.; Li, C.; Stubbs, L.; Van Meurs, M.; Kumbang, D. G. A.; Lim, S. C. Y.; Drent, E. Patent WO 2012/134397, 2012.

(62) Buntara, T.; Noel, S.; Phua, P. H.; Melián-Cabrera, I.; de Vries, J. G.; Heeres, H. J. Caprolactam from renewable resources: catalytic conversion of 5-hydroxymethylfurfural into caprolactone. *Angew. Chem., Int. Ed.* **2011**, *50* (31), 7083–7087.

(63) Buntara, T.; Noel, S.; Phua, P. H.; Melián-Cabrera, I.; de Vries, J. G.; Heeres, H. J. From 5-hydroxymethylfurfural (HMF) to polymer precursors: catalyst screening studies on the conversion of 1, 2, 6-hexanetriol to 1, 6-hexanediol. *Top. Catal.* **2012**, *55* (7–10), 612–619. (64) Motagamwala, A. H.; Won, W.; Sener, C.; Alonso, D. M.; Maravelias, C. T.; Dumesic, J. A. Toward biomass-derived renewable plastics: Production of 2,5-furandicarboxylic acid from fructose. *Sci. Adv.* **2018**, *4* (1), No. eaap9722.

(65) Gilkey, M. J.; Mironenko, A. V.; Vlachos, D. G.; Xu, B. Adipic Acid Production via Metal-Free Selective Hydrogenolysis of Biomass-Derived Tetrahydrofuran-2, 5-Dicarboxylic Acid. *ACS Catal.* **2017**, 7 (10), 6619–6634.

(66) Strassberger, Z.; Tanase, S.; Rothenberg, G. The pros and cons of lignin valorisation in an integrated biorefinery. *RSC Adv.* **2014**, *4* (48), 25310–25318.

(67) Liu, H.; Jiang, T.; Han, B.; Liang, S.; Zhou, Y. Selective phenol hydrogenation to cyclohexanone over a dual supported Pd–Lewis acid catalyst. *Science* **2009**, *326* (5957), 1250–1252.

(68) Vardon, D. R.; Franden, M. A.; Johnson, C. W.; Karp, E. M.; Guarnieri, M. T.; Linger, J. G.; Salm, M. J.; Strathmann, T. J.; Beckham, G. T. Adipic acid production from lignin. *Energy Environ. Sci.* **2015**, 8 (2), 617–628.

(69) Johnson, C. W.; Salvachúa, D.; Khanna, P.; Smith, H.; Peterson, D. J.; Beckham, G. T. Enhancing muconic acid production from glucose and lignin-derived aromatic compounds via increased protocatechuate decarboxylase activity. *Metab. Eng. Commun.* **2016**, *3*, 111–119.

(70) Vardon, D. R.; Rorrer, N. A.; Salvachúa, D.; Settle, A. E.; Johnson, C. W.; Menart, M. J.; Cleveland, N. S.; Ciesielski, P. N.; Steirer, K. X.; Dorgan, J. R.; Beckham, G. T. cis, cis-Muconic acid: separation and catalysis to bio-adipic acid for nylon-6, 6 polymerization. *Green Chem.* **2016**, *18* (11), 3397–3413.

(71) Rodriguez, A.; Salvachúa, D.; Katahira, R.; Black, B. A.; Cleveland, N. S.; Reed, M.; Smith, H.; Baidoo, E. E. K.; Keasling, J. D.; Simmons, B. A.; Beckham, G. T.; Gladden, J. M. Base-Catalyzed Depolymerization of Solid Lignin-Rich Streams Enables Microbial Conversion. ACS Sustainable Chem. Eng. **2017**, 5 (9), 8171–8180.

(72) Matthiesen, J. E.; Carraher, J. M.; Vasiliu, M.; Dixon, D. A.; Tessonnier, J.-P. Electrochemical conversion of muconic acid to biobased diacid monomers. *ACS Sustainable Chem. Eng.* **2016**, *4* (6), 3575–3585.

(73) Tomás, R. A. F.; Bordado, J. C. M.; Gomes, J. F. P. p-Xylene oxidation to terephthalic acid: a literature review oriented toward process optimization and development. *Chem. Rev.* **2013**, *113* (10), 7421–7469.

(74) Settle, A. E.; Berstis, L.; Rorrer, N. A.; Roman-Leshkóv, Y.; Beckham, G. T.; Richards, R. M.; Vardon, D. R. Heterogeneous Diels–Alder catalysis for biomass-derived aromatic compounds. *Green Chem.* **2017**, *19*, 3468–3492.

(75) Cho, H. J.; Ren, L.; Vattipalli, V.; Yeh, Y. H.; Gould, N.; Xu, B.; Gorte, R. J.; Lobo, R.; Dauenhauer, P. J.; Tsapatsis, M.; Fan, W. Renewable p-Xylene from 2, 5-Dimethylfuran and Ethylene Using Phosphorus-Containing Zeolite Catalysts. *ChemCatChem* **2017**, *9* (3), 398–402.

(76) Li, C.; Dai, T.; Li, L.; Zhao, Z. K.; Zhang, B.; Cong, Y.; Wang, A. Selective production of renewable para-xylene via tungsten carbide-

catalyzed atom-economic cascade reactions. Angew. Chem., Int. Ed. 2018, 57, 1808–1812.

(77) Pang, J.; Zheng, M.; Sun, R.; Wang, A.; Wang, X.; Zhang, T. Synthesis of ethylene glycol and terephthalic acid from biomass for producing PET. *Green Chem.* **2016**, *18* (2), 342–359.

(78) Pacheco, J. J.; Davis, M. E. Synthesis of terephthalic acid via Diels-Alder reactions with ethylene and oxidized variants of 5-hydroxymethylfurfural. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111* (23), 8363–8367.

(79) Lu, R.; Lu, F.; Chen, J.; Yu, W.; Huang, Q.; Zhang, J.; Xu, J. Production of Diethyl Terephthalate from Biomass-Derived Muconic Acid. *Angew. Chem.* **2016**, *128* (1), 257–261.

(80) Saraçi, E.; Wang, L.; Theopold, K. H.; Lobo, R. F. Bioderived Muconates by Cross-Metathesis and Their Conversion into Terephthalates. *ChemSusChem* **2018**, *11* (4), 773–780.

(81) Bai, Z.; Phuan, W. C.; Ding, J.; Heng, T. H.; Luo, J.; Zhu, Y. Production of Terephthalic Acid from Lignin-Based Phenolic Acids by a Cascade Fixed-Bed Process. ACS Catal. **2016**, 6 (9), 6141–6145.

(82) Zhang, Z.; Deng, K. Recent Advances in the Catalytic Synthesis of 2,5-Furandicarboxylic Acid and Its Derivatives. *ACS Catal.* **2015**, 5 (11), 6529–6544.

(83) Li, H.; Yang, S.; Riisager, A.; Pandey, A.; Sangwan, R. S.; Saravanamurugan, S.; Luque, R. Zeolite and zeotype-catalysed transformations of biofuranic compounds. *Green Chem.* **2016**, *18* (21), 5701–5735.

(84) De Clercq, R.; Dusselier, M.; Sels, B. F. Heterogeneous catalysis for bio-based polyester monomers from cellulosic biomass: advances, challenges and prospects. *Green Chem.* **2017**, *19* (21), 5012–5040.

(85) Šiankevich, S.; Savoglidis, G.; Fei, Z.; Laurenczy, G.; Alexander, D. T. L.; Yan, N.; Dyson, P. J. A novel platinum nanocatalyst for the oxidation of 5-Hydroxymethylfurfural into 2,5-Furandicarboxylic acid under mild conditions. *J. Catal.* **2014**, *315*, 67–74.

(86) Choudhary, H.; Ebitani, K. Hydrotalcite-supported PdPtcatalyzed Aerobic Oxidation of 5-Hydroxymethylfurfural to 2,5-Furandicarboxylic Acid in Water. *Chem. Lett.* **2016**, *45* (6), 613–615.

(87) Antonyraj, C. A.; Huynh, N. T. T.; Park, S.-K.; Shin, S.; Kim, Y. J.; Kim, S.; Lee, K.-Y.; Cho, J. K. Basic anion-exchange resin (AER)-supported Au-Pd alloy nanoparticles for the oxidation of 5-hydroxymethyl-2-furfural (HMF) into 2,5-furan dicarboxylic acid (FDCA). *Appl. Catal., A* **2017**, 547, 230–236.

(88) Gupta, K.; Rai, R. K.; Singh, S. K. Catalytic aerial oxidation of 5hydroxymethyl-2-furfural to furan-2,5-dicarboxylic acid over Ni-Pd nanoparticles supported on Mg(OH)2 nanoflakes for the synthesis of furan diesters. *Inorg. Chem. Front.* **2017**, *4* (5), 871–880.

(89) Wang, Q.; Hou, W.; Li, S.; Xie, J.; Li, J.; Zhou, Y.; Wang, J. Hydrophilic mesoporous poly(ionic liquid)-supported Au-Pd alloy nanoparticles towards aerobic oxidation of 5-hydroxymethylfurfural to 2,5-furandicarboxylic acid under mild conditions. *Green Chem.* **2017**, *19* (16), 3820–3830.

(90) Chen, C.; Li, X.; Wang, L.; Liang, T.; Wang, L.; Zhang, Y.; Zhang, J. Highly Porous Nitrogen- and Phosphorus-Codoped Graphene: An Outstanding Support for Pd Catalysts to Oxidize 5-Hydroxymethylfurfural into 2,5-Furandicarboxylic Acid. ACS Sustainable Chem. Eng. 2017, 5 (12), 11300–11306.

(91) Donoeva, B.; Masoud, N.; de Jongh, P. E. Carbon Support Surface Effects in the Gold-Catalyzed Oxidation of 5-Hydroxymethylfurfural. *ACS Catal.* **2017**, *7* (7), 4581–4591.

(92) Gao, T.; Gao, T.; Fang, W.; Cao, Q. Base-free aerobic oxidation of 5-hydroxymethylfurfural to 2,5-furandicarboxylic acid in water by hydrotalcite-activated carbon composite supported gold catalyst. *Mol. Catal* **2017**, 439, 171–179.

(93) Mishra, D. K.; Lee, H. J.; Kim, J.; Lee, H.-S.; Cho, J. K.; Suh, Y.-W.; Yi, Y.; Kim, Y. J. $MnCo_2O_4$ spinel supported ruthenium catalyst for air-oxidation of HMF to FDCA under aqueous phase and base-free conditions. *Green Chem.* **2017**, *19* (7), 1619–1623.

(94) Yang, Z.; Qi, W.; Su, R.; He, Z. Selective Synthesis of 2,5-Diformylfuran and 2,5-Furandicarboxylic Acid from 5-Hydroxymethylfurfural and Fructose Catalyzed by Magnetically Separable Catalysts. *Energy Fuels* **2017**, *31* (1), 533–541. (95) Yan, D.; Xin, J.; Zhao, Q.; Gao, K.; Lu, X.; Wang, G.; Zhang, S. Fe-Zr-O catalyzed base-free aerobic oxidation of 5-HMF to 2,5-FDCA as a bio-based polyester monomer. *Catal. Sci. Technol.* **2018**, *8* (1), 164–175.

(96) Zuo, X.; Venkitasubramanian, P.; Busch, D. H.; Subramaniam, B. Optimization of Co/Mn/Br-Catalyzed Oxidation of 5-Hydroxymethylfurfural to Enhance 2,5-Furandicarboxylic Acid Yield and Minimize Substrate Burning. ACS Sustainable Chem. Eng. **2016**, 4 (7), 3659–3668.

(97) Watanabe, H.; Asano, S.; Fujita, S.-i.; Yoshida, H.; Arai, M. Nitrogen-Doped, Metal-Free Activated Carbon Catalysts for Aerobic Oxidation of Alcohols. *ACS Catal.* **2015**, *5* (5), 2886–2894.

(98) Nguyen, C. V.; Liao, Y.-T.; Kang, T.-C.; Chen, J. E.; Yoshikawa, T.; Nakasaka, Y.; Masuda, T.; Wu, K. C. W. A metal-free, high nitrogen-doped nanoporous graphitic carbon catalyst for an effective aerobic HMF-to-FDCA conversion. *Green Chem.* **2016**, *18* (22), 5957–5961.

(99) Hazra, S.; Deb, M.; Elias, A. J. Iodine catalyzed oxidation of alcohols and aldehydes to carboxylic acids in water: a metal-free route to the synthesis of furandicarboxylic acid and terephthalic acid. *Green Chem.* **2017**, *19* (23), 5548–5552.

(100) McKenna, S. M.; Leimkuhler, S.; Herter, S.; Turner, N. J.; Carnell, A. J. Enzyme cascade reactions: synthesis of furandicarboxylic acid (FDCA) and carboxylic acids using oxidases in tandem. *Green Chem.* **2015**, *17* (6), 3271–3275.

(101) McKenna, S. M.; Mines, P.; Law, P.; Kovacs-Schreiner, K.; Birmingham, W. R.; Turner, N. J.; Leimkuhler, S.; Carnell, A. J. The continuous oxidation of HMF to FDCA and the immobilisation and stabilisation of periplasmic aldehyde oxidase (PaoABC). *Green Chem.* **2017**, *19* (19), 4660–4665.

(102) Xu, S.; Zhou, P.; Zhang, Z.; Yang, C.; Zhang, B.; Deng, K.; Bottle, S.; Zhu, H. Selective Oxidation of 5-Hydroxymethylfurfural to 2,5-Furandicarboxylic Acid Using O_2 and a Photocatalyst of Cothioporphyrazine Bonded to g-C₃N₄. *J. Am. Chem. Soc.* **2017**, *139* (41), 14775–14782.

(103) You, B.; Jiang, N.; Liu, X.; Sun, Y. Simultaneous H_2 Generation and Biomass Upgrading in Water by an Efficient Noble-Metal-Free Bifunctional Electrocatalyst. *Angew. Chem., Int. Ed.* **2016**, *55* (34), 9913–9917.

(104) You, B.; Liu, X.; Liu, X.; Sun, Y. Efficient H_2 Evolution Coupled with Oxidative Refining of Alcohols via A Hierarchically Porous Nickel Bifunctional Electrocatalyst. *ACS Catal.* **2017**, 7 (7), 4564–4570.

(105) Nam, D.-H.; Taitt, B. J.; Choi, K.-S. Copper-Based Catalytic Anodes To Produce 2,5-Furandicarboxylic Acid, a Biomass-Derived Alternative to Terephthalic Acid. ACS Catal. 2018, 8 (2), 1197–1206. (106) Degtyar, E.; Harrington, M. J.; Politi, Y.; Fratzl, P. The Mechanical Role of Metal Ions in Biogenic Protein-Based Materials. Angew. Chem., Int. Ed. 2014, 53 (45), 12026–12044.

(107) Jensen, A. C. S.; Imberti, S.; Parker, S. F.; Schneck, E.; Politi, Y.; Fratzl, P.; Bertinetti, L.; Habraken, W. J. E. M. Hydrogen Bonding in Amorphous Calcium Carbonate and Molecular Reorientation Induced by Dehydration. *J. Phys. Chem. C* **2018**, *122* (6), 3591–3598.

(108) Fernandez, J. G.; Ingber, D. E. Manufacturing of Large-Scale Functional Objects Using Biodegradable Chitosan Bioplastic. *Macromol. Mater. Eng.* **2014**, *299* (8), *932–938*.

(109) Jayakumar, R.; Menon, D.; Manzoor, K.; Nair, S.; Tamura, H. Biomedical applications of chitin and chitosan based nanomaterials— A short review. *Carbohydr. Polym.* **2010**, *82* (2), 227–232.

(110) Jayakumar, R.; Prabaharan, M.; Nair, S. V.; Tamura, H. Novel chitin and chitosan nanofibers in biomedical applications. *Biotechnol. Adv.* **2010**, *28* (1), 142–150.

(111) Politi, Y.; Priewasser, M.; Pippel, E.; Zaslansky, P.; Hartmann, J.; Siegel, S.; Li, C.; Barth, F. G.; Fratzl, P. A spider's fang: How to design an injection needle using chitin-based composite material. *Adv. Funct. Mater.* **2012**, *22* (12), 2519–2528.

(112) Crini, G. Recent developments in polysaccharide-based materials used as adsorbents in wastewater treatment. *Prog. Polym. Sci.* 2005, 30 (1), 38–70.

(113) Bhatnagar, A.; Sillanpää, M. Applications of chitin-and chitosan-derivatives for the detoxification of water and wastewater a short review. *Adv. Colloid Interface Sci.* **2009**, *152* (1), 26–38.

(114) Yu, S.; Zang, H.; Chen, S.; Jiang, Y.; Yan, B.; Cheng, B. Efficient conversion of chitin biomass into 5-hydroxymethylfurfural over metal salts catalysts in dimethyl sulfoxide-water mixture under hydrothermal conditions. *Polym. Degrad. Stab.* **2016**, *134*, 105–114.

(115) Zhang, M.; Zang, H.; Ma, B.; Zhang, X.; Xie, R.; Cheng, B. Green Synthesis of 5-Hydroxymethylfurfural from Chitosan Biomass Catalyzed by Benzimidazole-Based Ionic Liquids. *ChemistrySelect* **2017**, *2* (32), 10323–10328.

(116) Jiang, Y.; Zang, H.; Han, S.; Yan, B.; Yu, S.; Cheng, B. Direct conversion of chitosan to 5-hydroxymethylfurfural in water using Brønsted–Lewis acidic ionic liquids as catalysts. *RSC Adv.* **2016**, 6 (105), 103774–103781.

(117) Zang, H.; Yu, S.; Yu, P.; Ding, H.; Du, Y.; Yang, Y.; Zhang, Y. Hydrothermal conversion of N-acetyl-d-glucosamine to 5-hydroxyme-thylfurfural using ionic liquid as a recycled catalyst in a water-dimethyl sulfoxide mixture. *Carbohydr. Res.* **2017**, *442*, 1–8.

(118) Gao, X.; Chen, X.; Zhang, J.; Guo, W.; Jin, F.; Yan, N. Transformation of Chitin and Waste Shrimp Shells into Acetic Acid and Pyrrole. ACS Sustainable Chem. Eng. 2016, 4 (7), 3912–3920.

(119) Zhang, J.; Yan, N. Formic acid-mediated liquefaction of chitin. Green Chem. 2016, 18 (18), 5050–5058.

(120) Bobbink, F. D.; Zhang, J.; Pierson, Y.; Chen, X.; Yan, N. Conversion of chitin derived N-acetyl-D-glucosamine (NAG) into polyols over transition metal catalysts and hydrogen in water. *Green Chem.* **2015**, *17* (2), 1024–1031.

(121) Einbu, A.; Vårum, K. M. Characterization of chitin and its hydrolysis to GlcNAc and GlcN. *Biomacromolecules* **2008**, 9 (7), 1870–1875.

(122) Yabushita, M.; Kobayashi, H.; Kuroki, K.; Ito, S.; Fukuoka, A. Catalytic Depolymerization of Chitin with Retention of N-Acetyl Group. *ChemSusChem* **2015**, *8* (22), 3760–3763.

(123) Zhang, J.; Yan, N. Production of Glucosamine from Chitin by Co-solvent Promoted Hydrolysis and Deacetylation. *ChemCatChem* **2017**, *9* (14), 2790–2796.

(124) Kobayashi, H.; Techikawara, K.; Fukuoka, A. Hydrolytic hydrogenation of chitin to amino sugar alcohol. *Green Chem.* **2017**, *19*, 3350–3356.

(125) Pierson, Y.; Chen, X.; Bobbink, F. D.; Zhang, J.; Yan, N. Acid-Catalyzed Chitin Liquefaction in Ethylene Glycol. *ACS Sustainable Chem. Eng.* **2014**, *2* (8), 2081–2089.

(126) Rose, M.; Palkovits, R. Isosorbide as a renewable platform chemical for versatile applications—Quo Vadis? *ChemSusChem* 2012, 5 (1), 167–176.

(127) Drover, M. W.; Omari, K. W.; Murphy, J. N.; Kerton, F. M. Formation of a renewable amide, 3-acetamido-5-acetylfuran, via direct conversion of N-acetyl-D-glucosamine. *RSC Adv.* **2012**, *2* (11), 4642–4644.

(128) Omari, K. W.; Dodot, L.; Kerton, F. M. A Simple One-Pot Dehydration Process to Convert N-acetyl-D-glucosamine into a Nitrogen-Containing Compound, 3-acetamido-5-acetylfuran. *ChemSuschem* **2012**, *5* (9), 1767–1772.

(129) Liu, Y.; Rowley, C. N.; Kerton, F. M. Combined Experimental and Computational Studies on the Physical and Chemical Properties of the Renewable Amide, 3-Acetamido-5-acetylfuran. *ChemPhysChem* **2014**, *15* (18), 4087–4094.

(130) Chen, X.; Chew, S. L.; Kerton, F. M.; Yan, N. Direct conversion of chitin into a N-containing furan derivative. *Green Chem.* **2014**, *16* (4), 2204–2212.

(131) Chen, X.; Liu, Y.; Kerton, F. M.; Yan, N. Conversion of chitin and N-acetyl-d-glucosamine into a N-containing furan derivative in ionic liquids. *RSC Adv.* **2015**, 5 (26), 20073–20080.

(132) Wolter, F. E.; Schneider, K.; Davies, B. P.; Socher, E. R.; Nicholson, G.; Seitz, O.; Süssmuth, R. D. Total Synthesis of Proximicin A–C and Synthesis of New Furan-Based DNA Binding Agents. *Org. Lett.* **2009**, *11* (13), 2804–2807. (133) Ikegami, S.; Isomura, H.; Tsuchimori, N.; Osano, Y. T.; Hayase, T.; Yugami, T.; Ohkishi, H.; Matsuzaki, T. Structure of pyrrolosine: a novel inhibitor of RNA synthesis, from the actinomycete Streptomyces albus. J. Am. Chem. Soc. **1990**, 112 (26), 9668–9669.

(134) Takahashi, Y.; Iinuma, Y.; Kubota, T.; Tsuda, M.; Sekiguchi, M.; Mikami, Y.; Fromont, J.; Kobayashi, J. i. Hyrtioseragamines A and B, New Alkaloids from the Sponge Hyrtios Species. *Org. Lett.* **2011**, *13* (4), 628–631.

(135) Brun, N.; Hesemann, P.; Esposito, D. Expanding the biomass derived chemical space. *Chem. Sci.* 2017, *8*, 4724–4738.

(136) Liu, Y.; Stähler, C.; Murphy, J. N.; Furlong, B. J.; Kerton, F. M. Formation of a Renewable Amine and an Alcohol via Transformations of 3-Acetamido-5-acetylfuran. *ACS Sustainable Chem. Eng.* **2017**, *5* (6), 4916–4922.

(137) Pham, T. T.; Chen, X.; Yan, N.; Sperry, J. A novel dihydrodifuropyridine scaffold derived from ketones and the chitinderived heterocycle 3-acetamido-5-acetylfuran. *Monatsh. Chem.* [Online Early Access]. DOI: 10.1007/s00706-017-2112-8. Published Online: December 6, 2017. https://link.springer.com/article/10. 1007%2Fs00706-017-2112-8 (accessed January 11, 2018).

(138) Brucoli, F.; Natoli, A.; Marimuthu, P.; Borrello, M. T.; Stapleton, P.; Gibbons, S.; Schätzlein, A. Efficient synthesis and biological evaluation of proximicins A, B and C. *Bioorg. Med. Chem.* **2012**, 20 (6), 2019–2024.

(139) Sadiq, A. D.; Chen, X.; Yan, N.; Sperry, J. Towards the Shell Biorefinery: Sustainable Synthesis of the Anticancer Alkaloid Proximicin A from Chitin. *ChemSusChem* **2018**, *11* (3), 532–535.

(140) Muñoz, I.; Rodríguez, C.; Gillet, D.; Moerschbacher, B. M. Life cycle assessment of chitosan production in India and Europe. *Int. J. Life Cycle Assess.* **2017**, 1–10.

(141) Chen, Q.; Xiao, W.; Zhou, L.; Wu, T.; Wu, Y. Hydrolysis of chitosan under microwave irradiation in ionic liquids promoted by sulfonic acid-functionalized ionic liquids. *Polym. Degrad. Stab.* **2012**, 97 (1), 49–53.

(142) Beaney, P.; Lizardi-Mendoza, J.; Healy, M. Comparison of chitins produced by chemical and bioprocessing methods. *J. Chem. Technol. Biotechnol.* **2005**, 80 (2), 145–150.

(143) Shahidi, F.; Synowiecki, J. Isolation and characterization of nutrients and value-added products from snow crab (Chionoecetes opilio) and shrimp (Pandalus borealis) processing discards. *J. Agric. Food Chem.* **1991**, 39 (8), 1527–1532.

(144) No, H. K.; Meyers, S. P.; Lee, K. S. Isolation and characterization of chitin from crawfish shell waste. *J. Agric. Food Chem.* **1989**, 37 (3), 575–579.

(145) Devi, R.; Dhamodharan, R. Pre-treatment in Hot Glycerol for Facile and Green Separation of Chitin from Prawn Shell Waste. *ACS Sustainable Chem. Eng.* **2018**, *6* (1), 846–853.

(146) Jia, X.; Ma, J.; Wang, M.; Ma, H.; Chen, C.; Xu, J. Catalytic conversion of 5-hydroxymethylfurfural into 2,5-furandiamidine dihydrochloride. *Green Chem.* **2016**, *18* (4), 974–978.

(147) Pelckmans, M.; Renders, T.; Van de Vyver, S.; Sels, B. F. Biobased amines through sustainable heterogeneous catalysis. *Green Chem.* **2017**, *19* (22), 5303–5331.

(148) Ye, X.; Plessow, P. N.; Brinks, M. K.; Schelwies, M.; Schaub, T.; Rominger, F.; Paciello, R.; Limbach, M.; Hofmann, P. Alcohol Amination with Ammonia Catalyzed by an Acridine-Based Ruthenium Pincer Complex: A Mechanistic Study. *J. Am. Chem. Soc.* **2014**, *136* (16), 5923–5929.

(149) Pingen, D.; Lutz, M.; Vogt, D. Mechanistic Study on the Ruthenium-Catalyzed Direct Amination of Alcohols. *Organometallics* **2014**, 33 (7), 1623–1629.

(150) Storer, R. I.; Carrera, D. E.; Ni, Y.; MacMillan, D. W. C. Enantioselective organocatalytic reductive amination. *J. Am. Chem. Soc.* **2006**, *128* (1), 84–86.

(151) Abdel-Magid, A. F.; Carson, K. G.; Harris, B. D.; Maryanoff, C. A.; Shah, R. D. Reductive amination of aldehydes and ketones with sodium triacetoxyborohydride. studies on direct and indirect reductive amination procedures. *J. Org. Chem.* **1996**, *61* (11), 3849–3862.

(152) Liang, G.; Wang, A.; Li, L.; Xu, G.; Yan, N.; Zhang, T. Production of Primary Amines by Reductive Amination of Biomass-Derived Aldehydes/Ketones. *Angew. Chem.* **2017**, *129* (11), 3096– 3100.

(153) Pelckmans, M.; Vermandel, W.; Van Waes, F.; Moonen, K.; Sels, B. F. Low-Temperature Reductive Aminolysis of Carbohydrates to Diamines and Aminoalcohols by Heterogeneous Catalysis. *Angew. Chem.* **2017**, *129* (46), 14732–14736.

(154) Poissonnier, J.; Pelckmans, M.; Van Waes, F.; Moonen, K.; Sels, B. F.; Thybaut, J. W.; Marin, G. B. Kinetics of Homogeneous and Heterogeneous Reactions in the Reductive Aminolysis of Glucose with Dimethylamine. *Appl. Catal., B* **2018**, *227*, 161–169.

(155) Niemeier, J.; Engel, R. V.; Rose, M. Is water a suitable solvent for the catalytic amination of alcohols? *Green Chem.* **2017**, *19* (12), 2839–2845.

(156) Karp, E. M.; Eaton, T. R.; Sànchez i Nogué, V.; Vorotnikov, V.; Biddy, M. J.; Tan, E. C. D.; Brandner, D. G.; Cywar, R. M.; Liu, R.; Manker, L. P.; Michener, W. E.; Gilhespy, M.; Skoufa, Z.; Watson, M. J.; Fruchey, O. S.; Vardon, D. R.; Gill, R. T.; Bratis, A. D.; Beckham, G. T. Renewable acrylonitrile production. *Science* **2017**, *358* (6368), 1307–1310.

(157) Chen, X.; Yang, H.; Hülsey, M. J.; Yan, N. One-Step Synthesis of N-Heterocyclic Compounds from Carbohydrates over Tungsten-Based Catalysts. *ACS Sustainable Chem. Eng.* **2017**, *5* (11), 11096–11104.

(158) Song, L.; Zheng, M.; Pang, J.; Sebastian, J.; Wang, W.; Qu, M.; Zhao, J.; Wang, X.; Zhang, T. One-pot synthesis of 2-hydroxymethyl-5-methylpyrazine from renewable 1, 3-dihydroxyacetone. *Green Chem.* **2017**, *19*, 3515–3519.

(159) Mycroft, Z.; Gomis, M.; Mines, P.; Law, P.; Bugg, T. D. H. Biocatalytic conversion of lignin to aromatic dicarboxylic acids in Rhodococcus jostii RHA1 by re-routing aromatic degradation pathways. *Green Chem.* **2015**, *17* (11), 4974–4979.

(160) Jiao, G.; Yu, G.; Zhang, J.; Ewart, H. S. Chemical Structures and Bioactivities of Sulfated Polysaccharides from Marine Algae. *Mar. Drugs* **2011**, 9 (2), 196–223.

(161) Holdt, S. L.; Kraan, S. Bioactive compounds in seaweed: functional food applications and legislation. *J. Appl. Phycol.* 2011, 23 (3), 543–597.

(162) Ong, B. S.; Wu, Y.; Liu, P.; Gardner, S. High-performance semiconducting polythiophenes for organic thin-film transistors. *J. Am. Chem. Soc.* **2004**, *126* (11), 3378–3379.

(163) Foley, P.; Kermanshahi pour, A.; Beach, E. S.; Zimmerman, J.
B. Derivation and synthesis of renewable surfactants. *Chem. Soc. Rev.* 2012, 41 (4), 1499–1518.

(164) Feng, M.; Tang, B.; Liang, S. H.; Jiang, X. Sulfur containing scaffolds in drugs: synthesis and application in medicinal chemistry. *Curr. Top. Med. Chem.* **2016**, *16* (11), 1200–1216.

(165) Firdaus, M.; Montero de Espinosa, L.; Meier, M. A. R. Terpene-based renewable monomers and polymers via thiol-ene additions. *Macromolecules* **2011**, *44* (18), 7253–7262.

(166) Yao, K.; Tang, C. Controlled polymerization of nextgeneration renewable monomers and beyond. *Macromolecules* **2013**, 46 (5), 1689–1712.

(167) Lowe, A. B. Thiol-ene "click" reactions and recent applications in polymer and materials synthesis: a first update. *Polym. Chem.* **2014**, 5 (17), 4820–4870.

(168) Worthington, M. J. H.; Kucera, R. L.; Albuquerque, I. S.; Gibson, C. T.; Sibley, A.; Slattery, A. D.; Campbell, J. A.; Alboaiji, S. F. K.; Muller, K. A.; Young, J.; Adamson, N.; Gascooke, J. R.; Jampaiah, D.; Sabri, Y. M.; Bhargava, S. K.; Ippolito, S. J.; Lewis, D. A.; Quinton, J. S.; Ellis, A. V.; Johs, A.; Bernardes, G. J. L.; Chalker, J. M. Laying Waste to Mercury: Inexpensive Sorbents Made from Sulfur and Recycled Cooking Oils. *Chem. - Eur. J.* **2017**, *23* (64), 16219–16230.

(169) Bailey, S. E.; Olin, T. J.; Bricka, R. M.; Adrian, D. D. A review of potentially low-cost sorbents for heavy metals. *Water Res.* **1999**, 33 (11), 2469–2479.

(170) Bantchev, G. B.; Kenar, J. A.; Biresaw, G.; Han, M. G. Free Radical Addition of Butanethiol to Vegetable Oil Double Bonds. *J. Agric. Food Chem.* **2009**, *57* (4), 1282–1290.

(171) Lowe, A. B. Thiol-ene "click" reactions and recent applications in polymer and materials synthesis. *Polym. Chem.* **2010**, *1* (1), 17–36. (172) Park, D. S.; Joseph, K. E.; Koehle, M.; Krumm, C.; Ren, L.; Damen, J. N.; Shete, M. H.; Lee, H. S.; Zuo, X.; Lee, B.; Fan, W.; Vlachos, D. G.; Lobo, R. F.; Tsapatsis, M.; Dauenhauer, P. J. Tunable Oleo-Furan Surfactants by Acylation of Renewable Furans. *ACS Cent. Sci.* **2016**, 2 (11), 820–824.

(173) Changi, S.; Matzger, A. J.; Savage, P. E. Kinetics and pathways for an algal phospholipid (1, 2-dioleoyl-*sn*-glycero-3-phosphocholine) in high-temperature (175–350 C) water. *Green Chem.* **2012**, *14* (10), 2856–2867.

(174) Kensil, C. R.; Dennis, E. A. Alkaline hydrolysis of phospholipids in model membranes and the dependence on their state of aggregation. *Biochemistry* **1981**, *20* (21), 6079–6085.

(175) Zuidam, N.; Crommelin, D. Chemical hydrolysis of phospholipids. J. Pharm. Sci. 1995, 84 (9), 1113–1119.

(176) Song, X.; Seidler, F.; Saleh, J.; Zhang, J.; Padilla, S.; Slotkin, T. Cellular mechanisms for developmental toxicity of chlorpyrifos: targeting the adenylyl cyclase signaling cascade. *Toxicol. Appl. Pharmacol.* **1997**, *145* (1), 158–174.

(177) Steinrücken, H. C.; Amrhein, N. The herbicide glyphosate is a potent inhibitor of 5-enolpyruvylshikimic acid-3-phosphate synthase. *Biochem. Biophys. Res. Commun.* **1980**, *94* (4), 1207–1212.

(178) Giannousis, P. P.; Bartlett, P. A. Phosphorus amino acid analogs as inhibitors of leucine aminopeptidase. *J. Med. Chem.* **1987**, 30 (9), 1603–1609.

(179) Harnisch, F.; Urban, C. Electrobiorefineries: Unlocking the Synergy of Electrochemical and Microbial Conversions. *Angew. Chem., Int. Ed.* [Online Early Access]. DOI: 10.1002/anie.201711727. Published Online: February 28, 2018. http://onlinelibrary.wiley. com/doi/10.1002/anie.201711727/abstract (accessed March 05, 2018).

(180) Da Vià, L.; Recchi, C.; Gonzalez-Yañez, E. O.; Davies, T. E.; Lopez-Sanchez, J. A. Visible light selective photocatalytic conversion of glucose by TiO₂. *Appl. Catal., B* **2017**, *202*, 281–288.

(181) Alonso, D. M.; Hakim, S. H.; Zhou, S.; Won, W.; Hosseinaei, O.; Tao, J.; Garcia-Negron, V.; Motagamwala, A. H.; Mellmer, M. A.; Huang, K.; Houtman, C. J.; Labbé, N.; Harper, D. P.; Maravelias, C. T.; Runge, T.; Dumesic, J. A. Increasing the revenue from lignocellulosic biomass: Maximizing feedstock utilization. *Sci. Adv.* **2017**, *3* (5), No. e1603301.

(182) Fiorentino, G.; Ripa, M.; Ulgiati, S. Chemicals from biomass: technological versus environmental feasibility. A review. *Biofuels, Bioprod. Biorefin.* **2017**, *11* (1), 195–214.

(183) Lari, G. M.; Pastore, G.; Haus, M.; Ding, Y.; Papadokonstantakis, S.; Mondelli, C.; Perez-Ramirez, J. Environmental and economical perspectives of a glycerol biorefinery. *Energy Environ. Sci.* [Online Early Access]. DOI: 10.1039/C7EE03116E. Published Online: February 26, 2018. http://pubs.rsc.org/en/content/ articlelanding/2018/ee/c7ee03116e#!divAbstract (accessed March 05, 2018).